

Toxicological Communication

Human Health Risk Surveillance of Organochlorine Pesticides in River Water and Fishes from Bangladesh

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ABSTRACT

Organochlorine pesticides (OCPs) are chlorinated hydrocarbons that used extensively in the last century for agricultural purposes. Excess use of OCPs results pesticide residues accumulation in the water and fishes and causes various health problems. The concentrations of Organochlorine pesticides (OCPs) residues in water and fish samples of six most contaminated rivers surrounding Dhaka, Bangladesh namely Buriganga, Turag, Balu, Sitalakhya, Bangshai and Dhaleswari were determined and assessed the possible health risks through consumption of accumulated OCPs in fishes from those rivers. As we know, OCPs are non-biodegradable and thus remain in the environment as pollutants. Therefore, OCPs exposure must be monitored and controlled to reduce the human health risks. Risk assessment is an important procedure to quantify the potential health risks and provides information the risk managers to control the overuse of OCPs. The concentrations of OCPs residues were determined by Gas Chromatography tandem Mass Spectrometry (GC-MS/MS). After sample collection, sample was extracted and analyzed according to the validated method. Several OCPs residues including Aldrin, p,p-DDE, Eldrine ketone, p,p-DDD, Endrin, α -Endosulfan, Heptachlorepoxide, p,p-DDT, Endosulfan sulfate and β -Endosulfan were detected in river water and residues including Aldrin, p,p-DDE, Heptachlorepoxide, p,p-DDD, Endrin and p,p-DDT were detected in fish samples. The Health risk index (HI) values of Aldrin, p,p-DDE, Heptachlorepoxide, p,p-DDD, Endrin and p,p-DDT from analyzed three fish species (*Acanthobrama microlepis*, *Barbonymus gonionotus* and *Batasio tengana*) were ranged from 0.036 to 1.696, 0.029 to 2.007, 0.022 to 1.117, 0.0231 to 0.721, 0.019 to 1.597 and 0.019 to 1.205 for Buriganga, Turag, Balu, Sitalakhya, Bangshai and Dhaleswari rivers respectively. OCPs are quantified in river water and fish samples and potential health risks are assessed. This study suggested that, there might have OCPs mediated health risks through long term exposure of OCPs residues from fishes of those polluted rivers. Although, our study provided estimation about the presence of OCPs in water and fishes from six contaminated rivers surrounding Dhaka city but, further studies are suggested to ensure the safety of peoples.

KEY WORDS: BANGLADESH, FISH, GAS CHROMATOGRAPHY, MASS SPECTROMETRY, OCPs.

INTRODUCTION

Organochlorine pesticides (OCPs) are synthetic chemicals with a strong chemical bond between their chlorine and carbon components. OCPs are toxic to the environment, insoluble in water and fat loving pesticides. OCPs are very dangerous for the environment because, they stay around for a long time after using and may exposure to water, soil and animal bodies. OCPs can contaminate the environment either through direct application or waste disposal and rain water.

Although, OCPs are banned in several developed countries because of their long lasting nature but, still they are used in some of the developing countries. Peoples especially from developing countries where OCPs are used regularly are at a higher risk of OCPs mediated health hazards. Peoples can directly uptake the OCPs residues through breathing and ingest OCPs by taking foods like fish, dairy products and other highly fat containing foods (Lee et al. 2020).

As OCPs maintains very strong bonding in the fatty parts so, they can accumulate in the animal or fish tissues and can pass to the human and other animals after consumption of those animals. When human takes OCPs residues through food for long time, they may suffer from serious health problems such as damage of liver, kidney, thyroid gland, bladder and

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central nervous system. In agricultural countries, pesticides are used to control of insects, weeds, fungi, bacteria, etc. OCPs are lipophilic in nature with longer half-life. The bioaccumulation and long range transportation ability of OCPs, makes them ideal candidate for contamination of water, air and soil. There has harmful impact of pesticides in aquatic eco systems (De Lorenzo et al. 2001; Frankart et al. 2003; Gupta 2004; Castillo et al. 2006; Aktar et al. 2009). Although, OCPs are used to prevent malaria and typhus but they are banned in the developed countries because of their harmful effects (Aktar et al. 2009). Several pesticides such as DDT, hexachlorocyclohexane (HCH) and aldrin used in some part of Asia due to cheaper price and activity against some harmful organisms. There have several reports on OCPs mediated toxicity in several fish models (Gupta 2004; Yohannes et al. 2014a; Yohannes et al. 2014b; Zhang et al. 2014; Byrne et al. 2015; Cui et al. 2015; Wang et al. 2015; Lee et al. 2020).

