

Biomedical Communication

Bupleurum turcicum: A Rich Source of Saikosaponin A and D with Potential Use as Adjunct for the Management of Acute Respiratory Diseases: A Meta Review

Hani A. Alfheeaid,,^{1,2} Bassam I. Alkhalifah,³ Abdulrahman A. Alsayegh⁴ Md Faruque Ahmad,⁴ Mohammed Idreesh Khan⁵ and Faiyaz Ahmed⁵

 ¹Department of Food Science and Human Nutrition, College of Agriculture and Veterinary Medicine, Qassim University, Buraydah, Saudi Arabia
 ²Human Nutrition, School of Medicine, College of Medical, Veterinary and Life Sciences, University of Glasgow, New Lister Building, Glasgow Royal Infirmary, Glasgow, United Kingdom
 ³Department of Radiology College of Medicine and Medical Sciences, Qassim University, Unaizah, Saudi Arabia
 ⁴Department of Clinical Nutrition, College of Applied Medical Sciences, Jazan University, Jazan, Saudi Arabia
 ⁵Department of Clinical Nutrition, College of Applied Health Sciences in Ar Rass, Qassim University, Ar Rass, Saudi Arabia

ABSTRACT

COVID-19 pandemic caused by SARS-CoV-2 has resulted in unprecedented havoc worldwide with significant morbidity and mortality. Till now, no effective antivirals are at disposal prompting researchers to explore potential lead molecules including from bioactive phytochemicals. An extensive literature search was carried out utilizing online resources; Google Scholar and PubMed to collect published reports on pharmacological potential of saikosaponin particularly in underexplored *Bupleurum* species. A number of molecular docking studies have reported promising antiviral effects of saikosaponins particularly of saikosaponin A, D, U and V with tremendous potential to be developed as anti-SARS-CoV-2 therapy. The search for potential sources of saikosaponin A and D led to the identification of *Bupleurum turcicum*; an unexplored, underutilized and endemic *Bupleurum* species. The observation that *B. turcicum* root extract contains highest amount of SSa and SSd among endemic Bupleurum pauciradiatum) and presence of significant amounts of antioxidant compounds led to the proposition of using *B. turcicum* extracts as adjust therapy in the management of COVID-19. The proposal also relies on the evidence of SSa and SSd being effective against a number of viruses including SARS-CoV. This review discusses phytochemical composition of *B. turcicum* root, antiviral, immunomodulatory and anti-inflammatory potential of saikosaponins in view of its plausible usefulness in the management of COVID-19. *B. turcicum* is an underutilized species rich in saikosaponin A and D with potential antiviral properties which could be effective alternative therapy in COVID-19 management.

KEY WORDS: ANTIVIRAL, BUPLEURUM TURCICUM, COVID-19, SAIKOSAPONINS, SARS-COV.

INTRODUCTION

SARS-CoV-2 (COVID-19) infection started as an outbreak of pneumonia of unknown origin in Wuhan City, China during late December 2019 and soon declared as a global epidemic by February 2020 (Lu et al. 2020; WHO 2020).

Article Information:*Corresponding Author: f.masfoor@qu.edu.sa Received 25/10/2021 Accepted after revision 18/12/2021 Published: 31st December 2021 Pp- 1444-1451 This is an open access article under Creative Commons License, Published by Society for Science & Nature, Bhopal India. Available at: https://bbrc.in/ DOI: http://dx.doi.org/10.21786/bbrc/14.4.11 Soon the sequencing of virus genome from patient samples and human to human transmission were confirmed by researchers (Zhu et al. 2020; Chan et al. 2020). Researchers began to explore novel therapeutic approaches to treat COVID-19 in view of its potential to rapidly develop into acute respiratory distress syndrome which can in some cases possibly cause multiple organ failure in the absence of effective drugs/vaccines (Patel and Jernigan 2020; Sahin et al. 2020; Gralinski and Menachery 2020).



It is reported that during the outbreak of SARS-CoV in 2003, traditional herbal medicines used in conjunction with conventional medicine were effective in alleviating the symptoms of SARS (Lin et al. 2003; Xiao et al. 2003; Zhao et al. 2003; Zhong et al. 2003). Though the mechanism of action of those herbal products was not fully understood, the general consensus was that they contain antiviral molecules which might be acting by inhibiting viral replication (Vlietinck et al. 1991; McCutcheon et al. 1995; Jassim and Naji 2003). Therefore, exploring natural products with demonstrated antiviral efficacy against other coronaviruses is one of the therapeutic approaches. Since SARS-CoV-2 is a betacoronavirus which invades the host cell through angiotensin converting enzyme 2 (ACE2) receptor similar to SARS-CoV, it is assumed that antiviral biomolecules that are active against SARS-CoV can be repurposed to be used in COVID-19 patients (Cheng et al. 2006; Huanga et al. 2020; Shahrajabian et al. 2020).

In view of this, an extensive literature search was carried out utilizing online resources; Google Scholar and PubMed to collect published reports on herbal medicines and natural products with antiviral activity. The search was then tapered down to the natural compounds active against SARS-CoV wherein, saikosaponins widely distributed in *Bupleurum* spp., Heteromorpha spp. and Scrophularia spp were selected to explore further as saikosaponin A, B2, C and D have been reported to show significant antiviral activity against human coronavirus 229E in vitro (Cheng et al. 2006). The search was further narrowed down to *Bupleurum* spp. and Bupleurum turcicum was selected for this study since it contained highest levels of saikosaponin A and saikosaponin D amongst Bupleurum sulphureum, Bupleurum lycaonicum, Bupleurum heldreichii, and Bupleurum pauciradiatum (Kars et al. 2012; Huanga et al. 2020).

