

Extraction and Characterization of Protein Hydrolysate and Trypsin from Fish viscera of Labeo rohita

Aurobido Das¹, Yashaswi Nayak^{2*} and Supriya Dash³

^{1,2}Department of Zoology, Centurion University of Technology and Management, Odisha, India ³Department of Biotechnology, College of Engineering and Technology, Odisha, India

ABSTRACT

Acidic and antacid proteases from instinctive misuse of *Labeo rohita* (Hamilton et. al., 1822) were confined, incompletely sanitized by ammonium sulfate precipitation followed by dialysis, their energy and attributes considered. The purging fold expanded from 1.24 to 2.49 and 1.19 to 1.55 in acidic and soluble protease individually along with the cleaning steps. The atomic weight was found in the scope of 15-35 kDa and 25-63 kDa individually in acidic and basic proteases. The pH and temperature optima for acidic and basic proteases were 3 and 10, at 40°C and 60°C individually. The Protease action was diminished by 40% and 60% when hatched at 90°C for 30 min. Both the proteases demonstrated a diminished movement of over half after brooding with NaCl centralization of 0.5%. The level of hydrolysis (DH) of the proteases on muscle protein expanded with an increment of chemical fixations. Both soybean trypsin inhibitor and EDTA displayed a high level of hindrance when proteases were hatched with 50 mM of both the inhibitors. The investigation demonstrated that proteases from Rohu instinctive misuse could discover use in applications where the greatest movement at moderate temperature and low NaCl fixation is wanted.

KEY WORDS: ACIDIC AND ANTACID, AMMONIUM SULFATE PRECIPITATION, LABEO ROHITA

INTRODUCTION

Fish preparing tasks produce more than 60 % of the crude material. In a non-industrial nation like India, these squander are arranged or changed over into creature feed, fish feast, and manure. This training prompts underutilization of crude material and may influence the manageable usage of accessible assets. The removal of fish handling waste is under exacting guidelines because of natural issues and it adds to the operational expense of the fish industry (Elavarasan et. al., 2016).Hence, the successful use of fish preparing waste is picking up significance. A rough amount of waste created during

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NAAS Journal Score 2020 (4.31) A Society of Science and Nature Publication, Bhopal India 2020. All rights reserved. Online Contents Available at: http://www.bbrc.in/ the preparation of a significant sort of fish items is introduced in Table.1. There is no verified information on the side-effect age from the Indian fish handling area. The measure of waste created will differ with the size, style of item and species, nature of taking care of (machine/manual taking care of, the aptitude of working/ taking care of individual). The significant squander from shellfish is shell squander which is used somewhat as a crude material in the chitin industry. Squanders from balance fishes are containing an impressive amount of proteins which can be changed over/recuperated into protein hydrolysates for improved usage. From the fish protein hydrolysate industry perspective, the amount of blade fish squanders/side-effects are more significant.

An enormous amount of instinctive squanders is produced in the retail fish advertises due to pre-preparing. Such organic squanders, if not used something else, would represent an issue of their removal and ensuing natural contamination. Fish preparing squanders is about 30% of the entire fish and contained head, scales, skins, and viscera (Klomklao et. al., 2006) and is considered as a



phenomenal wellspring of protein and bio-dynamic peptides (Arnesen et. al., 2007). Even though there is a degree to recoup proteins and chemicals from the instinctive squanders of fish, however,a colossal amount of such waste is disposed of with no such endeavor (Bhaskar and Mahendrakar, 2007). As indicated by (Bezerra et. al.,2005), the fish instinctive waste generally represents 5% of the complete mass and incorporates stomach, pyloric caeca, digestion tracts, liver, pancreas, etc and different organs like spleen and balls. The stomach related chemicals from the fish instinctive waste are exceptionally dynamic over a wide scope of pH and temperature conditions and consequently speak to a significant esteemed side-effect of the fishing industry (Castillo-Yanez et. al., 2004).

Among the hydrolytic proteins, proteases speak to a significant class of mechanical compounds; have been utilized in various applications, generally in food, cleanser, material, calfskin, and pharmaceutics just as in squandering the executives and bioremediation measure (Anwar et. al., 1998)(Gupta et. al., 2002). Notwithstanding, proteases require their purging and portrayal before any application. Proteases contribute about 60% of the world's all out catalyst creation and utilized around the world (Gupta et. al., 2002). As of now, the greater part of the proteolytic proteins are separated from microscopic organisms, and generally, barely any endeavors have been made on the application of fish proteases as mechanical handling helps. Normally, the fishery results are commonly utilized as feeds and manures. As of late, intrigue has developed to look through high-esteem useful biomolecules from the fishery squanders, prominently catalysts. All things considered, a few specialists researched proteases from the instinctive squander from marine fish (Nasri et. al., 2011). However, the portraval of fish proteases particularly from the instinctive squanders of freshwater fish is only occasionally revealed.

In light of the above reasoning, the current examination was done to describe mostly purged acidic and soluble proteases from the fish instinctive squanders for deciding their application in food handling activities just as to decrease garbage removal issues. Rohu (Labeo rohita), overwhelmingly a section feeder and feeds basically on filamentous green growth, disintegrated vegetation, and mud was chosen for concentrate as it is the most regularly devoured freshwater fish in India among the carps.

1.2. Worldwide protein market: Worldwide protein fixing market examination uncovered that 43.2 % of incomes in the worldwide wellbeing fixings market contributed from protein. Europe holds its enormous lead in the worldwide protein market. Key items under creature protein fixings incorporate, gelatin, collagen, egg, and dairy. There is a steady interest in creature protein fixings. The worldwide protein market is required to arrive at the estimation of US \$ 40.88 Billion (2.7843618 trillion Indian rupees) by 2022. The utilization of protein fixings in newborn child equation diminished protein insufficiencies.

The utilization of protein fixings in the drug and the restorative industry is expanding continuously.Global protein fixing a piece of the pie is reasonably solidified with DuPont, Bunge, ADM, Cargill, and Mead Johnson being the significant business players.

Raw material accessibility in China and India impact industry players to move to fabricate base in the region. The protein supplements market in India is developing at 6% and is presently esteemed at Rs. 252 crores every year. Fish protein hydrolysate Fish protein hydrolysate is an item set up from proteins sourced from fish meat/ fish handling by items using enzymatic or synthetic cycle. Enzymatically created hydrolysates are broadly acknowledged which contain a combination of peptides of changing sizes and free amino acids.

1.3. Cycle for creation of protein hydrolysate: Protein hydrolysates from fish handling dispose of can be readied utilizing four distinctive cycles in the particular corrosive cycle, antacid cycle, enzymatic cycle, and microbial maturation. The fundamental component of enzymatic hydrolysis and the impact of various elements is examined underneath.



1.4. Enzymatic cycle: Enzymatic hydrolysis of the fishery by items uses either autolysis measure or by adding exogenous protein. Autolysis measure includes brooding ground fishery squander at ideal response states of endogenous chemicals and utilizes the fish instinctive waste (Kristinsson et. al., 2000). The endogenous catalysts trigger the stalling of biomolecules to more modest peptides through autolysis measures. The autolysis is generally directed at impartial or somewhat basic pH, abusing the presence of serine protease of the digestive tract in basic or the carboxyl favorable to a prod of gastric juice in acidic pH (Pastorizaet et. al., 2004).