Endosulfan was found to responsible for necrosis of focal liver cells (Capkin et al. 2006). Endosulfan sulphate is a toxic OCPs residue. The toxicity was studied in Zebra fish (Lee et al. 2020). Fishes accumulate and concentrate these hydrophobic compounds in their tissues directly from water and the pesticide residues are transferred to the fish consumers directly by trophic food systems. Humans can be affected by OCPs after long time exposure. The poisoning of OCPs may cause headache, nausea, dizziness, vomiting, tremor, lack of co-ordination and mental confusion. The use of several OCPs can cause cancer. DDT is reported as a potential cancer-causing agent in humans according to several researchers (Enan and Matsumura 1998; Diel et al. 2002; Han et al. 2010; Ndebele et al. 2010). Reduction of sperm count and damage of spermatogonial cells and sperm morphology can be occurred because of high endosulfan dose (Ennaceur et al. 2008; Wong et al. 2015).

Higher concentrations of DDE and HCB reduced cell proliferation and produced binucleated cells (Ennaceur et al. 2008; Wong et al. 2015). Bangladesh is called land of rivers. Rivers are very important part of Bangladesh. The rivers provide water for cultivation in the vast majority of areas. Rivers are used for transportation in some areas of Bangladesh. There are about 230 rivers currently flowing throughout Bangladesh during summer and winter seasons. The major river includes Padma, Meghna, Jamuna, Brahmaputra, Karnaphuli etc. All of the rivers are connected to each other and finally falls to the Bay of Bengal. Dhaka, the capital of Bangladesh is surrounded with some rivers. The rivers are: Buriganga River, Turag River, Balu River, Shitalakhya River, Dhaleshwari River and Bangshai River. They supply water to the huge population of Dhaka, Bangladesh. The present condition of these rivers is at very high risk (Li et al. 2020).

The water quality of these six major rivers around Dhaka city is getting polluted day by day and becoming very dangerous for public health. Water from these rivers cannot be used as drinking water. The deteriorating water quality of these six rivers has been a major concern. Rivers are main sources of fish. Bangladeshi peoples prefer fishes on their dishes in almost every day. Fish is one of the importance

sources of protein, vitamin, fat and carbohydrate. Peoples from rural areas get their required protein through consumption of fishes. Fish is considered as one of the most significant indicators of metal pollution in aquatic environment (Rashed 2001). QuEChERS extraction method in combination with GC-MS was developed for the detection of OCPs in shellfish. OCPs analysis method was validated for shellfish and cephalopods (Li et al. 2020; Hwang et al. 2020). A method was developed for the detection of OCPs in potato and sweet potato. A QuEChERS Method with slight modification using N-Doped Graphitized Carbon coupled with GC-MS/MS has been developed for determination of OCPs in tomatoes (Ye et al. 2020; Singh et al. 2020).

In this study, we detected several OCPs residues from water and fish samples from six contaminated rivers (Buriganga River, Turag River, Balu River, Shitalakhya River, Bangshi River and Dhaleshwari River) surrounding Dhaka, the capital of Bangladesh and accessed potential health hazards of peoples who consumed the fishes from those rivers. The limit of detection (LOD) values were appropriate for the detection of OCP residues from both water and fish samples. Considering the problems in pesticide residues detection, this study conducted on quantitative detection of OCPs by GC-MS/MS. Although, LC-MS/MS is preferred method for the detection of OCPs but, GC-MS/MS is more sensitive method (Pico et al. 2020). Excess use of OCPs in agriculture results pesticide residues accumulation in the water and fishes which in turn causes various health problems. This research will helpful for the assessment of the impact of OCPs on public health, agriculture and environment in Dhaka, Bangladesh.

MATERIAL AND METHODS

In this study, both fish and water samples were collected from 6 (Six) most polluted rivers of Bangladesh namely Buriganga, Turag, Balu, Sitalakhya, Bangshai and Dhaleswari. These Rivers are very important as they surrounds Dhaka city. For the sample collection, several sampling points were selected. This study was carried out from November, 2020 to April, 2021. Water samples were collected from the 30-50 cm depth of water surface. Fish samples were collected where the fishing effort is very high. Three fish species from those rivers were collected for this assay: *Acanthobrama microlepis*, *Barbonymus gonionotus* and *Batasio tengana*. Fishes were purchased from the local fishermen. While sampling, fishes of almost same size and weight were selected. Fish samples were stored in polythene bags and transported to the laboratory. For the analysis, fish muscle tissues were selected. A total of 30 water samples and 60 fish samples of three species were collected randomly. After collection, samples were stored in proper condition to maintain the sample quality. OCPs were extracted from river water samples using liquid-liquid extraction (LLE) method (APHA 2002).