Bupleurum L. genus belongs to Apiaceae family which consists of around 200 species distributed across Northern Hemisphere, Eurasia, and North Africa. The plants are often annual or perennial herbs/shrubs up to 2 m tall with simple, entire and alternate leaves and the flowers are small, radical and yellowish (Xie et al. 2009). Many plants particularly roots from *Bupleurum* genus are used in traditional systems of medicine across China, Japan, Korea and Taiwan in the treatment of fevers associated with common cold and malaria, inflammation, hepatitis, diabetes, cancer and for wound healing, while the essential oils are used as anti-inflammatory and antiseptic agents (Nose et al. 1989; Motoo et al. 1994; Benito et al. 1998; Van-Wyk and Wink 2004; Wu 2005). Bupleurum species are used as analgesics in amenorrhea and cholecystitis. They also find their use in nephrotic syndrome, autoimmune diseases deafness, dizziness, dry throat, vomiting, diarrhea and hemorrhoids (Ashour and Wink 2011). These plants reportedly contain diverse classes of bioactive components including essential oils, alkaloids, flavonoids, coumarins, polysaccharides, lignans, triterpene saponins, phytosterols and polyacetylenes. WHO monographs list Bupleurum species as commonly used medicinal plants of China and Korea and also officially listed in Japanese and Chinese Pharmacopoeias (Ashour and Wink 2011; Huanga et al. 2020).

Bupleurum Bioactive Saikosaponins: Saikosaponins are triterpene oleanane saponin glycosides widely distributed bioactive compounds in *Bupleurum* spp amounting to up to 7% of the total dry weight in roots. Though saikosaponins are classified into seven types based on the type of aglycone (closely related oxygenated pentacyclic triterpenoidal structures), four different types namely saikosaponin-A (SSa), saikosaponin-B (SSb), saikosaponin-C (SSc) and saikosaponin-D (SSd) are considered as most commonly occurring and biologically active saikosaponins (Yuan et al. 2017). SSa, SSd and SSc are epoxy-ether saikosaponins designated as type I and SSb2 is a heterocyclic diene saikosaponin designated as type II (Lin et al. 2013; Huanga et al. 2020).

Plant extracts/herbal medicines containing saikosaponins are commonly prescribed as anti-inflammatory and antiinfectious medicine in Asian countries including China, Taiwan and Japan. More than 120 types of saikosaponins have been isolated from *Bupleurum* species till date and some of them have significant bioactivity both *in vitro* and in vivo (Kim 2018). Previous studies have reported antioxidant, antidepressant, anti-inflammatory, antimicrobial, antiviral, anticancer, anti-tumor and immunomodulatory effects of saikosaponins (Wu et al. 2008; Wu et al. 2010; Jin et al. 2013; Sui et al. 2011; Wu et al. 2011; Ying et al. 2014; Zhang et al. 2014; Huanga et al. 2020).

Bupleurum turcicum: B. turcicum belongs to Apiaceae family which mostly has flower bearing shrubs (Davis 1972). The essential oils from flowers, fruits and roots contain 39 distinctive compounds (Table 1). Heptanal, pentadecane and undecane were major compounds of flower and fruit essential oils. The flowers contained 33.2% heptanal, 19.6% pentadecane, 6.6% undecane, while fruits contained 23.5% heptanal, 13.4% pentadecane and 8.9% undecane. On an interesting note, essential oil from root did not contain heptanal, pentadecane or undecane which were major compounds of flowers and fruits. The major compounds of root essential oil were pentacosane (9%), 1-undecanol (8.8%) and hexacosane (8.0%) (Huanga et al. 2020).

The oils were evaluated for their antibacterial activity against Staphylococcus aureus, Staphylococcus aureus, Escherichia coli, Escherichia coli, Bacillus cereus, Streptococcus salivarius, Pseudomonas aeruginosa, Pseudomonas aeruginosa and Proteus mirabilis. It was found that the essential oils of flowers and roots did not show any activity against tested bacteria, while the oil obtained from roots exhibited good antibacterial activity which was comparable to that of chloramphenicol against E. coli, B. cereus, S. salivarius and *P. aeruginosa*. The observed phenomenon is attributable their chemical composition wherein, both fruits and flowers contained heptanal, pentadecane or undecane as major components which were not found in root samples. It was concluded that the essential oils from the roots of B. turcicum could be used as a potential source of novel antibacterial agents (Cheng et al. 2006; Kars et al. 2012; Saraçoğlu et al. 2012; Tykheev et al. 2020).