Proteolysis is the enzymatic hydrolysis of the amide bond in peptides and proteins. The compounds are misused to perform wanted capacities in handling and examination 186 and to encourage the changes of crude materials into top-notch more alluring staples (Richardson et. al., 1984). Compounds utilized in the food business and exploration are dominatingly hydrolases. Proteolytic proteins are financially the main gathering of compounds and their utilization is entrenched in the food business (Godfrey et. al., 1983). The utilization of proteases in the planning of fish protein hydrolysates has gotten a wide consideration among scientists as it is more efficient and simple of cycle control. The idea of catalysts, substrate, and hydrolysis will decide the properties of (Fish Protein Hydrolysate) FPH.

The overall streamline for the creation of FPH by the utilization of chemicals is portrayed in Fig 1. The cycle includes the homogenization of fish meat or fish squanders with the expansion of water. The homogenate is brought to the ideal temperature and pH. The hydrolysis is started by the expansion of protein at the wanted focus. After a specific length of hatching, the hydrolysis is ended by applying heat or by changing the pH. The dissolvable part in the wake of eliminating the unhydrolyzed partition is concentrated by freeze-drying/ stove drying/shower drying. The dried protein powder alludesto protein hydrolysate.

- To deliver the FPH with various and wanted properties, it is critical to know the component of protein hydrolysis. A few proteases specially catalyze the hydrolysis of bonds nearby a specific amino corrosive build-up, while some are less explicit. The catalysis by proteases happens basically as three continuous responses (Krsitinsson et. al., 2000): the development of complex between the first peptide chain and the catalyst alluded as the Michaelis complex
- Cleavage of the peptide cling to free one of the two peptides
- Nucleophilic assault on the remaining parts of the complex to separate the other peptide and to reconstitute the free catalyst

The hydrolysis of peptide bonds prompts an expansion in the quantities of ionizable gatherings (NH3+ and COO-), with an associative expansion in hydrophobicity and net charge, the decline in the atomic size of the polypeptide chain, and an adjustment of the sub-atomic structure prompting the introduction of the covered hydrophobic deposits to the fluid climate (Phillips et. al., 198)(Kester et. al., 1984)(Mahmoud et. al., 1992). Endless supply of catalyst to the proteins, the chemical substrate complex will be framed. This complex alludestoMichaelis complex which may separate back to reactant substrate and free compound, or to free chemical and item atoms (Adler et. al., 1986). The by and large acknowledged component for proteases shows that the second step that is the separation of compound substrate complex into free protein and item is the rate-deciding advance, which decides the general pace of response. Enzymatic hydrolysis of proteins is a mind-boggling measure as a result of a few peptide bonds and their particular openness to enzymatic responses (Linder et. al., 1995).

The particularity of proteins isn't the main factor that influences the peptide profile of the eventual outcome and factors, for example, temperature and pH assume a significant job. The temperature and pH can extraordinarily influence the compound response energy and their effect is diverse for every chemical. By and large, there is an ideal mix of both pH and temperature, where the compound is generally dynamic. Temperature and pH limits deactivate the compounds by denaturing them. The elements engaged with the hydrolysis of proteins are most significant both as far as energy and nature of the finished result. The main components impacting the properties of FPH are the nature of the substrate, nature of protease, and level of hydrolysis (DH) and drying strategy.

1.5. Protein hydrolysates from various fish handling: Various squanders created during fish preparation like head, skin, roe, outline waste and bone have been utilized to deliver the hydrolysate. Then again, the proteins confined from the waste parts can likewise be utilized for this reason. The protein content in various fish squanders parts are introduced in Table 1. A large portion of the investigations has been completed regarding the hydrolysis cycle and their bioactive and practical properties.

Figure 2: Shows the Preparation of Fish Protein



Table 1: Shows Protein Content In Significant Fish WasteParts

Waste Parts	Protein (%)
Head	11%-13%
Spine/outline	10-15
Cutoffs	12-22
Skin	8-12
Milt	14-27
Viscera	9-23

Logical investigations have been accounted for the planning of fish protein hydrolysate from the fish head, viscera, roe, skin, edge, and bone. The greater part of these examinations has zeroed in on their cancer prevention agent properties and different cell reinforcement peptide particles have been separated and portraved. (Chalamaih et. al., 2012) has thoroughly evaluated the protein hydrolysates from different pieces of fish squander. The fish head is a significant fishery squander contains gills. Eyes, head casing, and shoulder muscle. It is hard to recuperate the protein because of its basic unpredictability. The enzymatic cycle will solubilize the protein by changing over into peptide shapes at that point encourage the simple recuperation of proteins. Protein hydrolysates from fish head side-effect squander have been set up from different species. The significant protein present in the fish head is collagen.

Thus the peptides produced will have ordinarily the arrangement from collagen which are known for their enemy of joint pain and hostile to corpulence properties. Fish skin is again a rich wellspring of collagen. Endeavors have been made to create the hydrolysate either legitimately from the fish skin or in the wake of disconnecting the collagen or gelatin. The fish liver is another result that generally goes for oil and dinner creation. The fish liver has been utilized to set up the hydrolysateusing protamex, flavorzyme, alacalase, and neutrase (Je et. al., 2009)(Ahn et. al., 2010). Fish viscera is also a potential source of protein that can fill in as a crude material for the readiness ofprotein hydrolysates. Normally, instinctive waste protein hydrolysate may exhibitunique properties. As of late, numerous attempts have been performed for the use of fish instinctive waste for protein hydrolysates creation (Batista et. al., 2010).

Fish roe contains a considerableamount of protein. To use this underutilized protein source from fish roe, protein hydrolysates have been readied. For instance, roe protein hydrolysate from Cirrhinusmrigalausing alcalase andpapain has been reported(Chalamaiah et. al., 2010). Fishbone, which is isolated after the evacuation of muscle proteins on the casing, is another important source in distinguishing well-being advancing parts. The natural segment of fishbone, which represents 30% of the material, is made out of collagen. Consequently, fishbone is considered as a hotspot for protein hydrolysate particularly collagen peptides and gelatin hydrolysates (Kim et. al., 2006).

1.7. Use of fish protein hydrolysates Nutritional application: The proximate organization of fish protein hydrolysate would shift with the crude material (head, bone, skin, viscera), sort of cycle, kind of drying, degree of hydrolysis, and some other pre-treatment of crude material. The substance structure of food materials has a significant part in human wellbeing in gracefully of basic supplements for keeping up prosperous wellbeing. The compound arrangement of fish protein hydrolysates is significant from sustenance point of view of human wellbeing.

Amino corrosive creation of protein hydrolysates from various crude material delivered utilizing diverse chemical source under various hydrolysis conditions expected to have variety. All in all, required fundamental amino acids are bountiful in FPH with wealth in glutamic and aspartic corrosive substances. FPH do likewise have trivial amino acids. The presence of fragrant amino corrosive in fish outline protein hydrolysates has been accounted for. Studies have unmistakably indicated that FPH from fish meat/fish waste could be an ideal wellspring of basic amino acids (Chalamaiah et. al., 2010).