OCPs were extracted from river water samples using liquid-liquid extraction (LLE) method (APHA 2002). For the extraction of OCPs from fish samples, 2g of fish muscle samples were dissected. The muscle sample was grounded to fine powder using 5 g of Sodium sulfate. After extraction

with 50 ml of Acetone (2 times), the extracts were filtered and filtrates were again extracted by 350 ml of deionized water, 15 g of NaCl and 40 ml of n-hexane/ethyl acetate (3:2). After organic layer separation, the samples were again extracted using 40 ml of n-hexane / ethyl acetate (3:2) and again the organic layer was collected. The extracts were then passed through the anhydrous sodium sulfate, concentrated and added 20 ml of n-hexane. Finally, the extract was condensed to 1 ml in rotary evaporator (Model: RE100-Pro, DLAB, USA) for GC-MS/MS analysis. In this study, GC-MS system (Model: TRACE 1310, Thermo Fisher Scientific, USA) equipped with Thermo Scientific™ Trace GOLD™ TG-5MS GC Column (0.25 mm X 0.25 µm X 0.25 m) was used. Helium was the carrier gas with a flow rate of 1.2 ml/min. Injection port temperature was 230°C and temperature for GC was from 80°C to 290°C. 2 µL of sample was injected for analysis. Mass Spectrometer (Model: TSQ DUO, Thermo Scientific, USA) was used for MS analysis.

A stock solution of a mixture of 19 pesticides was used as standard solution. Several dilutions were prepared from Standard solution with a range of 5 ppb to 200 ppb. All of those dilutions were injected and analyzed. Blank solutions method was used to determine LOD and LOQ. Signal and noise ratio 3:1 was considered for LOD determination and 10 times of baseline value of blank samples was considered for LOQ detection. The accuracy of the method was validated by recovery performance evaluation. For the recovery performance evaluation, both river water and fish samples were spiked with two known concentrations: 5 µg / L and 10 µg/L for river water and 5 µg / kg and 10 µg/ Kg for fish samples. Then, the samples were extracted and analyzed with the following equation: $P_i = (S_i/T_i) \times 100$ (1); Where, P_i , S_i and T_i stands for recovery percentage; results of control from laboratory and percent recovery of the spiked samples respectively.

Table 1. Data obtained from the Chromatography of multi-standards of 19 OCPs. Pesticides, their retention times, limit of detection (LOD) and Limit of quantification (LOQ).

OCPs	Retention time (Min) RT	Water	Fish		
		LOQ	LOD	LOQ	LOD
		µg/L	µg/ kg		
Aldrin	20.39	0.042	0.012	0.039	0.013
p,p-DDE	24.51	0.067	0.023	0.056	0.029
Eldrine ketone	28.96	0.092	0.032	0.088	0.037
p,p-DDD	25.8	0.032	0.021	0.043	0.038
α-Endosulfan	23.47	0.076	0.023	0.067	0.032
Cis-Chordane	23.61	0.066	0.019	0.063	0.022
Heptachlorepoide	22.05	0.054	0.028	0.048	0.025
Heptachlor	18.94	0.087	0.026	0.078	0.034
p,p-DDT	27.2	0.053	0.013	0.064	0.039
Methoxychlor	26.92	0.064	0.012	0.048	0.028
Endrin	25.31	0.067	0.018	0.087	0.021
Endosulfan sulfate	27.28	0.054	0.024	0.059	0.036
α-Endosulfan	23.47	0.069	0.021	0.077	0.029
β-Endosulfan	25.67	0.042	0.019	0.054	0.022
Lindane-I	14.23	0.067	0.014	0.062	0.019
Lindane-II	15.48	0.054	0.031	0.048	0.027
Lindane -III	15.76	0.043	0.023	0.038	0.034
Lindane-IV	16.85	0.073	0.038	0.065	0.031
Trans-Chordane	23	0.064	0.027	0.059	0.018
Phthalic Acid	30.54	0.076	0.032	0.066	0.029

The estimated daily intake (EDI) of OCPs is based on the OCPs concentration in the fish and the amount of fish consumed. EDI was calculated by the following equation: $EDI = (C_{OCP \text{ residues}} \times D_{\text{fish intake}}) / B_{\text{Average weight}}$ (2); Here, C_{OCPs} (mg/kg), $D_{\text{fish intake}}$ (kg / person) and B average weight (kg / person) are the OCPs residual concentrations in fish, daily intake of fish and average body weight, respectively. HI is defined as health risk assessment of consumers from the consumption of fishes accumulated OCPs in their tissue.

HI is calculated from estimated daily intake (EDI) and FAO/WHO prescribed acceptable daily intake (ADI) of OCPs using the following formula:

$$HI = EDI/ADI \dots\dots\dots(3);$$

When, calculated HI value is less than 1, the consumers are considered safe from any OCPs mediated health risks (USEP 1986). But, if HI value is higher than 1 then it can be

indicated as potential health hazardous. For data collection, organization and summary preparation, Microsoft Office Excel 2007 and for statistical calculation Statistics 10 software was used.