Table 1. Photochemical composition of <i>Bupleurum turcicum</i> essential oils						
Flowers (%)	Fruits (%)	Roots (%)	Compounds	Flowers (%)	Fruits (%)	Roots (%)
0.4	0.5	0.9	Dodecanoic acid	0.4	0.8	
1.5	2.3	-	Farnesyl acetone	0.1	0.4	1.4
0.5	0.2	-	Heneicosane	-	-	1.1
-	-	0.8	Heptacosane	0.3		3.6
3.1	7.7	0.8	Heptadecane	0.8	0.8	0.6
-	0.5	-	Heptanal	33.2	23.5	-
0.8	0.8		Heptanoic acid	1.0	0.7	-
0.8	0.7	0.8	Hexacosane	0.2	-	8.0
1.4	2.5	-	Hexadecane	0.2	0.2	0.4
-	-	0.8	Hexadecanoic acid	1.9	2.8	0.2
-	-	6.3	Hexahydrofarnesyl acetone	1.4	3.6	3.7
0.5	0.5	-	Hexanal	0.9	0.5	-
0.3	0.7	-	Limonene	1.5	1.8	-
0.3	0.7	-	Nonacosane	0.2	-	2.4
0.2	-	-	Nonadecane	-	-	0.5
-	-	2.2	Nonanal	0.3	0.4	0.2
-	-	8.8	Octacosane	0.1	-	3.9
-	-	4.5	Octadecane	0.2	-	0.5
0.3	0.3	-	Octanal	0.5	0.5	-
0.3	-	-	Pentacosane	0.2	-	9.0
1.3	1.0	-	Pentadecane	19.6	13.4	3.0
0.3	0.7	0.5	Phytol	-	1.0	-
0.4	-	-	Spathulenol	3.5	5.9	6.3
0.2	-	-	Tetracosane	0.2	-	5.4
-	0.3	-	Tetradecane	0.3	-	-
-	0.2	-	Tetradecane	-	0.2	0.5
-	-	1.8	Tetradecanoic acid	0.6	2.6	-
-	-	0.2	Tricosane	-	-	3.3
3.5	2.4	-	Tridecane	1.8	1.9	1.8
-	-	0.5	Undecanal	-	-	0.5
-	-	0.4	Undecane	0.6	8.9	5.5
-	-	0.1	α-Pinene	0.8	2.0	-
-	0.2	-	β-Caryophyllene	0.5	-	-
-	-	1.4	β-Elemene	0.2	0.8	-
-	-	0.4	γ-Muurolene	1.6	-	-
	sition of <i>E</i> Flowers (%) 0.4 1.5 0.5 3.1 - 0.8 0.8 1.4 0.5 0.3 0.3 0.2 0.3 0.3 0.2 0.3 0.3 0.3 0.2 0.3 0.3 0.2 1.4 0.2 0.2 0.3 0.3 0.3 0.4 0.2 0.2 0.3 0.3 0.3 0.4 0.2 0.2 0.2 0.2 0 0.1 0 0.2 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	Bition of Bupleurun Flowers Fruits (%) 0.4 0.5 1.5 2.3 0.5 0.2 - - 3.1 7.7 - 0.5 0.8 0.8 0.8 0.7 1.4 2.5 - - 0.5 0.5 0.8 0.7 1.4 2.5 - - 0.5 0.5 0.3 0.7 0.2 - - - 0.3 0.7 0.3 0.7 0.3 0.7 0.3 0.7 0.3 0.7 0.3 0.7 0.4 - 1.3 1.0 0.3 0.7 0.4 - 0.2 - - - 0.2 - - - 0.2 - - -	sition of Bupleurum turcicul Flowers Fruits Roots (%) (%) (%) 0.4 0.5 0.9 1.5 2.3 - 0.5 0.2 - 0.5 0.2 - 0.5 0.2 - 0.5 0.2 - 0.5 0.2 - 0.5 0.2 - 0.5 0.2 - 0.8 0.8 - 0.8 0.8 - 0.8 0.7 0.8 1.4 2.5 - - 0.8 - 0.8 0.7 0.8 0.5 0.5 - 0.3 0.7 - 0.3 0.7 - 0.3 0.7 - 0.3 0.7 - 0.3 0.7 0.5 0.4 - - 0.3 0.7 0.5 0.4 - - <td< td=""><td>sition of between sevential oilsFlowers (%)Fruits (%)Roots (%)Compounds0.40.50.9Dodecanoic acid1.52.3-Farnesyl acetone0.50.2-Heneicosane0.50.2-Heneicosane1.17.70.8Heptanceane3.17.70.8Heptanecane0.50.5-Heptanoic acid0.80.8Heytacosane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.50.5-Hexanal0.80.70.8Hexanal0.30.7-Nonacosane0.42.2Nonanal1.52.2Nonanal1.6-2.2Nonane1.31.0-Pentacosane1.31.0-Pentacosane1.31.0-Pentacosane1.31.0-Tetradecane0.30.70.5Phytol0.4Tetradecanoic acid1.31.0</td><td>sition of between between</td><td>sition of Burlew burle</td></td<>	sition of between sevential oilsFlowers (%)Fruits (%)Roots (%)Compounds0.40.50.9Dodecanoic acid1.52.3-Farnesyl acetone0.50.2-Heneicosane0.50.2-Heneicosane1.17.70.8Heptanceane3.17.70.8Heptanecane0.50.5-Heptanoic acid0.80.8Heytacosane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.42.5-Hexadecane1.50.5-Hexanal0.80.70.8Hexanal0.30.7-Nonacosane0.42.2Nonanal1.52.2Nonanal1.6-2.2Nonane1.31.0-Pentacosane1.31.0-Pentacosane1.31.0-Pentacosane1.31.0-Tetradecane0.30.70.5Phytol0.4Tetradecanoic acid1.31.0	sition of between	sition of Burlew burle

*Source: (Cheng et al. 2006; Kars et al. 2012; Saraçoğlu et al. 2012; Tykheev et al. 2020)

In an important study, the levels of phenolic compounds (catechin, quercetin, isoquercitrin), SSa, SSd and podohyllotoxin were determined in the root extracts of *B. sulphureum, B. lycaonicum, B. turcicum, B. heldreichii* and *B. pauciradiatum* using HPLC. *B. turcicum* was found to contain significant amounts of total phenolics (34.48 mg GAE/g extract), catechin (0.11 mg/g extract), quercetin (0.21 mg/g extract) and isoquercitrin (1.85 mg/g extract).

