

***In silico* Identification of Protein in *Ralstonia solanacearum*, A Bacterial Wilt Pathogen for Drug Target by Subtractive Genomic Analysis**

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ABSTRACT

Ralstonia solanacearum is a devastating pathogenic soil borne bacterium causing Bacterial Wilt disease in 450 plant species belonging to 54 botanical families and it severely impairs global solanaceous crop production. The loss of crop may go up to 90% depending upon the environmental suitability. The bacterium is very robust and can survive in diverse host plants, soil, water and even in weeds. It possesses an arsenal of secretory molecules like diverse virulent factors, exopolysaccharide, cell wall degrading enzymes to subvert host defense mechanisms. The wilt pathogen is also very efficient to overcome existing control measures rendering it extremely difficult to control. Understanding of molecular mechanism of pathogenesis through genome analysis and identification of novel drug target could be an effective alternative. In this study, subtractive genome analysis of *Ralstonia solanacearum* GM1000 strain having total 5106 proteins obtained from Uniprot database was done and 4972 non paralogous sequence of proteins were selected applying CD-HIT tool. A total of 465 essential proteins were then identified using BLASTp tools of DEG database. Functional pathway assessment of 424 essential proteins revealed 117 metabolically active proteins using KAAS server and a total of 7 non homologous proteins exclusive to the pathogen were identified using BLASTp algorithm. After screening the druggability of 7 proteins in DrugBank Database, 4 proteins were shortlisted and further analyzed for subcellular localization using PSORTb tool. After survey of the existing literature, type II secretory pathway *gspe*-related protein has been identified and predicted to be the best possible target for drug designing. The present work reports for the first time that type II secretory system could serve as drug target and therefore, opens a new avenue for *in silico* screening of novel molecules for effective control of bacterial wilt in future.

KEY WORDS: DRUG DESIGN, *RALSTONIA SOLANACEARUM*, SUBTRACTIVE GENOME ANALYSIS, WILT DISEASE.

INTRODUCTION

Soil born bacterium *Ralstonia solanacearum* is the most devastating plant pathogenic bacteria that causes wilt

diseases in many wide varieties of plants (Yuliar, Nion, and Toyota, 2015). The strains of this pathogen can infect 450 plant species distributed in 54 botanical families, including potatoes, tomatoes, brinjal, tobacco etc. (Wicker et al., 2007). It invades through the wounded roots or natural opening and colonize in the vascular tissues and release viscous exopolysaccharide that causes obstruction in xylem conduction and lead to fatal wilting disease symptoms in the plants (Schell MA, 2000). Direct yield losses by *R. solanacearum* vary widely according to the host, cultivar, climate, soil type, cropping pattern.

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It has been reported that it accounts for 80% loss in tobacco, 100% in banana, and up to 20% in the groundnut (Elphinstone, 2005; Somani et al., 2010). *Ralstonia* infection causes more than 50% crop loss in India and that may reach up to 75% in some parts of Karnataka (Gadewar et al., 1991). Bacterial wilt disease affects potato cultivation in different parts of India and accounts for 30 to 70 % crop loss in these areas (Somani et al., 2010). The control of bacterial wilt pathogen is very challenging. Difficulties are associated with controlling this pathogen due to its abilities to grow endophytically, long survival in the soil especially in the deeper layers, travel along water, and its relationship with weeds (Wang et al., 2005; Mansfield et al., 2012; Santana et al., 2020; Yan and Gao, 2020).

The bacterial pathogen often undergoes VBNC (Viable but not culturable) state under unfavorable condition (Van Elsas et al., 2001). Furthermore, many environmental stresses weaken the defense systems of the plants allowing to proliferate *Ralstonia* and other bacterial endophytes inside the host. Conventional disease management practice such as preventive measures, cultural practices are inefficient to pre-existing infection and because of the pathogen's diverse host range and persistence in the weeds and soil (Mbaka et al., 2013). Chemical pesticides such as algicide (3-[3-indolyl] butanoic acid), fumigants (Metam sodium, 1, 3-dichloropropene, and chloropicrin), and plant activators (validamycin A and validoxylamine) inducing systemic resistance in the tomato have been used to control bacterial wilt but with limited success (Ishikawa et al., 2007; Yuliar et al., 2015; Coutinho et al., 2017).