RESULTS AND DISCUSSION

Method validation: The detection method was validated through recovery evaluation. The spiked river water and fish tissue samples were compared with that of blank samples. The percent recoveries of river water and fish samples were ranged from 87.5% to 99.4% and 87.0% to 100.4% respectively. A linear range was observed from the dilutions of standard solution. The coefficient values (R²) value from this study was 0.95 to 0.99. The chromatogram showing the corresponding peaks of OCPs in standard solution is presented in Figure 1. The detected LOD values were ranged from 0.012 to 0.032 µg/L and 0.013 to 0.39 µg/kg for river water and fish tissue respectively. Obtained LOQ values were ranged from 0.42 to 0.87 and 0.39 to 0.88 µg/kg for river water and fish tissue respectively. The retention time, LOD and LOQ values of each pesticide are presented in Table 1.

Figure 1: Chromatogram showing peaks of Organochlorine pesticide standards. A total of 19 pesticides were analyzed with different retention times (RT) for each pesticides.

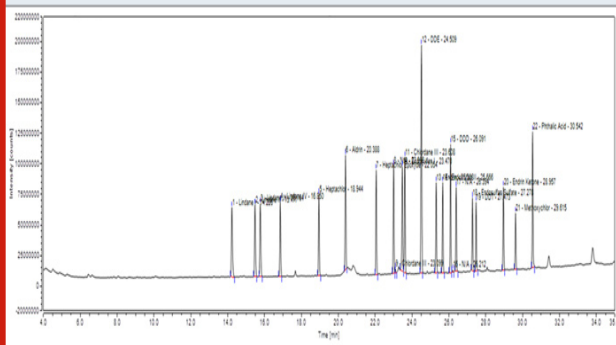


Table 2. OCPs residue levels ($\mu\text{g/L} \pm \text{SD}$) detected from Buriganga, Turag, Balu, Sitalakhya, Bangshai and Dhaleswari river water.

OCPs	Blank	Buriganga	Turag	Balu	Sitalakhya	Bangshai	Dhaleswari
Aldrin	ND	0.834 \pm 0.056	0.453 \pm 0.076	0.235 \pm 0.089	0.482 \pm 0.072	0.743 \pm 0.043	0.241 \pm 0.053
p,p-DDE	ND	1.782 \pm 0.021	1.267 \pm 0.098	1.0231 \pm 0.078	0.882 \pm 0.065	0.899 \pm 0.034	0.832 \pm 0.045
Eldrine ketone	ND	0.273 \pm 0.032	0.123 \pm 0.072	0.032 \pm 0.092	0.098 \pm 0.087	0.167 \pm 0.043	0.0743 \pm 0.032
p,p-DDD	ND	1.421 \pm 0.098	1.236 \pm 0.054	1.007 \pm 0.033	0.856 \pm 0.075	0.994 \pm 0.033	ND
Endrin	ND	1.12 \pm 0.068	1.09 \pm 0.032	0.989 \pm 0.034	0.776 \pm 0.071	0.907 \pm 0.072	0.997 \pm 0.0632
α -Endosulfan	ND	0.123 \pm 0.021	ND	0.078 \pm 0.032	0.0945 \pm 0.023	0.098 \pm 0.032	0.0743 \pm 0.021
Cis-Chordane	ND	ND	ND	ND	ND	ND	ND
Heptachlorepoxyde	ND	0.321 \pm 0.082	0.231 \pm 0.067	0.165 \pm 0.082	0.220 \pm 0.065	0.182 \pm 0.098	0.098 \pm 0.034
Heptachlor	ND	ND	ND	ND	ND	ND	ND
p,p-DDT	ND	1.243 \pm 0.043	1.076 \pm 0.021	1.243 \pm 0.078	0.605 \pm 0.066	0.882 \pm 0.023	0.993 \pm 0.082
Methoxychlor	ND	ND	ND	ND	ND	ND	ND
Endosulfan sulfate	ND	0.023 \pm 0.078	ND	ND	ND	ND	ND
β -Endosulfan	ND	0.242 \pm 0.098	0.175 \pm 0.078	0.08 \pm 0.023	ND	ND	ND
Lindane-I	ND	ND	ND	ND	ND	ND	ND
Lindane-II	ND	ND	ND	ND	ND	ND	ND
Lindane -III	ND	ND	ND	ND	ND	ND	ND
Lindane-IV	ND	ND	ND	ND	ND	ND	ND
Trans-Chordane	ND	ND	ND	ND	ND	ND	ND
Phthalic Acid	ND	ND	ND	ND	ND	ND	ND

*ND: Not detected

Table 3. OCPs residue levels ($\mu\text{g/kg} \pm \text{SD}$) in fish samples from Buriganga, Turag, Balu, Sitalakhya, Bangshai and Dhaleswari rivers.

