

## *In vitro* Evaluations of Anti-inflammatory and Antioxidant Activity of Ethanolic Leaf Extract of *Gymnema sylvestre* R. Br.

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### ABSTRACT

*Gymnema sylvestre* (Apocynaceae) is a well-known anti-diabetic herb used in various traditional Indian medicinal systems including Ayurveda and modern medicine. It is a source of diverse phytoconstituents and was reported to be used to treat various diseases. Hence our study aimed to evaluate the anti-inflammatory and antioxidant properties of *Gymnema sylvestre* by *in vitro* techniques. The ethanolic leaves extract of *G. sylvestre* was preliminarily screened to identify the presence of various phytoconstituents, and further analyzed for total alkaloids, flavonoids, phytosterols, and saponins estimations. The *in vitro* anti-inflammatory activity was assayed by human red blood cell (HRBC) membrane stabilization and protein denaturation assay, while antioxidant activity was assessed by DPPH and nitric oxide scavenging assay. Our study results showed that the ethanolic leaf extract of *G. sylvestre* exhibited dose-dependent anti-inflammatory and antioxidant activity. Preliminary phytochemical screening revealed the presence of alkaloids, flavonoids, phytosterols, tannins, phenolic compounds, terpenoids, glycosides, carbohydrates, sugars, proteins, amino acids, and saponins. Quantitative analysis showed high content of saponin  $32.44 \pm 5.65$  mg diosgenin/g, followed by steroidal content  $26.30 \pm 3.96$  mg cholesterol/g, flavonoids  $17.66 \pm 0.43$  mg QE/g, and alkaloids  $10.33 \pm 0.97$  mg AE/g. These results further authenticate the claim and use of *G. sylvestre* in traditional medicine to treat inflammation and oxidative stress.

**KEY WORDS:**ANTIOXIDANT, ANTI-INFLAMMATORY, GYMNEMA SYLVESTRE, DPPH, NITRIC OXIDE.,

### INTRODUCTION

Oxidative stress has a significant role in chronic inflammatory illnesses such as cancer, diabetes, neurological problems, and cardiovascular ailments. Prolonged exposure to high quantities of pro-oxidants can cause mitochondrial DNA damage and changes in cellular components, which contribute to gene expression anomalies (Sharifi-Rad et al., 2020). The process of oxidative stress that leads to the damage of numerous tissue's physiological and biochemical environments, a minor amount of oxidative stress that helps the immune system withstand microbial infections and intracellular cell signalling, is also thought to be an important physiological activity (Sies et al., 2017). When free radicals outnumber antioxidant defences, oxidative stress, a process that harms the physiological and biochemical balance of tissues, becomes worse (Kopáni et al., 2006). Inflammation increases reactive oxygen species (ROS) generation, which

exceeds the body's antioxidant capacity and causes oxidative stress-induced tissue damage (Sies 1997).

Early in the ages, medicinal plants have been extensively used to treat a wide range of diseases, notably in India, where indigenous medical systems such as Ayurveda, Siddha, and Unani have been practiced for centuries. Herbalism, or the medicinal use of herbal plants, is important in modern medicine. While synthetic drugs provide treatments for a variety of illnesses, their large quantity and related side effects sometimes limit accessibility. The use of herbs, on the other hand, is gaining popularity because of low toxicity, low cost, and broad availability, emphasizing the importance of herbs in everyday life (Das et al., 2022).

Nearly 80% of the world's population gets their basic medical treatment from medicinal plants, according to the World Health Organisation (WHO), highlighting the long-standing importance of herbal medicine in treating illnesses affecting human health (Dey et al., 2021). Traditional methods of treating inflammatory illnesses mainly target the fight against bacterial infections, which might not be effective enough. As a result, complementary approaches that target both inflammation and oxidative stress reduction

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have developed as viable therapeutic and preventative treatments. In this context, antioxidant compounds, which may be found in a variety of foods, drinks, plants, vitamins, and minerals are essential. Both conventional insight and contemporary studies have highlighted the anti-inflammatory and antioxidant qualities of medicinal plants. Despite their effectiveness, modern anti-inflammatory drugs can have a variety of negative effects. On the other hand, because of their natural source, herbal extracts are thought to provide a safer substitute. Treating chronic inflammatory diseases is made easier when a single plant extract combines anti-inflammatory and antioxidant effects (Somashekar et al., 2022).