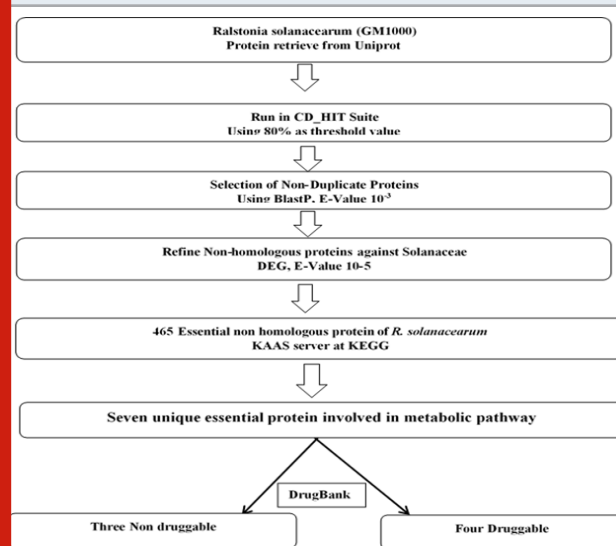
Copper compounds (copper hydroxide (CH), copper hydroxide-oxadixyl, and copper oxychloride-dithianon), and essential oils (Cinnamon oil, Clove oil) have been partially effective to control bacterial wilt. (Elphinstone, 2005; Lee et al., 2012). Many bactericides such as triazolothiadiazine [0.5 to 12 mM, in solution], streptomycin sulfate [400 mg kg⁻¹ of soil] have been employed to control bacterial wilt pathogen with average rate of success (Khanum et al., 2005; Lin et al., 2010). Additionally, emergence antibiotic resistance and environmental pollution due to long-term use of chemical pesticides rendered bacterial wilt disease management very difficult. Although, there are many studies have been done employing biocontrol strategy to control bacterial wilt but of limited success due to inefficient colonization, narrow range and requirement of high inoculum of biocontrol agents. Therefore, identification of novel pathogenic target protein and discovery of its corresponding drug could be an attractive alternative for controlling bacterial wilt disease (Whipps and Gerhardson, 2007; Coutinho et al., 2017).

Rapid advancement in the field of biotechnology enabled us to have vast genomic data from the prokaryotic whole genome projects that in turn may be exploited for finding novel drug targets and virulent factors in microbes. With the availability of whole genome sequence, subtractive genome analysis has been evolved as a very efficient

tool to identify novel drug targets and virulent factors in pathogenic microbes (Miesel et al., 2003; Amineni et al., 2010; Keshri et al., 2014). Subtractive genome analysis is a smart technique to identify essential metabolic gene present exclusively in the pathogen having no homologue in the host and therefore, the targeted drug developed against the pathogenic essential metabolic gene will impair only the metabolic function of the pathogen leaving the host metabolism undisturbed (Vetrivel et al., 2011; Barh et al., 2011). Many possible drug targets have been identified in human pathogenic bacteria (Barh et al., 2011; Sudha et al., 2019; Santana et al., 2020; Yan and Gao, 2020).

However, there are very few reports regarding drug target identification in plant pathogenic bacteria using *in silico* techniques (Allen et al., 2009; Silver, 2011). Subtractive hybridization technique has been exploited to underpin drug targets in rice bacterial pathogen, *Xanthomonas* by some researchers (Keshri et al., 2014; Prava et al., 2019). Although, the complete genome sequence of *Ralstonia solanacearum* is available in the database, but there is no report available so far that have tried subtractive genome analysis to identify drug targets in this bacterium. Therefore, the present work is attempted to identify possible drug targets in *Ralstonia solanacearum* through subtractive genome analysis and other *in silico* analysis tools (Prava et al., 2019).

Figure 1: The conceptual framework showing the methodology followed for the analysis



MATERIAL AND METHODS

Subtractive genomic approach was applied for the identification of essential proteins in the *Ralstonia solanacearum* (GM1000) which were then analyzed for the identification potential drug targets. The identified drug target was then screened through DrugBank database to evaluate druggability scope. Network based analysis was done for the identification of metabolic activity of target protein (Yu et al., 2010).