OCPs	Buriganga			Turag			Balu			Sitalakhya			Bangshai			Dhaleswari			
	A. microl epist	B. gonio notus	B. ten ga na	A. microl epist	B. gonio notus	B. ten ga na	A. microl epist	B. gon ion otu s	B. ten gan a	A. microl epist	B. gonio notus	B. ten ga na	A. microl epist	B. gonio notus	B. ten ga na	A. microl epist	B. gon ion otu s	B. ten ga na	
Aldrin	0.4 32 \pm 0. 03 2	0.3 42 \pm 0. 02 3	ND	0.2 34 \pm 0.0 54	0.2 31 \pm 0.0 64	0.2 36 \pm 0.0 67	0.1 13 \pm 0.0 43	0.1 12 \pm 0.0 67	ND	0.1 17 \pm 0.0 2	ND	0.2 12 \pm 0.0 84	0.2 22 \pm 0. 04 5	0.2 34 \pm 0. 06 7	ND	ND	ND	ND	ND
p,p-DDE	0.9 21 \pm 0. 02 1	0.8 83 \pm 0. 03 4	0.9 87 \pm 0. 02 3	0.8 83 \pm 0. 04 3	0.8 76 \pm 0. 03 2	0.6 68 \pm 0. 02 3	0.9 91 \pm 0. 02 1	0.5 431 \pm 0. 028	0.4 321 \pm 0. 067	0.6 75 \pm 0. 03 2	0.4 32 \pm 0. 05 4	0.4 53 \pm 0. 03 6	0.4 56 \pm 0. 02 3	0.1 18 \pm 0. 05 5	0.5 56 \pm 0. 02 4	0.3 42 \pm 0. 02 4	0.2 67 \pm 0.0 68	0.1 21 \pm 0. 07 9	
Eldrine ketone	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	

Eldrine ketone	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
p,p-DDD	0.87 ±0.06 7	0.785 ±0.08 6	0.943 ±0.08 8	0.786 ±0.03 4	0.675 ±0.04 3	0.987 ±0.08 9	0.883 ±0.04 5	0.885± 0.087	0.342± 0.021	0.386 ±0.04 7	0.554 ±0.03 4	0.432 ±0.03 5	0.118 ±0.02 9	0.564 ±0.08 5	0.776 ±0.04 7	ND	ND	ND	
Endrin	0.232 ±0.08 8	0.156 ±0.08 2	0.271 ±0.07 6	0.397 ±0.03 3	0.143 ±0.02 2	0.376 ±0.01 2	0.145 ±0.02 3	0.253± 0.056	0.112± 0.054	0.153 ±0.04 2	0.115 ±0.03 4	0.275 ±0.08 0	0.364 ±0.06 8	0.223 ±0.08 5	0.142 ±0.06 3	0.143 ±0.08 7	0.143 ±0.08 5	0.198± 0.0543	0.209 ±0.08 9
α-Endosulfan	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Cis-Chorane	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Heptachloroepoxide	0.123 ±0.053	0.231 ±0.021	0.111 ±0.043	0.113 ±0.076	0.110 ±0.032	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Heptachlor	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
OCPs	Buriganga			Turag			Balu			Sitalakhya			Bangshai			Dhaleswari			
p,p-DDT	0.584 ±0.02 2	0.678 ±0.02 1	0.384 ±0.03 2	0.287 ±0.01 1	0.564 ±0.03 4	0.484 ±0.03 7	0.587 ±0.06 7	0.398± 0.027	ND	0.334 ±0.02 5	0.467 ±0.03 9	0.256 ±0.03 0	0.432 ±0.08 7	0.223 ±0.06 8	0.253 ±0.05 7	0.321 ±0.07 3	0.221 ±0.07 8	0.232± 0.078	0.353 ±0.09 3
Methoxychlor	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Endosulfan sulfate	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
β-Endosulfan	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Lindane-I	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Lindane-II	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Lindane-III	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Lindane-IV	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Trans-Chorane	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Phthalic Acid	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND

The OCPs concentration in fishes from these rivers is presented in Table 3. The average OCPs concentration (mg/kg) in fishes from those rivers is shown in Table 4. Endrin normally stored in the fat tissues and may act as neurotoxin affecting the central nervous system leading to convulsions, seizures, or even death. Although, Endrin is not considered

as mutagen and it has no carcinogenic activity but still endrin is a toxic compound and it may cause serious health problems. Aldrin is a neurotoxic pesticide. It can elevate the neurotoxicity. Aldrin can stimulate the central nervous system (CNS) leading to hyperexcitation and seizures. Consumption of fishes from those rivers may be health

hazardous as HI values of some of the OCPs are above the safety range.