Furthermore, *B. turcicum* root extract contained highest amounts of SSa (12.99 mg/g extract) and SSd (17.96 mg/g extract) among all the other extracts investigated (Huanga et al. 2020; Shahrajabian et al. 2020).

B. turcicum root extract exhibited significant free radical scavenging activity with lowest IC50 value (57.3 μ g/mL) amongst other extracts in DPPH radical scavenging assay.

In antiproliferative assay, *B. turcicum* root extract exhibited potent activity against sensitive and drug resistant MCF-7 cells (Kars et al. 2012). The observation that *B. turcicum* root extract contains highest amount of SSa and SSd among endemic *Bupleurum* species also found in Turkey (*B. sulphureum*, *B. lycaonicum*, *B. turcicum*, *B. heldreichii*, *B. pauciradiatum*) and presence of significant amounts of antioxidant compounds led to the proposition of using *B. turcicum* extracts as adjust therapy in the management of COVID-19. The proposal also relies on the evidence of SSa and SSd being effective against a number of viruses including SARS-CoV (Huanga et al. 2020; Shahrajabian et al. 2020).

Antiviral properties of saikosaponins: Recent studies have evaluated the anti-SARS-CoV-2 effects of different saikosaponins found in *Bupleurum* Spp. using advanced molecular docking techniques and have found promising results with potential to be studied further with respect to saikosaponins A, D, U and V. Similarly, in the experimental setup saikosaponins isolated from Bupleurum spp. have been shown to possess potent antiviral activity against a number of viruses including herpes simplex virus, influenza virus (IAV), hepatitis B virus (HBV), hepatitis C virus (HCV), measles and varicella zoster viruses (Ashour and Wink 2011). Based on these findings, saikosaponins from *Bupleurum* Spp. have been proposed to be repurposed for the treatment of COVID-19 (Bahbah et al. 2020).

In a molecular docking study, SSa has been shown to possess significant affinity towards binding ACE 2 receptors through which SARS-CoV-2 infect lung epithelial cells (Yan et al. 2020). Another molecular docking simulation study showed that Saikosaponins exhibit high affinity towards RBD region of the spike glycoprotein of SARS-CoV (Goswami and Bagchi 2020). Another molecular docking study conducted to evaluate the affinity of Saikosaponins towards SARS-CoV-2 binding protein showed, Saikosaponins U and V to exhibit strong affinity towards SARS-CoV-2 binding protein. The study concluded that Saikosaponins U and V could be future research molecules for SARS-CoV-2 research (Sinha et al. 2020).

A study conducted to evaluate the antiviral and immunoregulatory activities of SSa and SSd against Porcine Coronavirus 2 (PCV2) showed that saikosaponins reduced the incidence and severity of PCV2-induced immunopathological damage in terms of pyrexia, weight loss, anemia, and internal organ oedema in mice. Immunoglobulin and protein absorption levels were also affected suggesting immunoregulatory effect of saponins (Yang et al. 2017). In another study, antiviral activity of saikosaponins against herpes simplex type I (HSV-1), vesicular stomatitis virus (VSV) and poliovirus type 1 was evaluated *in vitro* at non-cytotoxic concentrations. Buddlejasaponin 4 was found to be potent against vesicular stomatitis virus (Bermejo et al. 2002; Shahrajabian et al. 2020).

Ushio and Abe evaluated the virus neutralizing effect of SSd against measles virus and herpes simplex virus *in vitro*

(Ushio and Abe 1992). SSd at above 5 μ M concentration rendered measles virus and herpes simplex virus (HSV) completely non-infective when incubated together for 10 min at room temperature. Lin et al (2015) evaluated the antiviral activity of SSa, SSb₂, SSc, and SSd against cultured hepatitis C virus (HCV) *in vitro*. The effect of saikosaponins on virus entry, RNA replication/translation and particle production were studied using different HCV genotypes, clinical isolates and infection of primary human hepatocytes. All saikosaponins were found to exhibit potent inhibition of HCV infections even at non-cytotoxic concentrations by targeting early steps of the viral life cycle. SSb₂ was found to significantly prevent virus entry by neutralizing virus particles, preventing their attachment and inhibiting viral entry/fusion.

SSb₂ also inhibited other genotypic strains and prevented HCV binding onto hepatoma cells thereby blocking HCV infection of primary human hepatocytes. The study proposed further research into SSb₂ to develop it as an HCV entry antagonist. These finding were reiterated by another study in 2019, wherein SSb2 was found to inhibit viral entry, replication, and translation of hepatitis C virus (HCV) in a cell culture-derived HCV system *in vitro*. SSb₂ also inhibited daclatasvir-resistant mutant strains of HCV when used in combination with daclatasvir indicating potential antiviral effects of SSb₂. Antiviral activity of SSa, SSc and SSd against anti-hepatitis B virus (HBV) was evaluated *in vitro* using HBV-transfected human hepatoma cells (Lee et al. 2019; Shahrajabian et al. 2020).

SSc was found to inhibit DNA replication of HBV which was higher than that of lamivudine- a known antiviral drug in clinical use (Chiang et al. 2003; Chang et al. 2007). In another study SSd showed significant antiviral activity against HBV through inhibition of HBV-DNA replication (Yin et al. 2008). The anti-viral activity of SSa was evaluated against influenza A virus (IAV) infections in vitro and in vivo. SSa reduced replication of three different influenza A virus strains, including a H5N1 strain, in human alveolar epithelial A549 cells by downregulating NF-kB signaling and caspase 3-dependent virus ribonucleoprotein nuclear export. SSa decreased viral replication, production of proinflammatory cytokines in H1N1 PR8 model of influenza A virus lethality in C57BL/6 mice. SSa also attenuated lung neutrophil and monocyte recruitment during the early stages of immune response to PR8 infection (Chen et al. 2015; Shahrajabian et al. 2020).