The complete proteome of *Ralstonia solanacearum* GM1000 strain was retrieved from UniProt (<http://www.uniprot.org>). The UniProt Knowledgebase is the central hub for the collection of functional information on proteins, with accurate, consistent and rich annotation (The UniProt Consortium, 2019). Identification of nonhomologous protein and essential gene of *Ralstonia solanacearum* - Paralogueous sequences were excluded from the complete proteome of *Ralstonia solanacearum*

GM1000 strain by using CD-HIT at 80% threshold. BLASTp was performed for the remaining proteins against Solanaceae using threshold expectation value (E Value) 10^{-3} as parameter. Non homologous protein sequences were then subjected to BLASTp against the database of essential genes (DEG) assessed at DEG database (<http://tubic.tju.edu.cn/deg/>) using E-Value cut off of 10^{-5} , to screen out essential gene proteins (Li et al., 2001).

Table 1. Unique metabolic pathway essential proteins

Sl no	DEG ID	UNIPROT ID/ DRUGGABILITY	METABOLIC PATHWAY
1	DEG10570448	Q8XW91 Druggable	QUORUM SENSING
2.	DEG10570275	Q8XX10 Druggable	BACTERIAL SECRETION SYSTEM
3.	DEG10570247	Q8Y3B8 Druggable	PEPTIDOGLYCAN BIOSYNTHESIS BETA LACTAM RESISTANCE
4.	DEG10570255	Q8XVI1 Druggable	BETA LACTAM RESISTANCE PEPTIDOGLYCAN BIOSYNTHESIS
5.	DEG10570232	Q8XQ85 Not Druggable	BACTERIAL CHEMOTAXIS
6.	DEG10570446	Q8XVG2 Not Druggable	QUORUM SENSING
7.	DEG10570220	Q8XX15 Not Druggable	BACTERIAL SECRETION SYSTEM

Sub Cellular Localization

Name of Protein	Uniprot ID	Location
Probable conjugal transfer protein trbb	Q8XW91	cytoplasmic
Probable type II secretory pathway gspe-related protein (RSc2308)	Q8XX10	cytoplasmic
Peptidoglycan D, D-transpeptidase MrdA	Q8Y3B8	cytoplasmic
Peptidoglycan D, D-transpeptidase FtsI	Q8XVI1	Cytoplasmic Membrane

KEGG Automatic annotation Server (KAAS) was accessed to analyze the metabolic pathway of the essential proteins of *Ralstonia solanacearum* GM1000 strain for the identification of potential drug target. The server performs BLASTp comparisons of the query protein against Kyoto Encyclopedia of Genes and Genomes (KEGG) Genes Database (Moriya et al., 2007). Sub Cellular localization of non-homologous essential proteins of bacteria illustrates their potential of becoming the possible drug targets. Therefore PSORTb tools at Expsy server was utilized to identify the subcellular localization of non-homologous essential protein sequences (Yu et al., 2010). The modulation of the activity of a protein target with a small molecule of a drug accounts for its prospective druggability. DrugBank Database was accessed to calculate the druggability potential of each identified drug target (Knox et al., 2011). BLASTp with

default parameters was used to align the potential drug targets from *Ralstonia solanacearum* against the list of the of compounds found within the DrugBank (Szklarczyk et al., 2019).

Selected indispensable proteins were then subjected to STRING database (<http://string.embl.de>) to construct protein-protein interaction network (Li, Jaroszewski and Godzik, 2001). Interactors with confidence score greater than or equal to 0.700 alone included here in the protein network and with low and medium confidence score were eliminated to avoid false positive and false negative results. Target protein with more interactors is considered as a metabolically active protein which could serve as appropriate Drug target (Peyraud et al., 2017; Szklarczyk et al., 2019).

RESULTS AND DISCUSSION

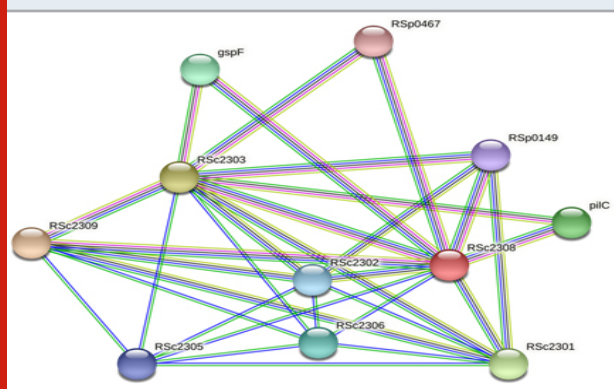
The main goal of the subtractive genomic analysis was to examine *Ralstonia solanacearum* GM1000 strain critical proteins as a possible drug target for future strategic drug discovery. Total 5106 proteins of total proteome

were originally obtained from *Ralstonia solanacearum* GM1000 Uniprot database. The CD-HIT tool was used to differentiate paralogous and non-paralogous proteins. 134 paralogous proteins were screened and 4972 non paralogous sequence of proteins were selected for further analysis.