HI values of some of the OCPs are within the range of safety limit indicated that, consumers may not suffer from health risks from those pesticides but, long time exposure of those OCPs may accumulate in the fat tissues and can be hazardous. In addition, there have several

other sources (vegetables, milk, eggs, meat and fruits) from where consumers can exposure OCPs. So, Health conscious peoples should very careful while eating fishes from the rivers contaminated with OCPs. Our current study estimated the level of OCPs in river water and fishes from six contaminated rivers surrounding Dhaka city but, more studies are required to ensure the safety of peoples (Lobato et al. 2021).

Table 4. Mean values of detected OCPs residues in fish tissues.

OCPs	Mean (mg/kg)					
	Buriganga	Turag	Balu	Sitalakhya	Bangshai	Dhalewari
Aldrin	0.000258	0.000233667	0.000075	0.000109667	0.000152	0
p,p-DDE	0.000930333	0.000809	0.0006554	0.00052	0.000376667	0.000243333
p,p-DDD	0.000871667	0.000816	0.000703333	0.000457333	0.000486	0
Endrin	0.000219667	0.000305333	0.00017	6.26667E-05	0.000243	0.000183333
Heptachlorepoide	0.000155	4.95556E-05	0	0	0	0
p,p-DDT	0.000548667	0.000445	0.000328333	0.000352333	0.000302667	0.000302

Table 5. EDI values calculated from the OCPs from the fish samples.

OCPs	EDI						ADI
	Buriganga	Turag	Balu	Sitalakhya	Bangshai	Dhalewari	
Aldrin	0.00017	0.00015	4.9E-05	7.2E-05	1E-04	0	0.0001
p,p-DDE	0.00061	0.00053	0.00043	0.00034	0.00025	0.00016	0.001
p,p-DDD	0.00057	0.00054	0.00046	0.0003	0.00032	0	0.001
Endrin	0.00014	0.0002	0.00011	4.1E-05	0.00016	0.000121	0.0001
Heptachlor epoxide	0.0001	3.3E-05	0	0	0	0	0.0001
p,p-DDT	0.00036	0.00029	0.00022	0.00023	0.0002	0.000199	0.01

*ADI= Acceptable daily intake.

Table 6. Health index (HI) values of detected OCPs in the fishes of six contaminated rivers.

OCPs	HI					
	Buriganga	Turag	Balu	Sitalakhya	Bangshai	Dhalewari
Aldrin	1.696423714	1.536425095	0.493146429	0.721089667	0.999443429	0
p,p-DDE	0.611720748	0.531940614	0.430944226	0.341914857	0.247669095	0.159998619
p,p-DDD	0.573145738	0.536543314	0.462461762	0.300709733	0.319558886	0
Endrin	1.444371095	2.007653905	1.117798571	0.412051238	1.597794429	1.205469048
Heptachlor epoxide	1.019169286	0.325841937	0	0	0	0
p,p-DDT	0.036076401	0.029260021	0.021588855	0.023166923	0.019901198	0.019857363

CONCLUSION

The findings of the present study indicated OCPs concentrations in six contaminated river water and fishes, and assessed the possible human health risks through consumption of those fishes. This study will help to figure out the presence of pesticide residues in the contaminated

rivers surrounding Dhaka city in very small concentration. Consumers' health relies on food safety which is ensured by a safe environment. As these rivers are polluting day by day so, the responsible authorities should monitor the presence of pesticide residues on regular basis for public health safety.