*Gymnema sylvestre* (*G. sylvestre*), Apocynaceae family, is a well-established antidiabetic herb utilized in various traditional Indian medicinal systems including Ayurveda and modern medicine. Known as "Gudmar," this herbaceous plant is a climbing species found in dry forests up to 600 meters in height. Its leaves are extensively employed for their multifaceted therapeutic properties, encompassing antidiabetic, anti-inflammatory, antiarthritic, anti-obesity, wound healing, astringent, bitter, acrid, thermogenic, anodyne, digestive, antipyretic, stomachic, diuretic, laxative, cardiogenic, and liver tonic effects. Rich in tannins, flavonoids, saponins, and gymnemic acid, *G. sylvestre* exhibits additional bioactive characteristics such as antimicrobial, larvicidal, antiviral, hypolipidemic, anticancer, and antioxidant activities. particularly gymnemic acids, a blend of at least 17 distinct saponins, acidic glycosides, and anthraquinones (Sudhakar et al., 2018, Pachiappan et al., 2021 Pachiappan et al., 2023).

The research regarding the pharmacological effects of *G. sylvestre* is currently at an early stage and requires further investigation to fully understand the phytochemical advantages of this herb. This *in vitro* study is designed to examine the antioxidative and anti-inflammatory properties of *G. sylvestre* leaf ethanolic extract.

## MATERIALS AND METHODS

**Plant material:** The fresh leaves of *Gymnema sylvestre* R. Br. (GS) were collected from Kolli Hills Namakkal, Tamil Nadu, India, during January 2021. The plant material was further authenticated by Dr. P. Radha, Research officer (Botany), Siddha Medicinal Plants Garden (Central Council for Research in Siddha), Ministry of AYUSH, Govt. of India, Mettur Dam, Tamil Nadu, India, where the voucher specimen is preserved with the reference number of G180221012S.

**Preparation of extract:** The collected fresh leaves were washed with running water to remove the sand and dust. Then the shade-dried material was coarsely powdered for extraction. 100 gm of the powdered plant material was loaded in the Erlenmeyer flask and extracted by cold maceration with 95% ethanol for 72 hours at room temperature with occasional shaking. After 72 hours, the filtrate was separated from marc by using a muslin cloth and further filtered by Whatman no. 1 filter paper. The same procedure was performed two consecutive times with the

marc material. All three filtrates were mixed and evaporated under reduced pressure and controlled temperature at 40 °C in a rotary evaporator until all the solvent was removed. The dried material was stored in an airtight container at 4 °C until further use.

**Preliminary phytochemical analysis of ethanolic extract of *Gymnema sylvestre*:** The preliminary phytochemical screening of *G. sylvestre* leaves ethanolic extract was carried out using standard procedure to identify the presence of alkaloids, flavonoids, phytosterols, tannins, phenolic compounds, terpenoids, glycosides, carbohydrates, sugars, proteins, amino acids, and saponins (Evans & WC 2009, Kokate et al., 2017).

**Quantitative phytochemical analysis: Determination of total alkaloid content:** The extract (1 mg/ml), bromocresol green solution, and phosphate buffer 5 ml each were mixed in a separating funnel. It was diluted with chloroform in 10 ml volumetric flask. Atropine was used as standard. The absorbance of both the standard and test solution was observed at 470 nm. The total alkaloid content in the extract was expressed as mg of Atropine (AE)/g of plant extract (Shalini et al., 2021).

**Determination of total flavonoid content:** The plant extract 3 mg was dissolved in methanol 3 ml for flavonoid content estimation. The quercetin was used as standard. 3 ml of plant extract or standard was mixed with 1 ml of 2% AlCl<sub>3</sub> methanolic solution and allowed to stand for 60 min at room temperature. The absorbance was measured at 420 nm. The total flavonoid content was indicated as mg of QE/g of extract (Garg & Garg 2019).

**Determination of total steroidal content:** The total steroidal content was estimated by Liebermann-burchard colorimetric method with slight modification. The *G. sylvestre* extract was dissolved with chloroform and the freshly prepared Liebermann-burchard reagent was added. The absorbance of standard cholesterol and extract was measured at 650 nm against a reagent blank. The total steroidal content was expressed as mg of cholesterol/g of extract (Kim & Goldberg 1969).

**Determination of total saponin content:** The 1 ml of diluted *G. sylvestre* extract was mixed with 1 ml of 80% aqueous methanol, followed by 1 ml of 72% H<sub>2</sub>SO<sub>4</sub> added in the sides of test tube. The mixture was warmed on 60 °C for 10 minutes. The absorbance of standard diosgenin and extract solution was measured at 544 nm against 80% methanol as a blank solution. The total saponin content was expressed as mg of diosgenin equivalent to gm of dry weight of the extract (Nandhini & Ilango 2020).