The antiviral activities of saikosaponins against human coronavirus (HCoV), which cause severe acute respiratory syndrome (SARS), were also studied using 2,3-bis[2methoxy-4-nitro-5-sulfophenyl]-5-[(phenylamino) carbonyl-2H-tetrazolium hydroxide] (XTT) assay *in vitro*. Results indicated both SSa and SSb2 showed significant antiviral activity against HCoV at concentrations of 0.25-25 mmol/L. Both saikosaponins exhibited no cytotoxic effects on target cells at tested concentrations. The antiviral activity of SSb2 was more significant than that of SSa and mediated by inhibition of attachment and penetration of the virus to target cells (Cheng et al. 2006; Shahrajabian et al. 2020). **Immunomodulatory and anti-inflammatory activity of saikosaponins in view of** COVID-19: SARS-CoV-2 much like Influenza A virus (IAV) can cause severe pneumonia resulting in morbidities and mortalities (Chen et al. 2015). Generally, infection starts in the upper respiratory tract epithelial cells and spreads aggressively into deeper regions of the lung parenchyma and might also enter into macrophages and dendritic cells (Spiegel et al. 2006; Manicassamy et al. 2010; Shahrajabian et al. 2020).

The infected cells release pro-inflammatory cytokines (IL2, IL7, IL10, IP10 and TNFa) and chemokines (MIP-1a and MCP1). Although these responses are important for controlling viral replication in the initial phase of infection through recruitment of immune cell into lungs, excessive pro-inflammatory components raise the levels of cytotoxic and pro-apoptotic products which damage lung tissue (Herold et al. 2008). Thus, natural products with anti-inflammatory and immunomodulatory properties could be useful in the management of COVID-19 (Huanga et al. 2020). Kumazawa and coworkers investigated macrophage activation potential of SSa and SSd in mice. Intraperitoneal injection of saikosaponins induced a dose dependent activation of peritoneal macrophages leading to enhanced phagocytic activity, increased cellular lysosomal levels, induction of cytostatic activity and expression of Ia antigen on the cell surface. Authors opined that SSd could be a potent macrophage activator as it showed significantly higher activity compared to ginsenoside Rg1 and glycyrrhizin. Furthermore, SSd also modulates lymphocyte activity through suppression of T-cell and induction of B-cell response to different mitogens and up-regulates IL-2, IL-4 production in thymocytes through post-receptor signal transduction (Kumazawa et al. 1989; Ushio and Abe 1991; Kato et al. 1995; Lin et al. 2015; Huanga et al. 2020).

Wong and coworkers reported *in vitro* suppression of OKT3/ CD28-costimulated human T cell proliferation and inhibition of PMA, PMA/Ionomycin and Con A-induced mouse T cell activation by saikosaponin D *in vitro*. Examination of T cell activation signaling pathways suggested that saikosaponin inhibits T cell activation downregulating CD69, CD71 expression and IL-2 production through modulation of PKC pathway via PKCh, JNK, and NF-kB transcription factors. It also downregulates CD25, IL-6, TNF α and IFN γ through NF-kB, NF-AT and AP-1 (c-Fos) signaling pathways (Wong et al. 2009). These observations indicate that SSd could be a potential molecule with immunomodulatory functions (Leung et al. 2005; Huanga et al. 2020).

SSa was found to inhibit inflammatory factors such as cyclooxygenase-2 (COX-2) and inducible nitric-oxide synthase (iNOS) and production of pro-inflammatory cytokines TNF α , IL1 β and IL6 in an experimental model of inflammation (lipopolysaccharide (LPS)-stimulated RAW 264.7 cells). The study concluded that SSa exhibits significant anti-inflammatory activity through regulation of inflammatory mediators and suppression of MAPK and NF- κ B signaling pathways (Zhu et al. 2013). These finding were reiterated in another study, wherein SSa significantly inhibited COX-2, iNOS, tumor necrosis factor- α , IL1 β and

IL6 in an obesity experimental model using mouse embryo fibroblast 3T3-L1 cells. The study also suggested SSa to be a potential therapeutic agent against obesity associated inflammation (Kim et al. 2015). SSa and SSd are reported to exhibit potent anti-inflammatory activity by inhibiting production of nitric oxide induced by lipopolysaccharide in BV-2 microglial cells. The inhibitory effect of both SSa and SSd were comparable to that of dexamethasone (Wang et al. 2017; Huanga et al. 2020).

In vivo anti-inflammatory activity of SSa and SSd were evaluated in female albino rats by granuloma pouch method and antigranulomatous action by cotton pellet method. Oral administration of SSa and SSd showed significant anti-inflammatory effect without affecting hematocrit and plasma-11-OH-corticosteroid levels (Yamamoto et al. 1975). The anti-inflammatory effect of saikosaponins were evaluated in mice paw oedema model, wherein oedema was induced by injecting formalin into hind par and paw oedema index was used as a measure of anti-inflammatory effect.