Table 2: Shows Proximate SHydrolysate	Structure Of Fish Protein		
Waste Part	Protein (%)		
Moisture	< 10%		
Protein	60-90%		
Fat	<5%		
Debris	0.45-27%		

Nutraceutical applications: There are fish protein hydrolysate items/peptides explicitly showcased as wellbeing supplements in created nations (Table 3). These items are demonstrated to have an explicit well-being job other than the dietary advantage. Protein hyd5rolysates or peptides present in the hydrolysate have exhibited to have cell reinforcement, hostile to heftiness, invulnerable balance, against coagulation, hostile to microbial, anticancer and antihypertension and so forth (Elavarasan et. al., 2014)(Elavarasan et. al., 2016).

1.8. Fish protein hydrolysate as a practical ingredient: Fish protein hydrolysates are dissolvable in a wide scope of pH which is an ideal trademark assist with utilizing in a wide scope of items. Protein hydrolysates have improved water-holding, oil official, emulsifying, and frothing properties. Be that as it may, the key factor which decides the useful properties is the level of hydrolysis. When all is said in done, broad hydrolysis prompts loss of usefulness. There is a basic level of hydrolysis at which protein hydrolysates ought to be set up concerning specific capacity to be utilized as a practical fixing (Elavarasan et. al., 2016) (Gajanan et. al., 2017).

1.9. Fish protein hydrolysate as feed ingredients and different applications: Fish protein hydrolysates (FPHs) have been utilized in hydroponics to take care of to improve the development and endurance of fish. Studies have indicated that FPH has helped the development execution and immunological status of many culture species. The amino corrosive arrangement and the peptides present in hydrolysate are liable for the improved development and immunological status. FPH is additionally being utilized as a wellspring of protein in poultry feed detailing and in pet creature nourishments. Different applications incorporate FPH as a plant sponsor, fixing in microbiological media, and as a cryo-protectant in fish mince/surimi.

Table 3. Shows Monetarily Advertised Fish Protein Hydrolysate Items As Nutraceuticals							
Item brand name	Points of interest	Nutraceutical applications	Nation				
PROTIZEN®	Delivered by enzymatic hydrolysis of white fish proteins United Kingdom	It is "disposition food" and dietary enhancement to battle against pressure and its side effects (weight disorders, work pressure, rest troubles, fixation challenges, and mind-set inconveniences).					
Amizate®	Delivered from Atlantic salmon fish proteins via autolysis	Sports sustenance (bolsters the body's muscle anabolism and metabolic recuperation).	North America				
Seacure®	Delivered by hydrolyzing profound sea white fish proteins	Dietary enhancement assists with supporting the cells in the gastrointestinal plot and direct gut capacities.	Canada & USA				
Vasotensin®	Delivered from Bonito (Sardaorientalis) by thermolysin hydrolysis	It upholds solid vascular capacity for ideal bloodstream and sound pulse levels.	Japan & USA				
LIQUAMEN®	Prepared from Molvamolva via autolysis	The dietary enhancement that helps in decreasing oxidative stress, bringing down the glycemic record and hostile to stretch.	United Kingdom				
Stabilium® 200	Prepared from Molvadypterygia via autolysis	Supports the body's reaction to stretch and offers healthful help for memory and intellectual capacity.	United Kingdom				
PEPTACE®	Delivered from Bonito (Sardaorientalis) by thermolysin hydrolysis	It brings down the circulatory strain by repressing the ACE compound.	Japan & USA				
MOLVAL®	Delivered from North Atlantic fish Molvamolva by enzymatic hydrolysis	Dietary enhancement suggested for. cholesterol equilibrium, stress control, and advances great cardiovascular wellbeing	United Kingdom				

Security of protein hydrolysates in human nourishment: By and large, food business administrators ought to guarantee the well-being of items. The wellbeing parts of any food fixing should be recorded before discharge on the lookout. Protein hydrolysates can be considered as protected when they are hydrolyzed from proteins having a past filled with alright for utilization and they are created utilizing proteases that are of food-grade and utilized regular food-handling techniques. The wellbeing of parts and bioactive peptides, gotten from safe hydrolysates, ought to be assessed by the assembling before the market presentation. A survey of the wellbeing evaluation of the organization by an outside free council and resulting endorsement by the skilled specialists as per novel food methodology is fundamental when the wellspring of protein and cycle is novel and under strange high admission of amino acids (Schaafsma et. al., 2009).

MATERIAL AND METHODS

2.1. Rohu viscera: Viscera of Rohu was gathered in polyethylene packs and shipped with ice. In the research

center, viscera was washed with chilled water to eliminate the disciple blood, sludges, and soils, kept in plastic packs, and put away at – 20°C until utilized for chemical extraction.

2.2. Planning of unrefined acidic and soluble protease: The technique recommended by (Vannabun et. al., 2014) was followed for the readiness of unrefined acidic and antacid proteases. At first, the instinctive mass was defrosted and homogenization was accomplished for 2 min with various extraction supports, for example, citrate cushion (10mM Citrate/HCl pH 3.0) for corrosive protease and tris cradle (10mM Tris–HCl pH 8.0, 10mM CaCl2) for basic protease, in the proportion of 1:5(w/v). The homogenization, the pellet was disposed of to gather the supernatant which was utilized as "crude chemical extract".

2.3. Enzyme cleansing: The rough chemical concentrate was exposed to two-venture (NH4)2SO4 precipitation. According to primer measure, (NH4)2SO4 grouping of 40-60% gave the most noteworthy purging fold and

explicit action. The rough chemical was encouraged with a 40–60% immersion of ammonium sulfate and afterward permitted to agree to 24h at 4°C. The supernatant was disposed of and the accelerate was broken up in 0.02 M acetic acid derivation cushion, pH 3.0 and 0.02 M Tris–HCl support, pH 8.0 for acidic and soluble proteases individually, by centrifugation at 10,000×g for 30 min at 4°C. The protein consequently acquired was dialyzed against similar support for 24 h at 4°C with the discontinuous difference in the cradle after 12 h. After dialysis, the unrefined chemical was alluded to as "partially filtered proteases".

2.4. Assurance of atomic weight: The atomic weight (MW) of the somewhat cleaned compound was completed by SDS-PAGE, following the technique proposed by Laemmli (1970). Example support was set up by blending 2.5 ml 0.5 M Tris-HCl (pH 6.8), 4 ml 10% SDS, 2 ml glycerol, 1 ml 1% b-mercaptoethanol, 0.03 ml 0.002% bromophenol blue and the last volume was made to 10 ml. Protein arrangements were blended at a 1:2 (v/v) proportion and bubbled for 10 min. Tests (10 µl) were stacked on the gel made of 4% stacking and 12.5 % isolating gels and fractionated for 90 min at a consistent current of 400 mA. After electrophoresis, the gels were recolored with 0.05g Coomassie splendid blue R-250 in 15% methanol and 5% acidic corrosive and destained with destaining arrangements [solution-1 (half methanol and 7.5% acidic corrosive) and arrangement 2 (5% methanol and 7.5% acidic acid)]. The atomic weight was assessed utilizing protein standard (10-245kDa) (HiMedia, India).

2.5.Protein content: The protein content was assessed following Lowry "s technique (Lowry et. al., 1951) by estimating test absorbance at 280 and 260 nm, utilizing ox-like serum egg whites as standard.