Table 2. Non homologous essential protein of *Ralstonia solanacearum* strain similar to binding pattern of FDA approved drugs against DrugBank database using BLASTp

Sl. no	Protein name	DrugBank ID	Uniprot ID
1.	Conjugal transfer protein trbb	DB02930 DB04395	Q8XW91
2.	Type II secretory pathway gspe-related protein (RSc2308)	DB04395 DB02930	Q8XX10
3.	Peptidoglycan D, D-transpeptidase MrdA	DB01413, DB00438, DB14879, DB01598, DB01329, DB01327, DB01163, DB01163, DB01328, DB01413, DB01415, DB00948, DB00438, DB00303, DB00671, DB01326, DB00923DB00355, DB00493, DB04570, DB01413, DB01147, DB09050, DB06211, DB14879, DB04918DB00274, DB00430, DB01607, DB01000, DB02443, DB02968, DB04041, DB01603, DB00417	Q8Y3B8
4.	Peptidoglycan D, D-transpeptidase FtsI	DB06211, DB14879DB04918, DB00267, DB01413, DB01147, DB09050, DB01416, DB01329, DB01327, DB01331, DB01328, DB01413, DB01415, DB00430DB05659, DB00535, DB04918, DB01150DB03190	Q8XVI1

Figure 2: Interaction among Type II secretory pathway gspe-related protein (RSc2308) and other proteins of *R. solanacearum*.



The selected proteins were assessed against Solanaceae proteome in BLASTp, with an E-value cut off 10^{-3} . Selected non homologous proteins were employed for the identification of essential gene using BLASTp tools of DEG database at default parameter settings. The analysis identified 465 essential proteins. There are 41 hypothetical proteins were identified which were finally excluded in this study. The essential proteins of bacteria

are expected to be involved in housekeeping and are important for the survival of pathogen.

Total no of protein	5106
Duplicate (>80% identical) in CD-HIT	134
Essential proteins in DEG (E-value 10-5)	465
Number of hypothetical proteins as essential proteins	41
Essential proteins involved in metabolic pathway	117
Unique metabolic pathway essential proteins	7
Essential proteins found to be druggable	4

Functional pathway assessment of 424 essential proteins were conducted using KAAS server. Among 424 proteins 117 proteins were found to be involved in different metabolic pathway of the pathogen. These 117 proteins were further analyzed by the BLASTp algorithm for the comparison of metabolic pathway in *Ralstonia solanacearum* proteome and *Solanum tuberosum* proteome as a reference organism of Solanaceae family to exclude the common pathway. Total seven pathogen specific pathways of *Ralstonia solanacearum* GM 1000 were identified by KEGG which were absent in Solanaceae family.

Table 3. Interaction among Type II secretory pathway gspe-related protein (RSc2308) and other proteins of *R. solanacearum* and their combined score.

node1	node2	node1 string id	node2 string id	combined_score
RSc2300	RSc2308	267608.RSc2300	267608.RSc2308	0.762
RSc2301	RSc2308	267608.RSc2301	267608.RSc2308	0.922
RSc2302	RSc2308	267608.RSc2302	267608.RSc2308	0.886
RSc2303	RSc2308	267608.RSc2303	267608.RSc2308	0.955
RSc2304	RSc2308	267608.RSc2304	267608.RSc2308	0.867
RSc2305	RSc2308	267608.RSc2305	267608.RSc2308	0.884
RSc2306	RSc2308	267608.RSc2306	267608.RSc2308	0.887
RSc2307	RSc2308	267608.RSc2307	267608.RSc2308	0.845
RSc2309	RSc2308	267608.RSc2309	267608.RSc2308	0.981
RSc2310	RSc2308	267608.RSc2310	267608.RSc2308	0.869
RSp0143	RSc2308	267608.RSp0143	267608.RSc2308	0.772
RSp0149	RSc2308	267608.RSp0149	267608.RSc2308	0.884
RSp0467	RSc2308	267608.RSp0467	267608.RSc2308	0.882
RSp0474	RSc2308	267608.RSp0474	267608.RSc2308	0.715
gspD	RSc2308	267608.RSc3114	267608.RSc2308	0.756
gspF	RSc2308	267608.RSc3116	267608.RSc2308	0.895
pilC	RSc2308	267608.RSc2826	267608.RSc2308	0.896
pilD	RSc2308	267608.RSc2827	267608.RSc2308	0.790