Saikosaponins were found not only to decrease paw oedema but also decreased inflammatory metabolites including nicotinate, niacinamide, arachidonic acid (AA), and 20carboxy-leukotriene B4 as evidenced by HPLC metabolomic study. The study concluded that saikosaponins exert their anti-inflammatory effect through regulation of nicotinate and nicotinamide and arachidonic acid metabolism (Ma et al. 2016). The saikosaponins have also been reported to exhibit anti-inflammatory effect in TPP-induced ear oedema model in rats (Recio et al. 1995). In experimental model of cigarette smoke induced lung inflammation in mice, SSa was found to inhibit inflammatory cell infiltration, nitric oxide production, TNF- α , and IL-1 β production, MPO and MDA levels in lung tissues (Chen et al. 2018; Huanga et al. 2020).

CONCLUSION

The findings of the present study suggests that *B. turcicum* is an unexplored and underutilized species of genus *Bupleurum* L which is a rich source of saikosaponin A and D having potential antiviral properties against a number of virus including IAV, HBV, PCV-2, SARS-CoV and SARS-CoV-2. Further phytochemical characterization and standardization of *B. turcicum* root extract might be beneficial in finding safe and effective alternative therapy with respect to COVID-19.

Data Availability Statement: The database generated and /or analysed during the current study are not publicly available due to privacy, but are available from the corresponding author on reasonable request.

Conflict of Interests: Authors declare no conflicts of interests to disclose.

REFERENCES

Ashour, M. L., and Wink, M. L. (2011). Genus Bupleurum: a review of its phytochemistry, pharmacology and modes of action. Journal of Pharmacy and Pharmacology, 63(3),

305-321

Bahbah, E. I., Negida, A., and Nabet, M. S. (2020). Purposing Saikosaponins for the treatment of COVID-19. Medical Hypotheses, 140, 109782.

Benito, B. P, Martínez, A. M. J., Sen, S. A. M., et al. (1998). In vivo and *in vitro* anti-inflammatory activity of saikosaponins. Life Sciences, 63(13), 1147-1156.

Bermejo, P., Abad, M. A. A., Díaz, A. M., et al. (2002). Antiviral activity of seven iridoids, three saikosaponins and one phenylpropanoid glycoside extracted from *Bupleurum rigidum* and *Scrophularia scorodonia*. Planta Medica, 68(2), 106–110.

Chan, J. F., Yuan, S., Kok, K. H., et al. (2020). A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to person transmission: a study of a family cluster. Lancet, 395(10223), 514–523.

Chang, J. S., Wang, K. C., Liu, H. W., et al. (2007). Shosaiko-to (Xiao-Chai-Hu-Tang) and crude saikosaponins inhibit hepatitis B virus in a stable HBV producing cell line. American Journal of Chinese Medicine, 35(2), 341–351.

Chen, J., Duan, M., Zhao, Y., et al. (2015). Saikosaponin A inhibits influenza A virus replication and lung immunopathology. Oncotarget, 6(40), 42541-42556.

Chen, R., Guo, X., Cheng, B., et al. (2018). Saikosaponin a Inhibits Cigarette Smoke-Induced Oxidant Stress and Inflammatory Responses by Activation of Nrf2. Inflammation, 41(4), 1297–1303.

Cheng, P. W., Ng, L. T., Chiang, L. C., et al. (2006). Antiviral effects of saikosaponins on human coronavirus 229E *in vitro*. Clinical and Experimental Pharmacology and Physiology, 33(7), 612–616.

Chiang, L. C., Ng, L. T., Liu, L. T., et al. (2003). Cytotoxicity and anti-hepatitis B virus activities of saikosaponins from Bupleurum species. Planta Medica, 69(8), 705–709.

Davis, P. H. (1972). Flora of Turkey and the East Aegean Islands, Edinburgh University Press.

Goswami, T., and Bagchi, B. (2020). Molecular docking study of receptor binding domain of SARS-CoV-2 spike glycoprotein with saikosaponin, a triterpenoid natural product. Preprints ChemRxiv, https://doi.org/10.26434/ chemrxiv.12033774.v1

Gralinski, L., and Menachery, V. (2020). Return of the Coronavirus: 2019-nCoV. Viruses, 12(2), 135.

Herold, S., Steinmueller, M., Wulffen, W. V., et al. (2008). Lung epithelial apoptosis in influenza virus pneumonia: the role of macrophage-expressed TNF related apoptosisinducing ligand. The Journal of Experimental Medicine, 2008. 205(13), 3065-3077.

Huanga, F., Lib, Y., Leungc, E. L. H., et al. (2020). A review of therapeutic agents and Chinese herbal medicines against SARSCOV-2 (COVID-19). Pharmacological Research, 158(2020), 104929.

Jassim, S. A. A., and Naji, M. A. (2003). Novel antiviral agents: a medicinal plant perspective. Journal of Applied Microbiology, 95(3), 412–427.

Jin, X., Zhang, Y., Li, Q., et al. (2013). Mechanisms underlying the beneficial effects of Kaiyu Granule for depression. Neural Regeneration Research, 8(34), 3241– 3248.

Kars, G., Kars, M. D., Akin, M., et al. (2012). Determination of saikosaponin, phenolic and podophyllotoxin contents of five endemic Bupleurum root extracts and their effects on MCF-7 cells. Journal of Medicinal Plants Research, 6(5), 825-832.

Kato, M., Pu, M. Y., Isobea, K. I., et al. (1995). Cell typeoriented differential modulatory actions of saikosaponin-d on growth responses and DNA fragmentation of lymphocytes triggered by receptor-mediated and receptor bypassed pathways. Immunopharmacology, 29(3), 207– 213.