2.6.Test of protease action: The acidic protease action was resolved as recommended by (Natalia et. al., 2004) utilizing 2% cow-like hemoglobin arrangement containing 0.04M HCl (corrosive denatured) as substrate at pH 3.0 and 37°C, while, a technique for Rawdkuen et al., (2010) was followed to decide soluble protease movement utilizing casein as a substrate. The absorbance read at 280 nm and changed over into µmoles of tyrosine freed utilizing arrangements of 25-250 µg/ml centralization of tyrosine for alignment bend. Enzymatic action was communicated as one unit equal to the measure of chemical equipped for hydrolyzing ox-like hemoglobin to free 1 µmole tyrosine under standard examine conditions. All-out action and explicit action was communicated as units of enzymatic movement per ml protein (U/ml) and per mg protein (U/mg) individually.

2.7.Complete action: The complete enzymatic action was assessed utilizing the accompanying condition.

Total Activity $\left(\frac{U}{mL}\right)$

 μ mole of Tyrosine equivalent released \times Total Volume of reaction assay (mL)

Volume of Enzyme used(mL) × Time of Assay (min) × Volume used in Cuvette(mL)

2.8.Explicit action of protein: The particular action of both the proteins was resolved to utilize the condition as proposed by El-beltagy et al., (2005).

$$Specific \ Activity\left(\frac{U}{mg}\right) = \frac{Total \ activity\left(\frac{U}{mL}\right)}{Protein \ content\left(\frac{mg}{mL}\right)}$$

2.8.Protein decontamination fold: The degree of sanitization was assessed by deciding the purging fold following the condition given by El-beltagy et al., (2005).

 $Purification fold = \frac{Specific \ Activity}{Specific \ Activity \ of \ crude \ extract}$

2.9.Ideal pH and pH stability: The ideal pH for enzymatic action was resolved after the strategy for (Vannabun et. al., 2014), by testing protease action at various pH conditions utilizing 100mM cradle arrangements going from pH 1.0 to 12.0 (Glycine-HCl cushion for pH (1.0-3.0); sodium acetic acid derivation support for pH (4.0-6.0); Tris-HCl support for pH (7.0–9.0); and Glycine-NaOH support for (9.0–12.0), at the ideal temperature for action recently decided.

The impact of pH on protein security was dictated by the technique for (Vannabun et. al., 2014). The compound was hatched at different pH (1.0–12.0) utilizing various cushions of 100mM Glycine–HCl (1.0–3.0), Na-acetic acid derivation (4.0–6.0), Tris–HCl (7.0–9.0), and Glycine–NaOH (10.0–12.0) for 30 min alongside the spaces arranged all the while. The leftover enzymatic action after brooding was assessed and contrasted and the condition that indicated the most elevated worth (100% movement).

2.10.Ideal temperature and temperature stability: Protease movement at various temperatures (30–90°C) was performed by utilizing various cushions like Glycine–HCl (pH 3.0) and Tris–HCl (pH 8.0) for acidic and basic protease action separately as per the strategy given by Vannabun et al., (2014). To decide the warm steadiness of proteases, catalyst removal was brooded for different time lengths like 1,3,5,10,15,20,30,40,50, and 60 min at 90°C and the staying enzymatic movement was resolved. The control was not pre-hatched and considered as 100% action.

2.11.Impact of NaCl focus on enzyme movement: The response blend was made with various conc. of NaCl (0-2.5%, w/v) and protein was hatched trailed by the assurance of lingering action. The control was made without NaCl and its action was considered as 100% action (Vannabun et. al., 2014).

2.12.Impact of isolated enzymes on proteins hydrolysis: Separated acidic and antacid proteases were utilized to hydrolyze the groundfish muscle protein to decide the level of hydrolysis of catalyst on the fish muscle. The ground muscle (2g) was hatched with protein at various focuses (10-50 mL) for 30 minutes at 60°C. The response

was halted by adding 5 mL of 20% TCA followed by centrifugation at 3300 rpm for 10 minutes to gather the 10% TCA solvent material as the supernatant. The protein substance of the supernatant was assessed by the Biuret technique. The level of hydrolysis was dictated by the strategy (Hoyle et. al., 1994).

 $\% DH = \frac{10\% \, TCA \, soluble \, protein \, in \, the \, sample}{Total \, content \, of \, sample} \times 100$

2.13.Chemical inhibitors and activators: Chemical inhibitors, for example, soybean trypsin inhibitor (STPI) and ethylene diamine tetra acetic corrosive (EDTA) were utilized to decide their consequences for the enzymatic movement. Pre-hatching of the substrates with those at various conc. like 10, 20, 30, 40, and 50 mM were accomplished for 10 min at the ideal temperature of compound followed by an assurance of catalyst movement. The outcomes were communicated as an overall level of the movement without modifiers.

2.14. Analysis of Statistics: Investigation of change (ANOVA) trailed by Duncan's different reach test was done to decide contrasts between implies. The measurable investigation was performed utilizing the Statistical Package for Social Sciences (SPSS for Windows adaptation 16.0, SPSS, Inc., Chicago, IL).

RESULTS AND DISCUSSION

3.1.Halfway filtration of proteases: The protein content, absolute movement, explicit action, and cleaning fold for acidic and antacid proteases of instinctive misuse of Rohu is introduced in Table 4. The normal protein content was discovered to be 6.31 mg/ml and 7.79 mg/ ml in acidic and soluble rough proteases individually. After ammonium sulfate fractionation (40-60%), the protein content diminished in the unrefined proteases, and the qualities came to 3.72 mg/ml and 4.15 mg/ml in acidic and antacid rough proteases individually. The soaked ammonium sulfate arrangement specifically accelerates proteins from the rough catalyst separate by the salting-in and salting-out system to frame a mostly refined chemical concentrate (Klomklao et. al., 2006). This might be because of the pollutions present in the unrefined example which are taken out after ammonium sulfate precipitation. Dialysis, a stage in the refinement of proteases, displayed a further decrease of protein content in the (NH4)2SO4 accelerated proteases. In acidic and soluble proteases the normal protein content diminished to 1.68 mg/ml and 2.96 mg/ml separately. Such abatement of protein content after dialysis might be because of the additional evacuation of different proteins, not eliminated by ammonium sulfate fractionation.

3.2.The atomic weight of proteases: The electrophoretic example demonstrated a few clear groups showing the presence of various proteases of shifting sub-atomic mass if there should arise an occurrence of both basic and acidic protease tests (Fig. 3). If there should arise an occurrence of rough and somewhat sanitized acidic and antacid proteases 3-4 groups were watched going from

15-35 kDa and 25-63 kDa separately. A few creators revealed the sub-atomic loads of instinctive soluble and acidic proteases in the scope of 17-90 kDa. Atomic load of acidic protease from Tilapia nilotica after gel filtration on Sephadex G-100 was accounted for as 31.0 kDa (El-Beltagy et al., 2004). While, (Lopez-Liorca et. al., 1990) and (Liu et. al., 2008) revealed the sub-atomic load of the instinctive acidic protease of fish as around 32 kDa and 28.5 kDa individually.

Sub-atomic load of fish instinctive soluble proteases has been accounted for as 23.5 kDa (Bezerra et. al., 2005), 23-28 kDa (Balti et. al., 2009), 23 kDa (El-Beltagy et. al., 2004), 24-30 kDa (Sekizaki et. al., 2000). The current investigation uncovered that the atomic loads of soluble proteases are higher contrasted with acidic proteases. The presence of a few groups in the electrophoretic partition of stomach related proteases was disclosed as because of constituent chemicals like trypsin, chymotrypsin, collagenase, gastricin, pepsin, elastase, carboxypeptidase, and carboxylesterase (Barkia et. al., 2010), and due generally to the distinctive sub-atomic loads of individual catalyst. The current investigation supported the perceptions announced by before analysts with regards to atomic weight appropriation of stomach related proteases (Younes et. al., 2014)(Sila et. al., 2012).