Total seven nonhomologous proteins were identified that are thought to be essential and involved in pathogens unique metabolic pathway. Therefore, new drugs may be designed to target these essential proteins to inhibit one or more of these metabolic pathways thereby controlling the growth and viability of the pathogenic strain *Ralstonia solanacearum* omit GM 1000. The total seven non homolog essential proteins (Table1) so obtained were verified within DrugBank Database for possible druggability and four essential non homologous proteins (Table 2) were identified to have druggability potential. Thereafter, the four selected proteins were then subjected to PSORTb for their sub cellular localization.

Earlier, 20 proteins of *Ralstonia solanacearum* were targeted for drug design having Protein Data Bank (PDB) ID of 3ZI8, 4I68, 4KF9, 4FDB, 3UMB, 3TMB, 3TOT, 3TOU, 3NPN, 3NPQ, 3LOP, 3GG9, 3GHY, 3EN2, 2QGU, 2CHH, 2BT9, 2BS5, 2BS6, 1UQX. (Kotaki and Saikia, 2015). Peptidoglycan D, D-transpeptidase MrdA, Peptidoglycan D, D-transpeptidase FtsI, Type II secretory pathway gspe-related protein were identified as the best predicted protein for drug target in this study. Type II secretion system is a virulent factor of *R. solanacearum* (Peeters et al., 2013). Inhibition of Quorum sensing protein can only prevent biofilm formation of pathogenic bacteria without any apparent direct effect on survivability. However, Peptidoglycan D, D-transpeptidase MrdA, Peptidoglycan D, D-transpeptidase FtsI protein as drug targets have already been reported and efforts have been taken for drug design in many human pathogenic bacterial strains, but these drug targets are inapplicable for *Ralstonia*

solanacearum strains as β lactam antibiotics are less effective in controlling bacterial wilt disease (Souvage and Terrak, 2016; Waack et al., 2017).

Different secretion systems of bacteria are very attractive targets for alternative therapeutics because their inactivation interferes with the delivery of secreted virulence factors. There are many cell walls degrading enzymes are secreted through Type II secretory system (T2SS) in *Ralstonia solanacearum*. Therefore, inhibitor of Type II secretory system (T2SS) could be a good alternative for drug design. Rsc2308 (UniProtKB ID-Q8XX10) is the Type II secretory pathway gspe-related protein of *Ralstonia solanacearum* associated with secretory system of bacteria which is responsible for pathogenicity. Therefore, Type II secretory pathway gspe-related protein (RSc2308) of *Ralstonia solanacearum* could be a promising drug target for future drug design that has not been properly addressed so far. Network based analysis showed that this protein Rsc3208 is interconnected with eighteen proteins in network with combined score greater than 0.7 (Table3) (Salanoubat et al., 2001; Waack et al., 2017).

So, it may be assumed that this Type II secretory pathway gspe-related protein is a highly metabolically active protein and inhibition of this protein may arrest the growth of the bacteria. Therefore, the present work opens a new avenue for searching novel drug compounds that may interact with the target Type II secretory pathway gspe-related protein (RSc2308) and may pave the path for new control strategy (Souvage and Terrak, 2016).

CONCLUSION

Subtractive genome analysis revealed possible drug targets in many human pathogenic bacteria and only few in plant pathogenic bacteria. *In silico* identification of possible drug target in *Ralstonia solanacearum* is completely lacking. Therefore, the present work probably is the first report underpinning the druggability of type II secretory pathway gspe-related protein of *Ralstonia solanacearum* through subtractive genome analysis. The gspe-related protein is essential in type-2 secretion pathway for secreting cell wall degrading enzymes that are key to host penetration and colonization. Therefore, targeting the protein with new drugs may prevent host colonization and survival in the weeds thereby offering a good strategy for controlling the pathogen in future.

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Conflict of Interest: The authors declare that there is no conflict of interests.

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