***In vitro* anti-inflammatory activity screening : Human red blood cell (HRBC) membrane stabilization assay:** The *in vitro* anti-inflammatory activity of *G. sylvestre* was determined by the Human Red Blood Cell (HRBC) membrane stabilization method. The blood sample was collected from a healthy human volunteer who had not taken any NSAIDs for 2 weeks before the experiment, added with an equal amount of Alsever's solution. This mixture was

centrifuged at 3000 rpm for 15 min. The RBC pellet was washed thrice with sterile saline till the supernatant was clear and colorless.

The packed cellular content was formulated as 10 % v/v suspension with sterile isosaline. 1 ml of different concentrations (100-500 µg/ml) of *G. sylvestre* extract and standard diclofenac was mixed with 1 ml of phosphate buffer, 0.5 ml of HRBC suspension, and 2 ml of hyposaline. After 30 min incubation at 37°C, the reaction mixture was centrifuged at 3000 rpm for 10 min. The supernatant absorbance was observed spectrophotometrically at 560 nm. The percentage of hemolysis was estimated by considering the percentage of hemolysis of control as 100% (Senthil Kumar et al., 2018; Gupta et al., 2021). The percentage of protection/ percentage inhibition of hemolysis were evaluated using the formula

$$\% \text{ protection} = 100 - \frac{\text{Absorbance of test}}{\text{Absorbance of control}} \times 100$$

**Protein denaturation assay:** The *in vitro* anti-inflammatory activity of the *G. sylvestre* extracts by protein denaturation was performed using bovine serum albumin. The increasing concentrations of the extract (100-500 µg/ml) and reference compound diclofenac sodium were incubated with 0.5% w/v of bovine serum albumin at 37°C for 20 min and the temperature was increased to keep the samples at 57 °C for 30 min. After reaching room temperature, the turbidity was measured using UV-Visible spectrophotometer at 660 nm following the addition of 2.5 ml of phosphate-buffered saline (Shanmuganathan et al., 2017; Senthil Kumar et al., 2018). The percentage inhibition of protein denaturation was calculated by using the following formula.

$$\% \text{ inhibition} = \frac{(\text{Absorbance of control} - \text{Absorbance of test})}{\text{Absorbance of Control}} \times 100$$

**In vitro antioxidant activity screening :DPPH free radical scavenging assay:** The 2, 2 diphenyl-1-picrylhydrazyl (DPPH) 0.2mM solution 1 ml was added to 1 ml of different concentrations (100-500 µg/ml) of *G. sylvestre* extract, the mixture was kept room temperature for 50 mins in dark environment. The antioxidant activity was measured spectrophotometrically at 517 nm. Ascorbic acid was used as standard. The percentage of free radical scavenging was calculated as half minimal inhibitory concentration (IC<sub>50</sub>). IC<sub>50</sub> denotes the concentration of the sample required to inhibit 50% of DPPH free radicals (Govindappa et al., 2018). The DPPH radical scavenging capacity was calculated by using the following formula:

$$\text{DPPH radical scavenged \%} = \frac{(\text{Absorbance of control} - \text{Absorbance of test})}{\text{Absorbance of Control}} \times 100$$

**Nitric oxide (NO) radical scavenging activity:** The nitric oxide scavenging assay reaction mixture (3 ml) contains 2 ml of 10 mM sodium nitroprusside, 0.5 ml of phosphate buffer, and 0.5 ml of different concentrations (100-500 µg/ml) of *G. sylvestre* extract. The reaction mixture was incubated at 25 °C for 150 min. The 0.5 ml of reaction mixture was pipetted and mixed with 1 ml of sulfanilic acid reagent and allowed to stand for 5 min for diazotization. Then 1 ml of naphthyl ethylene diamine dihydrochloride

was added and incubated at 25 °C for 30 min. A pink-colored chromophore was formed in diffused light. Ascorbic acid was used as standard. The NO scavenging activity was measured at 550 nm and the results were expressed as a percentage (%) of scavenging using the following formula: (Adebayo et al., 2019)

$$\text{Nitric oxide radical scavenged \%} = \frac{(\text{Absorbance of control} - \text{Absorbance of test})}{\text{Absorbance of Control}} \times 100$$

**Statistical analysis:** The statistical analysis was performed in IBM SPSS version 18 (SPSS version 18.0; IBM Corporation, Armonk, NY, USA). The entire assay was performed in triplicate and the values were expressed as mean ± standard deviation (SD). Descriptive statistics were used for continuous variables and expressed in mean and standard deviation. A test of normality was applied.