3.3. Examine of proteolytic action: The normal all-out action of unrefined acidic and antacid Rohu viscera squander was resolved to be 18.33 U/ml and 34.11 U/ ml individually. Complete proteolytic action diminished after (NH4)2SO4 fractionation (ASF) and further decrease occurred after dialysis. If there should arise an occurrence of basic proteases the recuperation rate was discovered to be 63.67 and 58.95 after ASF and dialysis separately (Table 1). Comparative was seen if there should arise an occurrence of acidic proteases, wherein, recuperation of absolute action after ASF and dialysis were discovered to be 72.76 and 66.28 separately (Table 4). Purging may have eliminated other catheptic compounds that were likely present in the instinctive waste, and came about the decline of the absolute action. Such abatement of protease movement after sanitization was additionally detailed (Subash et. al., 2011)(Kim et. al., 2012).

The normal explicit action after dialysis was discovered to be 6.79 and 7.23 if there should be an occurrence of soluble and acidic proteases separately. Such increment of explicit action along the cleansing advances might be clarified as the evacuation of meddling proteins during (NH4)2SO4 fractionation and further during dialysis, coming about improved action. Increment of explicit action with the advancement of the refinement was likewise detailed by (Liu et. al., 2008) (Bezerra et. al., 2005) (El-Beltagy et. al., 2004). This examination additionally uncovered that the particular movement of post-dialysis acidic proteases was more than the soluble proteases, even though, the later indicated more all-out action and pre-dialysis explicit action contrasted with the acidic one. Since the Rohu being missing of the genuine stomach, maybe the explanation behind the low measure

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of acidic proteases discharge in the gut substance of viscera, as stomach establishes a significant wellspring of stomach related proteolytic catalysts (Simpson et. al., 2000).

The particular movement of the protein decides the sanitization overlay. In the two-venture purging framework, the cleansing fold encountered an expansion from stage two to stage three in the event of both acidic and basic proteases. If there should arise an occurrence of soluble proteases the outcome demonstrated that expansion in the purging fold was from 1.19 to 1.55, while, it was 1.24 to 2.49 in the event of acidic proteases. Increment of refinement crease following dialysis has likewise been accounted for by (El-Beltagy et. al., 2004) (Liu et. al., 2008). Immaculateness of the trypsin-like compound from anchovy stomach related plot was expanded by 2.7-overlap following ammonium sulfate precipitation (20-70%) (Martinez et. al., 1988).

3.4. Ideal pH and pH stability: Incompletely purged corrosive and soluble proteases were discovered to be dynamic over a scope of pH 1.0-12.0 utilizing casein and corrosive denatured ox-like hemoglobin as substrates for antacid and acidic proteases individually. The acidic protease showed high action in the pH range from 2-4 with an expected most extreme at 3.0 and afterward diminished essentially (p<0.05) with expanding pH (Fig. 4a). The relative movement of about over half was lost over pH 4. In other investigations, the ideal pH for hydrolysis of corrosive denatured cow-like hemoglobin by a halfway decontaminated acidic protease from Tilapia nilotica was discovered to be 2.5 (El-Beltagy et. al., 2004). Our outcomes substantiate well with the perception of (Bougatef et. al., 2009), who detailed pH optima for acidic proteases in the scope of 2-4. The basic protease displayed the most extreme action at pH 10 and afterward diminished fundamentally at higher pH levels (Fig. 4c). Ideal pH for most extreme movement of basic protease was accounted for in the scope of 8-10 (Nasri et. al., 2011). Assurance of pH optima of a protein is basic as this is viewed as a significant marker for its possible application for various purposes.

Both the acidic and basic proteases were exceptionally steady over a wide pH range, keeping up over 90% of its unique movement between pH 1.0-5.0 and pH 8.0-12.0 regarding corrosive and antacid proteases individually following 30 minutes brooding at 37°C (Fig. 4b, d). The pH steadiness of proteases relies upon the distinctions in atomic properties, which incorporates holding and soundness of the structure; adaptation of chemical in various anatomical areas among different species (Klomklao et. al., 2007). Comparative discoveries concerning pH dependability of acidic protease from fish have likewise been accounted for by (Castillo-Yanez et. al., 2004) for Monterey sardine. The pH solidness of antacid proteases in the scope of 6-12, has been accounted for by a few creators (Sila et. al., 2012)(Younes et. al., 2014). Acidic protease movement demonstrated a reduction of around 15-20% at pH over 6.0 though; a

comparative diminishing was appeared by basic protease at pH beneath 7.0.

3.5. Ideal temperature and thermostability: In this examination, the ideal movement of acidic protease was found at 40°C (Fig. 5a) which is like the prior reports from other fish, viz., pepsins from Sardinelle by (Ben Kahled et. al., 2008) and smooth dog by (Bougatef et. al., 2009). The ideal temperature of basic protease movement was found as 60°C (Fig. 5c) and the comparative outcome was accounted for by (Klomklao et. al., 2011) (Cao et. al., 2000) for trypsin from the pyloric caeca of Chinook salmon (Oncorhynchus tshawytscha) and Japanese seabass (Lateolabrax japonicas) separately.

The outcome demonstrated that the chemical action of proteases expanded in a specific way followed by a reduction with increment in temperature framing a ringer molded bend. At temperature above ideal, the local adaptation of protein is changed because of the breakdown of feeble intramolecular bonds capable of adjustment of the three-dimensional structure of the catalyst dynamic site (Klomklao et. al., 2011). As thought by (Klomklao et. al., 2006), natural and hereditary variables among the various species may be answerable for the local adaptations of catalysts.

The investigation likewise uncovered that acidic and soluble proteases" movement diminished by 40 and 60% individually when the hatching condition was 90°C for 30 min (Fig. 5b and d). This might be clarified as the inactivation of enzymatic action following loosening up of the enzymes local compliance during warm treatment (Klomklao et. al., 2011). As (Vannabun et. al., 2014) additionally announced comparable discoveries while portraying instinctive acidic and antacid proteases of cultivated monster feline fish. As proposed by (Sabtecha et. al., 2014), the soundness of a fish protein in various temperatures is affected by their natural surroundings, climate, and hereditary characters.

3.6. Impact of NaCl on catalyst action: As Fig. 6 shows the impact of NaCl on protein action of proteases. Relative protein action demonstrated a reduction of over half for both acidic and basic proteases at NaCl centralization of 0.5%. Further increment of NaCl focuses somewhat on diminished protease action. A 10% reduction in the general action of acidic protease from Sardinelle at 20% NaCl focus was accounted for by (Ben et. al., 2008). This demonstrates that the movement of proteases of freshwater fish contrasts from the marine fish, and this is essential because of salt centralization of the living space. (Klomklao et. al., 2011) explored trypsin action from crossover catfish and found that the compound action fundamentally diminished steadily with the expanding grouping of NaCl. This loss of compound movement maybe because of the denaturation of protein (Ben et. al., 2008) coming about the "salting out" impact.

The ionic quality is expanded with the expanding salt focus. In the high ionic quality, the compound movement

is decreased because of unrivaled hydrophobichydrophobic association between proteins of the chemical and upgraded liking of ionic salts for water consequently coming about precipitation of catalyst (Klomklao et. al., 2009).