REFERENCES

- Aktar, W., Sengupta, D. and Chowdhury, A. (2009). Impact of pesticides use in agriculture: Their benefits and hazards, *Interdisciplinary Toxicology*, 2(1), pp. 1–12. doi: 10.2478/v10102-009-0001-7.
- American Public Health Association (2002). American Water Works Association; Water Environment Federation, Standard Methods for the Examination of Water and Wastewater, 02, pp. 1–541.
- Arikan, K. and Turan, S.L. (2020). Organochlorine pesticide residues in feathers of four bird species from western part of Turkey, *Turkish Journal of Zoology*, 44(5), pp.401-407.
- Byrne, S., Miller, P., Waghiyi, V., et al. (2015). Persistent organochlorine pesticide exposure related to a formerly used defense site on St. Lawrence Island, Alaska: data from sentinel fish and human sera, *Journal of Toxicology and Environmental Health, Part A*, 78(15), pp.976-992. doi: 10.1080/15287394.2015.1037412.
- Capkin, E., Altinok, I. and Karahan, S. (2006). Water quality and fish size affect toxicity of endosulfan, an organochlorine pesticide, to rainbow trout, *Chemosphere*, 64(10), pp. 1793–1800. doi: 10.1016/j.chemosphere.2005.12.050.
- Carneiro, R.P., Oliveira, F.A., Madureira, F.D., et al. (2013). Development and method validation for determination of 128 pesticides in bananas by modified QuEChERS and UHPLC–MS/MS analysis, *Food Control*, 33(2), pp.413-423. doi: 10.1016/j.foodcont.2013.02.027.
- Castillo, L.E., Martínez, E., Ruedert, C., et al. (2006) Water quality and macroinvertebrate community response following pesticide applications in a banana plantation, Limon, Costa Rica, *Science of the total environment*, 367(1), pp.418-432. doi: 10.1016/j.scitotenv.2006.02.052.
- Cui, L., Ge, J., Zhu, Y., et al. (2015). Concentrations, bioaccumulation, and human health risk assessment of organochlorine pesticides and heavy metals in edible fish from Wuhan, China, *Environmental Science and Pollution Research*, 22(20), pp.15866-15879. doi: 10.1007/s11356-015-4752-8.
- Cui, Y., Ke, R., Gao, W., et al. (2020). Analysis of Organochlorine Pesticide Residues in Various Vegetable Oils Collected in Chinese Markets, *Journal of Agricultural and Food Chemistry*, 68(49), pp.14594-14602. <https://doi.org/10.1021/acs.jafc.0c05227>
- DeLorenzo, M. E., Scott, G. I. and Ross, P. E. (2001). Toxicity of pesticides to aquatic microorganisms: A review, *Environmental Toxicology and Chemistry*, 20(1), pp. 84–98. doi: 10.1002/etc.5620200108.
- Diel, P., Olf, S., Schmidt, S. et al. (2002). Effects of the environmental estrogens bisphenol A, o, p'-DDT, p-tert-octylphenol and coumestrol on apoptosis induction, cell proliferation and the expression of estrogen sensitive molecular parameters in the human breast cancer cell line MCF-7, *The Journal of steroid biochemistry and molecular biology*, 80(1), pp.61-70. doi: 10.1016/S0960-0760(01)00173-X.
- Enan, E. and Matsumura, F. (1998). Activation of c-Neu Tyrosine Kinase by o,p'-DDT and β-HCH in Cell-Free and Intact Cell Preparations from MCF-7 Human Breast Cancer Cells, *Journal of Biochemical and Molecular Toxicology*, 12(2), pp. 83–92. doi: 10.1002/(SICI)1099-0461(1998)12:2<83::AID-JBT3>3.0.CO;2-K.
- Ennaceur, S., Ridha, D. and Marcos, R. (2008). Genotoxicity of the organochlorine pesticides 1,1-dichloro-2,2- bis(p-chlorophenyl)ethylene (DDE) and hexachlorobenzene (HCB) in cultured human lymphocytes, *Chemosphere*, 71(7), pp. 1335–1339. doi: 10.1016/j.chemosphere.2007.11.040.
- Forum, R. A. and Agency, U. S. E. P. (1986). Guidelines for the Health Risk Assessment of Chemical Mixtures, *Epa*, 51(September), pp. 34014–34025.
- Frankart, C., Eullaffroy, P. and Vernet, G. (2003). Comparative effects of four herbicides on non-photochemical fluorescence quenching in *Lemna minor*, *Environmental and Experimental Botany*, 49(2), pp. 159–168. doi: 10.1016/S0098-8472(02)00067-9.
- Fry, D. M. (1995). Reproductive effects in birds exposed to pesticides and industrial chemicals, *Environmental Health Perspectives*, 103(7), pp. 165–171. doi: 10.1289/ehp.95103s7165.
- Gupta, P. K. (2004). Pesticide exposure - Indian scene, *Toxicology*, 198(1–3), pp. 83–90. doi: 10.1016/j.tox.2004.01.021.
- Han, E.H., Kim, H.G., Hwang, Y.P., et al. (2010). The role of cyclooxygenase-2-dependent signaling via cyclic AMP response element activation on aromatase up-regulation by o, p'-DDT in human breast cancer cells, *Toxicology letters*, 198(3), pp.331-341. doi: 10.1016/j.toxlet.2010.07.015.
- Hwang, B.H. and Lee, M.R. (2000). Solid-phase microextraction for organochlorine pesticide residues analysis in Chinese herbal formulations, *Journal of Chromatography A*, 898(2), pp.245-256. [https://doi.org/10.1016/S0021-9673\(00\)00874-8](https://doi.org/10.1016/S0021-9673(00)00874-8)
- Hwang, S.M., Lee, H.U., Kim, J.B. et al. (2020). Validation of analytical methods for organochlorine pesticide detection in shellfish and cephalopods by GC–MS/MS, *Food Science and Biotechnology*, 29(8), pp.1053-1062. <https://doi.org/10.1007/s10068-020-00748-0>
- Lee, H.K., Kim, K., Lee, J., et al. (2020). Targeted toxicometabolomics of endosulfan sulfate in adult zebrafish (*Danio rerio*) using GC-MS/MS in multiple reaction monitoring mode, *Journal of hazardous materials*, 389, p.122056. <https://doi.org/10.1016/j.jhazmat.2020.122056>