## RESULTS AND DISCUSSION

**Extractive yield and preliminary phytochemical analysis:** The ethanolic extractive percentage yield of *Gymnema sylvestre* leaf extract was calculated as 22.08 gm% w/w. The preliminary phytochemical analysis of ethanolic leaves extract of *G. sylvestre* showed the presence of alkaloids, flavonoids, phytosterols, tannins, phenolic compounds, terpenoids, glycosides, carbohydrates, sugars, proteins, amino acids, and saponins.

**Quantitative phytochemical analysis:** Table 1 illustrates the quantification of secondary metabolites alkaloids, flavonoids, steroids, and saponins in the ethanolic leaves extract of *G. sylvestre*. The secondary metabolite saponins content was found to be higher 32.44 ± 5.65 mg diosgenin/g, followed by steroidal content 26.30 ± 3.96 mg cholesterol/g, flavonoids 17.66 ± 0.43 mg QE/g, and alkaloids 10.33 ± 0.97 mg AE/g in the ethanolic leaves extract of *G. sylvestre*.

**Table 1. Quantitative phytochemical analysis of *G. sylvestre* leaves ethanolic extract**

S. No.	Phytoconstituent	Quantity of Phytoconstituent (mg/g of extract)
1.	Alkaloids	10.33 ± 0.97
2.	Flavonoids	17.66 ± 0.43
3.	Steroids	26.30 ± 3.96
4.	Saponins	32.44 ± 5.65

Values are expressed as mean ± SD, n=3.

**In vitro anti-inflammatory activity:** The *in vitro* anti-inflammatory activity of *G. sylvestre* was carried out by Human red blood cell membrane stabilization assay and protein denaturation assay techniques.

**HRBC membrane stabilization assay:** HRBC assay is based on the principle of evaluating lysosomal membrane

protection. During inflammation lysosomes undergo lysis and release specific enzymes into circulation leading to inflammatory diseases. The RBC membrane is similar to the lysosomal membrane, and its stability indicates anti-inflammatory properties (Yesmin et al., 2020). In this study, the hemolysis of RBC was impacted by the hypotonicity of hyposaline, which induces lysis of the cell membrane. The percentage of RBC lysis/protection was taken as an index for anti-inflammatory activity measurement (Kumar et al., 2011 and Kumar et al., 2020).

The treatment of *G. sylvestre* showed dose-dependent membrane stabilization action the maximum activity ( $72.46 \pm 2.58$ ) was observed at the concentration 500  $\mu\text{g/ml}$ . The standard diclofenac showed maximum membrane protection ( $89.27 \pm 1.08$ ) activity at the concentration 500  $\mu\text{g/ml}$ , comparably the treatment of *G. sylvestre* leaves extract showed less percentage inhibition of hemolysis compared to the standard diclofenac. The treatment of *G. sylvestre* ethanolic leaves extract exhibited remarkable anti-inflammatory activity by stabilizing the RBC membrane, preventing the discharge of lytic enzymes and other inflammation mediators (Table 2).

**Table 2. Human red blood cell membrane stabilization assay of *G. sylvestre* leaves ethanolic extract**

S. No.	Concentration ( $\mu\text{g/ml}$ )	% Protection	
		Ethanolic extract of GS	Diclofenac (Standard)
1.	100	$23.51 \pm 1.01$	$48.11 \pm 2.77$
2.	200	$34.25 \pm 0.98$	$66.96 \pm 2.54$
3.	300	$40.19 \pm 1.50$	$75.39 \pm 7.80$
4.	400	$56.88 \pm 3.55$	$78.93 \pm 4.13$
5.	500	$72.46 \pm 2.58$	$89.27 \pm 1.08$

Values are expressed as mean  $\pm$  SD, n=3

**Table 3. Protein denaturation inhibition assay of *G. sylvestre* leaves ethanolic extract**

S. No.	Concentration ( $\mu\text{g/ml}$ )	% inhibition	
		Ethanolic extract of GS	Diclofenac (Standard)
1.	100	$18.88 \pm 1.42$	$24.71 \pm 3.31$
2.	200	$27.12 \pm 1.32$	$39.14 \pm 6.04$
3.	300	$38.41 \pm 0.70$	$52.90 \pm 1.11$
4.	400	$58.62 \pm 2.45$	$67.22 \pm 0.83$
5.	500	$71.89 \pm 7.89$	$78.70 \pm 2.81$

Values are expressed as mean  $\pm$  SD, n=3.