3.7. Impact of Isolated enzymes on proteins hydrolysis: As the level of hydrolysis (DH) is the demonstrative of the degree of peptide bonds separated (Adler-Nissen et. al., 1979), its assurance is significant since a few

qualities of protein hydrolysates is DH subordinate. Utilizing ground muscle meat of fish as substrate, hydrolysis was directed at temperature 37°C and ideal pH for both the compounds. The level of hydrolysis (DH) as a component of the compound fixation is given in Fig. 7. The aftereffect of this examination connotes that higher measure of proteases in chemical division severed more peptide bonds and comparable perception was additionally announced by (Klompong et. al., 2008).

Table 4. Showa Purification Of Acidic And Basic Proteases From Instinctive Misuse Of Rohu								
	Steps for Cleansing	Protein Content (mg/mL)	Complete Activity (U/mL)	Explicit Activity (U/mg)	Recuperation (%)	Purification Fold		
Acidic Protease	Unrefined	6.31±0.08	18.33±0.06	2.90 <u>+</u> 0.05	100	01		
	Ammonium Sulfate	3.72±0.04	13.37±0.02	3.59 <u>+</u> 0.01	72.76	1.24		
	Fractionation (40-60%)							
	Dialysis	1.68±0.05	12.15±0.28	7.23±0.15	66.28	2.49		
Alkaline Protease	Unrefined	7.79±0.06	34.11±0.11	4.38±0.01	100	01		
	Ammonium Sulfate	4.15±0.03	21.72±0.76	5.23 <u>+</u> 0.04	63.67	1.19		
	Fractionation (40-60%)							
	Dialysis	2.96±0.03	20.11±0.61	6.79±0.07	58.95	1.55		

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*Values given in the table are implied \pm SD, n=3.

Figure 3: Shows Electrophoretic example of Rohu viscera squander (C=standard protein marker, A=crude catalyst concentrate, and B=purified chemical concentrate)



3.8. Impact of inhibitors on the catalyst action: High restraint rates of complete acidic protease action were gotten when the acidic protease was brooded with 50 mM of both soybean trypsin inhibitor or ethylenediaminetetraacetic corrosive (90.9% and 68.8%, separately), while they were 23.9% and 10.5% individually when 10 mM focus was utilized (Fig. 8). Practically comparative percent restraint was gotten in the event of basic protease. Our outcome is in concurrence with the discoveries of (Diaz-Lopez et. al., 1998) (El-Beltagy et. al., 2004) for the acidic protease. In an examination with tilapia stomach related proteases, a high hindrance of approx. 40% was accounted for utilizing low convergence of SBTI (Moyano et. al., 1999). Restraint of Rohu basic proteases at 250 µM centralization of SBTI was accounted for to be 78.1% (Kumar et. al., 2007).

Fig 4: Optimum pH and pH dependability for most extreme movement of proteases from Rohu viscera squander (a-ideal pH for acidic proteases, b-pH solidness of acidic proteases, c-ideal pH for antacid proteases, thed-pH steadiness of soluble proteases)





Fig 5: Optimum temperature and thermostability for most extreme action of proteases from Rohu viscera squander (a-ideal temp. a necessity for acidic proteases, b-thermostability of acidic proteases, c-ideal temp. a necessity for antacid proteases, and d-thermostability of basic proteases)









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CONCLUSION

Taking everything into account, this examination has uncovered that impressive measures of acidic and antacid proteases are available in the instinctive misuse of Rohu fish, and those in sanitized structure have the potential for application as various food handling helps, and then again, would add to tackle bio-garbage removal issue generally. Both the proteases showed considerable action in both acidic and basic conditions. As a necessity for their application reason, the most extreme movement of acidic and basic protease was discovered to be at 40°C and 60°C individually. By the by, the solidness of these compounds at raised temperature and NaCl focus was not discovered to be good.

Because of the current investigation, the proteins from Rohu instinctive waste could discover use in applications where most extreme action at moderate temperature and low NaCl focus is wanted. The fish preparing industry in India produces enormous protein-rich material that is untapped and can be used by changing over into protein hydrolysate. Relies upon the properties and synthetic synthesis, further FPH discovers application in different ventures going from nutraceutical to plant developmentboosting fixing. Ongoing enthusiasm of FPH as nutraceutical compound/bioactive peptide requests sterile taking care of and appropriate conservation of fish preparing waste. Be that as it may, the wellbeing of FPH when delivered from fishery squander, financial attainability, and business case are stays unaddressed around the world.

REFERENCES

Adler-Nissen, J. 1979. Determination of the degree of hydrolysis of food protein hydrolysates by trinitrobenzene sulfonic acid. Journal of agricultural and food chemistry. 27(6): 1256-1262.

Adler-Nissen, J.,1986. Enzymic Hydrolysis of Food Proteins, Elsevier Applied Science Publishers, Barking, UK.

Ahn CB, Lee KH, Je JY. Enzymatic production of bioactive protein hydrolysates from tuna liver: effects of enzymes and molecular weight on bioactivity. International journal of food science & technology. 2010 Mar 1;45(3):562-8.

Anwar, A., and Saleemuddin, M. 1998. Alkaline proteases: a review. Bioresource technology. 64(3): 175-183.

Arnesen, J. A., and Gildberg, A. 2007. Extraction and characterization of gelatine from Atlantic salmon (Salmo salar) skin. Bioresource Technology. 98(1): 53-57.

Balti, R., Barkia, A., Bougatef, A., Ktari, N., and Nasri, M. 2009. Heat-stable trypsin from the hepatopancreas of the cuttlefish (Sepia Officinalis): Purification and characterization. Food Chemistry. 113(1): 146-154.

Barkia, A., Bougatef, A., Nasri, R., Fetoui, E., Balti, R., and Nasri, M. (2010). Trypsin from the viscera of Bogue (Boops boops): isolation and characterization. Fish physiology and biochemistry. 36(4): 893-902.

Batista I, Ramos C, Coutinho J, Bandarra NM, Nunes ML. Characterization of protein hydrolysates and lipids obtained from black scabbardfish (Aphanopuscarbo) by-products and antioxidative activity of the hydrolysates produced. Process Biochemistry. 2010 Jan 31;45(1):18-24.

Ben Khaled, H., Bougatef, A., Balti, R., Triki- Ellouz, Y., Souissi, N., and Nasri, M. 2008. Isolation and characterization of trypsin from sardinelle (Sardinella aurita) viscera. Journal of the Science of Food and Agriculture. 88(15): 2654-2662.

Bezerra, R. S., Lins, E. J., Alencar, R. B., Paiva, P. M., Chaves, M. E., Coelho, L. C., and Carvalho Jr, L. B. 2005. Alkaline proteinase from intestine of Nile tilapia (Oreochromis niloticus). Process Biochemistry. 40(5):

Das et al.,

1829-1834.

Bhaskar, N., & Mahendrakar, N. S. 2007. Chemical and microbiological changes in acid ensiled visceral waste of Indian major carp Catla (Hamilton) with emphasis on proteases. Indian Journal of Fisheries. 54(2): 217-225. Bougatef, A., Hajji, M., Balti, R., Lassoued, I., Triki-Ellouz, Y., and Nasri, M. 2009. Antioxidant and free radical-scavenging activities of smoothhound (Mustelusmustelus) muscle protein hydrolysates obtained by gastrointestinal proteases. Food chemistry. 114(4): 1198-1205.