Kim, B. M. (2018). The role of saikosaponins in therapeutic strategies for age-related diseases. Oxidative Medicine and Cellular Longevity, Article ID 8275256.

Kim, S. O., Park, J. Y., Jeon, S. Y., et al. (2015). Saikosaponin a, an active compound of *Radix bupleuri*, attenuates inflammation in hypertrophied 3T3-L1 adipocytes via ERK/NF-κB signaling pathways. International Journal of Molecular Medicine, 35(4), 1126-1132.

Kumazawa, Y., Takimoto, H., Nishimura, C., et al. (1989). Activation of murine peritoneal macrophages by saikosaponin a, saikosaponin d and saikogenin d. International Journal of Immunopharmacology, 11(1), 21–28.

Lee, W. P., Lan, K. L., Liao, S. X., et al. (2019). Antiviral effect of saikosaponin B2 in combination with daclatasvir on NS5A resistance-associated substitutions of hepatitis C virus. Journal of Chinese Medical Association, 82(5), 368-374.

Leung, C. Y., Liu, L., Wong, R. N., et al. (2005). Saikosaponin-d inhibits T cell activation through the modulation of PKC θ , JNK, and NF- κ B transcription factor. Biochemical and Biophysical Research Communications, 338(4), 1920-1927.

Lin, L. T., Chung, C. Y., Hsu, W. C., et al. (2015). Saikosaponin b2 is a naturally occurring terpenoid that efficiently inhibits hepatitis C virus entry. Journal of Hepatology, 62(3), 541-548.

Lin, L., Han, Y., and Yang, Z. M. (2003). Clinical observation on 103 patients of severe acute respiratory syndrome treated by integrative traditional Chinese and Western medicine. Zhongguo Zhong Xi Yi Jie He Za Zhi, 23(6), 409–413.

Lin, T. Y., Chiou, C. Y., and Chiou, S. J. (2013). Putative genes involved in saikosaponin biosynthesis in Bupleurum

species. International Journal of Molecular Sciences, 14(6), 12806–12826.

Lu, H., Stratton, C.W., and Tang, Y. W. (2020). Outbreak of pneumonia of unknown etiology in Wuhan, China: the mystery and the miracle. Journal of Medical Virology, 92(4), 401–402.

Ma, Y., Bao, Y., Wang, S., et al. (2016). Anti-Inflammation effects and potential mechanism of saikosaponins by regulating nicotinate and nicotinamide metabolism and arachidonic acid metabolism. Inflammation, 39(4), 1453-1461.

Manicassamy, B., Manicassamy, S., Belicha-Villanueva, A., et al. (2010). Analysis of in vivo dynamics of influenza virus infection in mice using a GFP reporter virus. Proceedings of National Academy of Science USA, 107(25), 11531-11536.

McCutcheon, A. R., Roberts, T. E., Gibbons, E., et al. (1995). Antiviral screening of British Columbian medicinal plants. Journal of Ethnopharmacology, 49(2), 101–110.

Motoo, Y., and Sawabu, N. (1994). Antitumor effects of saikosaponins, baicalin and baicalein on human hepatoma cell lines. Cancer Letters, 86(1), 91-95.

Nose, M., Amagaya, S., and Ogihara, Y. (1989). Corticosterone secretion-inducing activity of saikosaponin metabolites formed in the alimentary tract. Chemical and Pharmaceutical Bulletin, 37(10), 2736-2740.

Patel, A., and Jernigan, D. B. (2020). Initial public health response and interim clinical guidance for the 2019 novel coronavirus outbreak - United States, December 31, 2019- February 4, 2020. MMWR Morbidity and Mortality Weekly report, 69 (5), 140–146.

Recio M. D. C., Just, M. J., Giner, R.M., et al. (1995). Anti-inflammatory activity of saikosaponins from *Heteromorpha trifoliata*. Journal of Natural Products, 58(1), 140-144.

Sahin, A. R., Erdogan, A., Agaoglu, M. P., et al. (2020). 2019 Novel Coronavirus (COVID-19) Outbreak: A Review of the Current Literature. Eurasian Journal of Medicine and Oncology, 4(1), 1-7.

Saraçoğlu, H. T., Akın, M., Demirci, B., et al. (2012). Chemical composition and antibacterial activity of essential oils from different parts of endemic *Bupleurum* L. species. Ankara Üniv Vet Fak Derg, 59, 265-270.

Shahrajabian, M. H., Sun, W., Shenb, H., et al. (2020). Chinese herbal medicine for SARS and SARS-CoV-2 treatment and prevention, encouraging using herbal medicine for COVID-19 outbreak. Acta Agriculturae Scandinavica, 70(5), 437–443.

Sinha, S. K., Shakya, A., Prasad, S. K., et al. (2020). An *in-silico* evaluation of different Saikosaponins for their potency against SARS-CoV-2 using NSP15 and fusion spike glycoprotein as targets. Journal of Biomolecular Structure and Dynamics, DOI: 10.1080/07391102.2020.1762741

Spiegel, M., Schneider, K., Weber, F., et al. (2006). Interaction of severe acute respiratory syndromeassociated coronavirus with dendritic cells. Journal of General Virology, 87(7), 1953-1960.

Sui, C., Zhang, J., Wei, J., Chen, S., Li, Y., Xu, J., and Xu, Y. (2011). Transcriptome analysis of *Bupleurum chinense* focusing on genes involved in the biosynthesis of saikosaponins. BMC Genomics, 12(1), 539.