**Table 4. DPPH radical scavenging activity of *G. sylvestre* leaves ethanolic extract**

S. No.	Concentration ( $\mu\text{g/ml}$ )	% scavenging activity	
		Ethanolic extract of GS	Ascorbic acid (Standard)
1.	100	$27.32 \pm 2.56$	$32.07 \pm 1.22$
2.	200	$38.58 \pm 5.92$	$59.33 \pm 5.05$
3.	300	$46.65 \pm 9.02$	$62.43 \pm 3.25$
4.	400	$68.45 \pm 4.52$	$74.21 \pm 3.82$
5.	500	$77.32 \pm 6.29$	$82.91 \pm 6.64$
	IC <sub>50</sub>	$289.15 \pm 11.45$	$196.38 \pm 8.19$

Values are expressed as mean  $\pm$  SD, n=3.

**Table 5. Nitric oxide radical scavenging activity of *G. sylvestre* leaves ethanolic extract**

S. No.	Concentration ( $\mu\text{g/ml}$ )	% scavenging activity	
		Ethanolic extract of GS	Ascorbic acid (Standard)
1.	100	$29.45 \pm 0.89$	$48.33 \pm 6.02$
2.	200	$40.99 \pm 1.34$	$61.73 \pm 4.05$
3.	300	$56.05 \pm 2.62$	$75.00 \pm 2.39$
4.	400	$71.30 \pm 3.27$	$86.19 \pm 4.31$
5.	500	$80.12 \pm 6.80$	$97.68 \pm 3.24$
	IC <sub>50</sub>	$258.93 \pm 9.06$	$107.07 \pm 12.46$

Values are expressed as mean  $\pm$  SD, n=3.

**Protein denaturation assay:** The denaturation of cellular protein is the most known cause of inflammation and arthritis. The compounds that prevent the protein denaturation have advantageous anti-inflammatory properties (Osman et al., 2016). The results in table 3 shows that maximum ( $71.89 \pm 7.89$ ) percentage denaturation inhibition was observed in the *G. sylvestre* 500  $\mu\text{g/ml}$  treatment, when compared to standard diclofenac it shows maximum inhibition ( $78.70 \pm 2.81$ ) at the concentration of 500  $\mu\text{g/ml}$  which is almost equal. The ethanolic leaves extract of *G. sylvestre* showed dose dependent protein denaturation inhibition property. As results the protein denaturation inhibition capacity confirms the anti-inflammatory property of *G. sylvestre* extract.

**In vitro antioxidant assay:** The antioxidant potential of *G. sylvestre* leaves extract was assayed by DPPH free radical scavenging assay and Nitric oxide (NO) radical scavenging assay.

**DPPH free radical scavenging assay:** DPPH radical scavenging is the most widely used technique for

measuring free radical scavenging. The antioxidant ability of compounds was determined by their capacity to donate hydrogen was assumed to be responsible for DPPH scavenging. The DPPH antioxidant model may have some positive benefits in certain inflammatory disorders (Nagulsamy et al., 2015, Kawra et al., 2020). The *G. sylvestre* leaf extract and ascorbic acid showed a dose-response relationship with the DPPH scavenging activity which was directly proportional to their concentrations. The IC<sub>50</sub> value for ascorbic acid was 196.88 µg/ml, which was significantly lower than the *G. sylvestre* extract 289.15 µg/ml (Table 4).

**Nitric oxide radical scavenging assay:** Inflammatory diseases have a high production of nitric oxide. Overproduction of NO can cause tissue damage and contribute to inflammatory diseases including atherosclerosis and hypertension. The compounds that can scavenge or inhibit the production of NO are known to have antioxidant properties (Moncada et al., 1991, Adebayo et al., 2019). The *G. sylvestre* leaf extract and standard ascorbic acid showed a dose-response relationship with the NO scavenging activity which was directly proportional to their concentrations. The IC<sub>50</sub> value for ascorbic acid was 107.07 µg/ml, which was significantly lower than the *G. sylvestre* extract at 258.93 µg/ml (Table 5).

## CONCLUSION

The results of this study demonstrated that the ethanolic leaf extract of *G. sylvestre* inhibits hemolysis by protecting the RBC cell membrane in HRBC membrane stabilization assay and preventing protein denaturation in protein denaturation assay. Free radical scavenging property was demonstrated through DPPH radical scavenging and nitric oxide radical scavenging in a concentration-dependent manner. These suggest that *G. sylvestre* leaf ethanolic extract has extensive anti-inflammatory and antioxidant potency. The presence of secondary metabolites might contribute to its actions, the free radical scavenging activity of *G. sylvestre* leaf extract can play a vital role in modulating inflammatory reactions. Hence *Gymnema sylvestre* leaf extract may utilized for the herbal medicine development to the inflammation condition associated with oxidative stress. Further, in vivo studies and the specific bioactive compound isolation are needed to explore *Gymnema sylvestre* as a newer therapeutic agent.

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