Cao, M. J., Osatomi, K., Suzuki, M., Hara, K., Tachibana, K., and Ishihara, T. 2000. Purification and characterization of two anionic trypsins from the hepatopancreas of carp. Fisheries Science, 66(6): 1172acidic protease from the viscera of bolti fish (Tilapia nilotica). Food Chemistry. 86(1): 33-39.

Castillo-Yanez, F. J., Pacheco-Aguilar, R., Garcia-Carreño, F. L., and de los Angeles Navarrete-Del, M. 2004. Characterization of acidic proteolytic enzymes from Monterey sardine (Sardinops sagax caerulea) viscera. Food Chemistry. 85(3): 343-350.

Chakrabarti, R., and Sharma, J. G. 2005. Digestive physiology of fish larvae during ontogenic development: a brief overview. The Indian Journal of Animal Sciences. 75(11): 1337–1347.

Chalamaiah M, Hemalatha R, Jyothirmayi T. Fish protein hydrolysates: proximate composition, amino acid composition, antioxidant activities, and applications: a review. Food Chemistry. 2012 Dec 15;135(4):3020-38. Chalamaiah M, Jyothirmayi T, Bhaskarachary K, Vajreswari A, Hemalatha R, Kumar BD. Chemical composition, molecular mass distribution, and antioxidant capacity of rohu (Labeorohita) roe (egg) protein hydrolysates prepared by gastrointestinal proteases. Food research international. 2013 Jun 30;52(1):221-9.

Characterization of Protease. Journal of Cell and Tissue Research. 11(1): 2589.

Diaz-López, M., Moyano-López, F. J., Alarcón- López, F. J., Garcia-Carreno, F. L., and del Toro, M. A. N. 1998. Characterization of fish acid proteases by substrate-gel electrophoresis. Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology. 121(4): 369-377.

El- Beltagy, A. E., El- Adawy, T. A., Rahma, E. H., and El-Bedawey, A. A. 2005. Purification and characterization of an alkaline protease from the viscera of bolti fish (Tilapia nilotica). Journal of food biochemistry. 29(5): 445-458.

Elavarasan K, Naveen Kumar V, Shamasundar BA. Antioxidant and functional properties of fish protein hydrolysates from freshwater carp (Catlacatla) as influenced by the nature of the enzyme. Journal of Food Processing and Preservation. 2014 Jun 1;38(3):1207-14.

Elavarasan K, Shamasundar BA, Badii F, Howell N.

Angiotensin I-converting enzyme (ACE) inhibitory activity and structural properties of oven-and freeze-dried protein hydrolysate from freshwater fish (Cirrhinusmrigala). Food chemistry. 2016 Sep 1;206:210-6.

Elavarasan K, Shamasundar BA. Effect of oven drying and freeze-drying on the antioxidant and functional properties of protein hydrolysates derived from freshwater fish (Cirrhinusmrigala) using papain enzyme. Journal of food science and technology. 2016 Feb 1;53(2):1303-11.

El-Beltagy, A. E., El-Adawy, T. A., Rahma, E. H., and El-Bedawey, A. A. 2004. Purification and characterization of an alkaline protease from the viscera of bolti fish (Tilapia nilotica). Journal of food biochemistry. 29(5): 445-458.

Gajanan PG, Elavarasan K, Shamasundar BA. Bioactive and functional properties of protein hydrolysates from fish frame processing waste using plant proteases. Environmental Science and Pollution Research. 2016 Dec 1;23(24):24901-11.

Godfrey, T., and Reichelt, J., 1983. Industrial Enzymology. The Application of Enzymes in Industry, MacMillan, London, UK.

Godinho IS. Production of fish protein hydrolysates by a marine proteolytic strain (Doctoral dissertation, ISA).

Grabner, M. 1985. An in vitro method for measuring protein digestibility of fish feed components. Aquaculture. 48(2): 97-110.

Gupta, R., Beg, Q., and Lorenz, P. 2002. Bacterial alkaline proteases: molecular approaches and industrial applications. Applied microbiology and biotechnology. 59(1): 15-32.

Hoyle, N. T., and Merrltt, J. H. 1994. Quality of fish protein hydrolysates from herring (Clupea harengus). Journal of food science. 59(1): 76-79.

Je JY, Lee KH, Lee MH, Ahn CB. Antioxidant and antihypertensive protein hydrolysates produced from tuna liver by enzymatic hydrolysis. Food Research International. 2009 Nov 30;42(9):1266-72.

Jemil I, Jridi M, Nasri R, Ktari N, Salem RB, Mehiri M, Hajji M, Nasri M. Functional, antioxidant and antibacterial properties of protein hydrolysates prepared from fish meat fermented by Bacillus subtilis A26. Process Biochemistry. 2014 Jun 30;49(6):963-72.

Jhingran, V. G. 1991. Fish and Fisheries of India. (3rd Ed.). Hindustan Publishing Corporation, New Delhi.

Kester, J.J., and Richardson, T. 1984. Modification of whey proteins to improve functionality. J. Dairy Sci., 67: 2757-2774

Kim SK, Mendis E. Bioactive compounds from marine processing byproducts–a review. Food Research International. 2006 May 31;39(4):383–93.

Kim, S. K., Ngo, D. H., and Vo, T. S. 2012. Marine fishderived bioactive peptides as potential antihypertensive agents. In Kim, S. K. (Eds.), Advances in food and nutrition research. Academic Press, United States. pp. 249-260.

Klomklao, S., Benjakul, S., Kishimura, H., and Chaijan, M. 2011. 24 kDa Trypsin: A predominant protease purified from the viscera of hybrid catfish (Clarias macrocephalus× Clarias gariepinus). Food chemistry. 129(3): 739-746.

Klomklao, S., Benjakul, S., Visessanguan, W., Kishimura, H., and Simpson, B. K. 2006. Purification and characterization of trypsin from the spleen of tongol tuna (Thunnus tonggol). Journal of agricultural and food chemistry. 54(15): 5617-5622.

Klomklao, S., Benjakul, S., Visessanguan, W., Kishimura, H., and Simpson, B. K. 2007. Trypsin from the pyloric caeca of bluefish (Pomatomus saltatrix). Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology. 148(4): 382-389.

Klomklao, S., Benjakul, S., Visessanguan, W., Kishimura, H., Simpson, B. K., and Saeki, H. 2006. Trypsins from yellowfin tuna (Thunnus albacores) spleen: purification and characterization. Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology. 144(1): 47-56.

Klomklao, S., Kishimura, H., Nonami, Y., and Benjakul, S. 2009. Biochemical properties of two isoforms of trypsin purified from the intestine of skipjack tuna (Katsuwonus pelamis). Food Chemistry. 115(1): 155-162.

Klompong, V., Benjakul, S., Kantachote, D., Hayes, K. D., and Shahidi, F. 2008. Comparative study on antioxidative activity of yellow stripe trevally protein hydrolysate produced from Alcalase and Flavourzyme. International journal of food science & technology. 43(6): 1019-1026.

Kristinsson HG, Rasco BA. Fish protein hydrolysates: production, biochemical, and functional properties. Critical reviews in food science and nutrition. 2000 Jan 1;40(1):43-81.