Tykheev, Z. A., Anenkhonov, O. A., Zhigzhitzhapova, S. V., et al. (2020). Do compositions of lipid fraction correspond to species differentiation in *Bupleurum* L. (Apiaceae)?. Plants. Nov;9(11):1407.

Ushio, Y., and Abe, H. (1991). The effects of saikosaponin on macrophage functions and lymphocyte proliferation. Planta Medica, 57(6), 511–514.

Ushio, Y., and Abe, H. (1992). Inactivation of measles virus and herpes simplex virus by saikosaponin d. Planta Medica, 58(2), 171–173.

Van-Wyk, B. E., and Wink, M. (2004). Medicinal Plants of the World: an Illustrated Scientific Guide to Important Medicinal Plants and Their Uses. 1st ed. Portland, Orlando, Timber Press.

Vlietinck, A. J., and Vanden Berghe, D. A. (1991). Can ethnopharmacology contribute to the development of antiviral drugs? Journal of Ethnopharmacology, 32(1-3), 141–153.

Wang, Y., Guo, Q., Cheng, Z., et al. (2017). New saikosaponins from the roots of *Bupleurum chinense*. Phytochemistry Letters, 21, 183-189.

WHO (2020). WHO Director-General's remarks at the media briefing on 2019-nCoV on 11 February 2020, (2020), https://www.who.int/dg/ speeches/detail/who-director-general-s-remarks-at-the-media-briefing-on-2019-ncov-on-11-february-2020.

Wong, V. K., Zhou, H., Cheung, S. S., et al. (2009). Mechanistic study of saikosaponin-d (Ssd) on suppression of murine T lymphocyte activation. Journal of Cellular Biochemistry, 107(2), 303-315.

Wu, G. C., Wu, H., Fan, L. Y., et al. (2011). Saikosaponins: a potential treatment option for systemic lupus erythematosus. Irish Journal of Medical Science, 180(1), 259–261.

Wu, J. N. (2005), An Illustrated Chinese Materia Medica. New York, Oxford University Press.

Wu, S. J., Lin, Y. H., Chu, C. C., et al. (2008). Curcumin or saikosaponin a improves hepatic antioxidant capacity and protects against CCl4-induced liver injury in rats. Journal of Medicinal Food, 11(2), 224–229.

Wu, S. J., Tam, K. W., Tsai, Y. H., et al. (2010). Curcumin and saikosaponin a inhibit chemical-induced liver inflammation and fibrosis in rats. The American Journal of Chinese Medicine, 38(1), 99–111.

Xiao, Z., Li, Y., Chen, R., et al. (2003). A retrospective study of 78 patients with severe acute respiratory

syndrome. Chinese Medical Journal, 116 (6), 805–810. Xie, H., Huo, K. K., Chao, Z., et al. (2009). Identification of crude drugs from Chinese medicinal plants of the genus *Bupleurum* using ribosomal DNA ITS sequences. Planta Medica, 75(1), 89–93.

Yamamoto, M., Kumagai, A., and Yamamura, Y. (1975). Structure and actions of saikosaponins isolated from *Bupleurum falcatum* L. I. Anti-inflammatory action of saikosaponins. Arzneimittelforschung, 25(7), 1021-3102.

Yan, Y. M., Shen, X., Cao, Y. K., et al. (2019). Discovery of anti-2019-nCoV agents from Chinese patent drugs via docking screening. Preprints Preprints.org, https://doi: 10.20944/preprints202002.0254.v1

Yang, H., Chen, X., Jiang, C., et al. (2017). Antiviral and immunoregulatory role against PCV2 in vivo of Chinese herbal medicinal ingredients. Journal of Veterinary Research 61(4), 405-410.

Yin, F., Pan, R., Chen, R., and Hu, Let al. (2008). Saikosaponins from *Bupleurum chinense* and inhibition of HBV DNA replication activity. Natural Product Communications, 3(2), 155–157.

Ying, Z. L., Li, X. J., Dang, H., et al. (2014). Saikosaponin-d affects the differentiation, maturation and function of monocyte-derived dendritic cells. Experimental and

Therapeutic Medicine, 7(40), 1354-1358.

Yuan, B., Yang, R., Ma, Y., et al. (2017). A systematic review of the active saikosaponins and extracts isolated from *Radix bupleuri* and their applications. Pharmaceutical Biology, 55(1), 620-635.

Zhang, B. Z., Guo, X. T., Chen, J. W., et al. (2014). Saikosaponin-D attenuates heat stress-induced oxidative damage in LLC-PK1 cells by increasing the expression of anti-oxidant enzymes and HSP72. The American Journal of Chinese Medicine, 42(5), 1261–1277.

Zhao, C. H., Guo, Y. B., Wu, H., et al. (2003). Clinical manifestation, treatment, and outcome of severe acute respiratory syndrome: analysis of 108 cases in Beijing. Zhonghua Yi Xue Za Zhi, 83(11), 897–901.

Zhong, N. S., and Zeng, G. Q. (2003). Our strategies for fighting severe acute respiratory syndrome (SARS). American Journal of Respiratory and Critical Care Medicine, 168(1), 7–9.

Zhu, J., Luo, C., Wang, P., et al. (2013). Saikosaponin A mediates the inflammatory response by inhibiting the MAPK and NF- κ B pathways in LPS-stimulated RAW 264.7 cells. Experimental and Therapeutic Medicine, 2013, 5, 1345-1350.

Zhu, N., Zhang, D., Wang, W., et al. (2020). Novel coronavirus from patients with pneumonia in China, 2019. New England Journal of Medicine, 382, 727–733.