- Li, W., Zhang, Z.M., Zhang, R.R., et al. (2020). Effective removal matrix interferences by a modified QuEChERS based on the molecularly imprinted polymers for determination of 84 polychlorinated biphenyls and organochlorine pesticides in shellfish samples, *Journal of hazardous materials*, 384, p.121241. <https://doi.org/10.1016/j.jhazmat.2019.121241>
- Lobato, A., Fernandes, V.C., Pacheco, J.G., et al. (2021). Organochlorine pesticide analysis in milk by gas-diffusion microextraction with gas chromatography-electron capture detection and confirmation by mass spectrometry, *Journal of Chromatography A*, 1636, p.461797. <https://doi.org/10.1016/j.chroma.2020.461797>
- Ndebele, K., Graham, B. and Tchounwou, P. B. (2010). Estrogenic activity of coumestrol, DDT, and TCDD in human cervical cancer cells, *International Journal of Environmental Research and Public Health*, 7(5), pp. 2045–2056. doi: 10.3390/ijerph7052045.
- Olisah, C., Okoh, O.O. and Okoh, A.I. (2020). Occurrence of organochlorine pesticide residues in biological and environmental matrices in Africa: A two-decade review, *Heliyon*, 6(3), p.e03518. <https://doi.org/10.1016/j.heliyon.2020.e03518>
- Pico, Y., Alfarhan, A.H. and Barcelo, D. (2020). How recent innovations in gas chromatography-mass spectrometry have improved pesticide residue determination: An alternative technique to be in your radar, *TrAC Trends in Analytical Chemistry*, 122, p.115720. <https://doi.org/10.1016/j.trac.2019.115720>
- Rashed, M. N. (2001). Monitoring of environmental heavy metals in fish from nasser lake, *Environment International*, 27(1), pp. 27–33. doi: 10.1016/S0160-4120(01)00050-2.
- Singh, S.P., Sharma, J. and Prakash, P. (2020). Development of a Gas Chromatography Tandem Mass Spectrometry-Based Analytical Method for the Quantitative Determination of Organochlorine Pesticide Residues in Potato Crops, *Analytical Chemistry Letters*, 10(6), pp.811-826. <https://doi.org/10.1080/22297928.2021.1875872>
- Tahboub, Y.R., Zaater, M.F. and Barri, T.A. (2006). Simultaneous identification and quantitation of selected organochlorine pesticide residues in honey by full-scan gas chromatography–mass spectrometry, *Analytica Chimica Acta*, 558(1-2), pp.62-68. <https://doi.org/10.1016/j.aca.2005.11.004>
- Wang, J., Liang, W., Henkelmann, B., et al. (2015). Organochlorine pesticides accumulated by SPMD-based virtual organisms and feral fish in Three Gorges Reservoir, China, *Environmental pollution*, 202, pp.160-167. doi: 10.1016/j.envpol.2015.03.031.
- Wong, L.I., Labrecque, M.P., Ibuki, N., et al. (2015). p, p'-Dichlorodiphenyltrichloroethane (p, p'-DDT) and p, p'-dichlorodiphenyldichloroethylene (p, p'-DDE) repress prostate specific antigen levels in human prostate cancer cell lines, *Chemico-biological interactions*, 230, pp.40-49. doi: 10.1016/j.cbi.2015.02.002.
- Wu, P., Gu, M., Wang, Y., et al. (2020). Transfer of organochlorine pesticide residues during household and industrial processing of ginseng, *Journal of Food Quality*, 2020. <https://doi.org/10.1155/2020/5946078>
- Ye, X., Shao, H., Zhou, T., et al. (2020). Analysis of Organochlorine Pesticides in Tomatoes Using a Modified QuEChERS Method Based on N-Doped Graphitized Carbon Coupled with GC-MS/MS, *Food Analytical Methods*, 13(3), pp.823-832. <https://doi.org/10.1007/s12161-019-01674-6>
- Yohannes, Y.B., Ikenaka, Y., Nakayama, S.M. et al. (2014). Organochlorine pesticides in bird species and their prey (fish) from the Ethiopian Rift Valley region, Ethiopia, *Environmental pollution*, 192, pp.121-128. doi: 10.1016/j.envpol.2014.05.007.
- Yohannes, Y.B., Ikenaka, Y., Saengtienchai, A., et al. (2014). Concentrations and human health risk assessment of organochlorine pesticides in edible fish species from a Rift Valley lake—Lake Ziway, Ethiopia, *Ecotoxicology and environmental safety*, 106, pp.95-101. doi: 10.1016/j.ecoenv.2014.04.014.
- Zhang, Y., Lin, N., Su, S., et al. (2014). Freeze drying reduces the extractability of organochlorine pesticides in fish muscle tissue by microwave-assisted method, *Environmental pollution*, 191, pp.250-252. doi: 10.1016/j.envpol.2014.04.018.