Kumar, S., Garcia- Carreño, F. L., Chakrabarti, R., Toro, M. A. N., and Córdova- Murueta, J. H. 2007. Digestive proteases of three carps Catla, Labeorohita, and Hypophthalmichthys molitrix: partial characterization and protein hydrolysis efficiency. Aquaculture Nutrition. 13(5): 381-388.

Laemmli, U. K. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. Nature. 227(5259): 680.

Linder, M., Fanni, J., Parmentier, M., Sergent, M. and Phan-Tan-Luu, R., 1995, Protein recovery from veal bones by enzymatic hydrolysis. J. Food Sci., 60: 949–952

Liu, Z. Y., Wang, Z., Xu, S. Y., and Xu, L. N. 2007. Two trypsin isoforms from the intestine of the grass carp (Ctenopharyngodon idellus). Journal of Comparative Physiology B. 177(6): 655- 666.

López, F. M., Dıaz, I. M., López, M. D., and López, F. A. 1999. Inhibition of digestive proteases by vegetable meals in three fish species; seabream (Sparus aurata),

tilapia (Oreochromis niloticus), and African sole (Solea senegalensis). Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology. 122(3): 327-332.

Lopez-Llorca, L. V. 1990. Purification and properties of extracellular proteases produced by the nematophagous fungus Verticillium suchlasporium. Canadian Journal of Microbiology. 36(8): 530-537.

Lowry, O. H., Rosebrough, N. J., Farr, A. L., and Randall, R. J. 1951. Protein measurement with the Folin phenol reagent. Journal of biological chemistry. 193(1): 265-275.

Mahmoud, M.I., Malone, W.T., and Cordle, C.T., 1992. Enzymatic hydrolysis of casein: effect of degree of hydrolysis on antigenicity and physical properties, J. Food Sci., 57: 1223-1229

Maitra, S., and Ray, A. K. 2003. Inhibition of digestive enzymes in rohu, Labeo rohita (Hamilton), fingerlings by tannin: an in vitro study. Aquaculture Research. 34(1): 93-95.

Martínez, A., Olsen, R. L., and Serra, J. L. 1988. Purification and characterization of two trypsinlike enzymes from the digestive tract of anchovy Engraulis encrasicholus. Comparative biochemistry and physiology. B, Comparative biochemistry. 91(4): 677-684.

Moyano, F. J., Diaz, M., Alarcon, F. J., and Sarasquete, M. C. 1996. Characterization of digestive enzyme activity during larval development of gilthead seabream (Sparus aurata). Fish Physiology and Biochemistry. 15(2): 121-130.

Murthy PS, Rai AK, Bhaskar N. Fermentative recovery of lipids and proteins from freshwater fish head waste concerning antimicrobial and antioxidant properties of protein hydrolysate. Journal of food science and technology. 2014 Sep 1;51(9):1884-92.

Nasri, R., Younes, I., Lassoued, I., Ghorbel, S., Ghorbel-Bellaaj, O., and Nasri, M. 2011. Digestive alkaline proteases from Zosterisessor ophiocephalus, Raja clavata, and Scorpaena scrofa: characteristics and application in chitin extraction. Journal of amino acids. 2011, 1-9.

Natalia, Y., Hashim, R., Ali, A., and Chong, A. 2004. Characterization of digestive enzymes in a carnivorous ornamental fish, the Asian bony tongue Scleropages formosus (Osteoglossidae). Aquaculture. 233(1-4): 305-320.

Pastoriza L, Sampedro G, Cabo ML, Herrera JJ, Bernárdez M. Solubilisation of proteins from rayfish residues by endogenous and commercial enzymes. Journal of the Science of Food and Agriculture. 2004 Jan 15;84(1):83-8.

Raksakulthai, R., and Haard, N. F. 1999. Purification and characterization of aminopeptidase fractions from squid (Illex illecebrosus) hepatopancreas. Journal of food biochemistry. 23(2): 123-144.

Rawdkuen, S., Chaiwut, P., Pintathong, P., and Benjakul,

Das et al.,

S. 2010. Three-phase partitioning of protease from Calotropis procera latex. Biochemical Engineering Journal. 50(3): 145-149.

Richardson, T., and Hyslop, D.B., 1984. Enzymes. In: Food Chemistry Edt. Fennema, O.R. Edn. 2nd ., Marcel Dekker Inc., New York.

Rustad T. Physical and chemical properties of protein seafood by-products. Maximizing the value of marine by-products. 2007:3-21.

Sabtecha, B., Jayapriya, J., and Tamilselvi, A. 2014. Extraction and characterization of proteolytic enzymes from fish visceral waste: potential applications as destainer and dehairing agent. International Journal of ChemTech Research. 6(10): 4504-4510.

Schaafsma G. Safety of protein hydrolysates, fractions thereof, and bioactive peptides in human nutrition. European journal of clinical nutrition. 2009 Oct 1;63(10):1161- 8.

Sekizaki, H., Itoh, K., Murakami, M., Toyota, E., and Tanizawa, K. 2000. Anionic trypsin from chum salmon: activity with p- amidinophenyl ester and comparison with bovine and Streptomyces griseus trypsins. Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology. 127(3): 337-346.

Sila, A., Nasri, R., Jridi, M., Balti, R., Nasri, M., and Bougatef, A. 2012. Characterization of trypsin purified from the viscera of Tunisian barbel (Barbus callensis) and its application for recovery of carotenoproteins from shrimp wastes. Food chemistry. 132(3): 1287-1295.

Simpson, B.K. 2000. Digestive proteinases from marine

animals. In: Seafood Enzymes: Utilization and Influence on Postharvest Seafood Quality (Eds.) B.K. Simpson and N.F. Haard. Marcel Dekker, New York. Pp. 531–540.

Subash, A., Shobana, A., Shanmugakani, G., Swetha, V. P., and Josephine, J. L. 2011. Utilization of Visceral Waste of Ribbon Fish (Trachipterus Trachypterus) for the Extraction, Purification and

Temiz, H., Ustun, N. S., Turhan, S., and Aykut, U. 2013. Partial purification and characterization of alkaline proteases from the Black Sea anchovy (Engraulis encrasicholus) digestive tract. African Journal of Biotechnology. 12(1): 56-63.

Vannabun, A., Ketnawa, S., Phongthai, S., Benjakul, S., and Rawdkuen, S. 2014. Characterization of acid and alkaline proteases from viscera of farmed giant catfish. Food Bioscience. 6, 9-16.

Venugopal, V. Production of fish protein hydrolysates by microorganisms. pp. 223 – 243, in Martin, AM (Ed) 1994. Fisheries processing: biotechnological applications. Chapman & Hall, London, UK.

Villalba-Villalba, A. G., Ramírez-Suárez, J. C., Valenzuela-Soto, E. M., Sanchez, G. G., Ruiz, G. C., and Pacheco-Aguilar, R. 2013. Trypsin from viscera of vermiculated sailfin catfish, Pterygoplichthys disjunctivus, Weber, 1991: Its purification and characterization. Food chemistry. 141(2): 940-945.

Younes, I., Nasri, R., Bkhairia, I., Jellouli, K., and Nasri, M. 2015. New proteases extracted from red scorpionfish (Scorpaena scrofa) viscera: Characterization and application as a detergent additive and for shrimp waste deproteinization. Food and Bioproducts Processing. 94, 453-462.