

## Biological Control of Fusarium Wilt of *Cajanus cajan*

Pushpa. H<sup>1</sup>, Swetha. P<sup>2</sup>, Vishal. M<sup>3</sup> and Vidyashree. M<sup>4</sup>

<sup>1</sup>Department of Microbiology, Ramaiah College of Arts, Science and Commerce, Bengaluru

<sup>2</sup>Forest Protection Division, Institute of wood science and technology, Bengaluru

<sup>3</sup>Department of Studies and Research in Microbiology, Sahyadri Science College,

Kuvempu University, Shivamogga, India

<sup>4</sup>Tungal PU Science college, Bijapur, India

### ABSTRACT

*Cajanus cajan* (L.) Millsp. commonly called Pigeon Pea a leguminous plant grown extensively for food, feed, fodder, fuel also grown as an intercrop and in crop rotation to improve the fertility of the soil in sustainable organic farming. This plant gets infected by the pathogen *Fusarium udum* causing wilt disease which is one of the major constraints in the production and productivity of pigeon pea. This pathogen is reported throughout the world infecting *C. cajan*. The extensive use of chemical fungicides results in environmental pollution, the resistance of pathogens towards fungicides, hazardous to humans and animals. This necessitated the need to adopt eco-friendly and sustainable management of diseases, like using antagonistic fungi against the pathogen. In our present study, 15 different fungal isolates were isolated from different rhizosphere soil and used for Our present study involved the in vitro - dual culture assay as well as in vivo - greenhouse bioassay was performed to analyze the antifungal efficacy of antagonistic fungi against *F. udum*. The *in vitro* and *in vivo* investigations showed that *Cephalosporium acremonium*, *Lasiodiplodia pseudotheobromae*, *Penicillium frequentans* and *Epicoccum sorghinum* equally inhibited *F. udum* when compared to various *Trichoderma* spp. Also, under greenhouse conditions, the root length, shoot length, and the number of leaves of plants was found to be increased significantly ( $p \leq 0.05$ ) in treatments with the talc-based biopesticide formulations of the antagonistic fungi. Hence these fungal isolates can also be used as a potential biocontrol agent for sustainable wilt diseases management caused by *Fusarium udum* and the extensive usage of chemical fungicides can be avoided to control the wilt disease of *Cajanus cajan*.

**KEY WORDS:** BIOCONTROL, EPICOCUM SORGHINUM, FUSARIUM SP., LASIODIPLODIA PSEUDOTHEOBROMAE, WILT DISEASE.

### INTRODUCTION

*Cajanus cajan* is an important constituent in the category of pulses among all Indians due to the availability of

#### Article Information:

\*Corresponding Author: [pushpa\\_microbio@msrcasc.edu.in](mailto:pushpa_microbio@msrcasc.edu.in)

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20–21% protein providing an energy-rich cereal diet. According to FAO statistics, this crop is cultivated in an area covering 4.6 million hectares globally and India accounted for about 73% of the global production in the year 2007. It is also an important crop of Karnataka contributing about 18% and 12% to total area and production respectively (GOI, Agricultural statistics, 2013). This protein-rich legume is prone to a multitude of diseases of which fungal diseases lead to a productivity loss of approximately 22 – 25 % each year amounting to a loss of up to Rs. 50,000 crores annually. Among the fungal diseases, *Fusarium* wilt disease caused by a soil-borne pathogen *Fusarium udum* (Fu) Butler is associated with extensive yield losses of pigeon pea in India. *Fusarium*

species are common soil inhabitants that usually infect through roots and penetrates the vascular system of plants, causing wilt and substantially reducing yields due to their ability to survive in soil, plant roots, and dead plant debris for several years Saremi and Burgess (2000).

There is an increasing demand for controlling crop diseases in an eco-friendly way by using biological pesticides rather than the traditional way of using chemical pesticides. The extensive use of chemical pesticides poses a negative impact on the environment as well as humans. They also are toxic to normal microflora of the rhizosphere Molli et al, (2015). Therefore, the use of biocontrol agents is promising in protecting the rhizosphere or spermosphere by inhibiting the pathogen (Marx, 1972) and by competing with the pathogen for limiting the growth nutrients (Chet, 1979; Couteaudier and Alabouvette, 1990). Scientists have studied the mechanism of biological control by antagonistic fungi and bacteria to control plant pathogens extensively in the past two decades (Janisiewicz et al. 2000). In our present study, we explore the possibilities to isolate a potentially promising biocontrol agent against wilt disease of pigeon pea.

## MATERIAL AND METHODS

For the isolation of pathogen from wilt infected *C. cajan*, the wilt infected plant samples were collected from the pigeon pea field of GVK, Bangalore. Isolation of fungi from diseased root and leaves samples was done by standard tissue isolation method (Aneja, 2003). The isolated pathogen was subjected to pathogenicity assay by sick soil method to confirm the pathogenicity of the pathogen (Dehariya et al. 2015; Sumitra 2006); Nikam et al., 2011; Arunodhayam et al., 2014). The isolated pathogen was identified by morphological characteristics and the pure cultures of the pathogen were maintained on PDA slants and stored at 4°C (Leslie and Summerell, 1991).

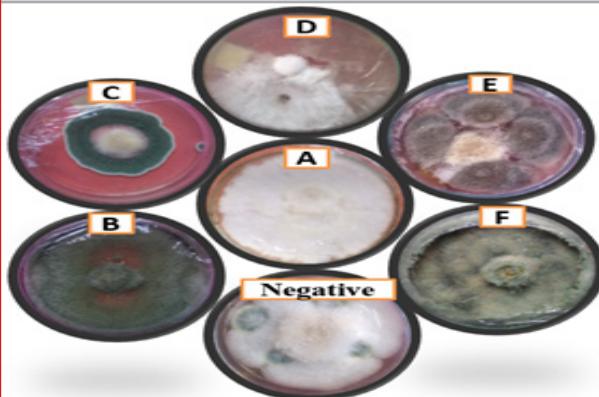
**Figure 1:** Isolated Pure culture and Microscopic view of *Fusarium udum*



For the isolation of antagonistic fungi, rhizosphere soil of wilt infected pigeon pea was collected. The obtained samples were processed and plated by a serial dilution method and pour plate technique (Aneja, 2003). All the plates were incubated for 5- 7 days at 21-24°C. The pure culture of the fungal isolates was prepared, and

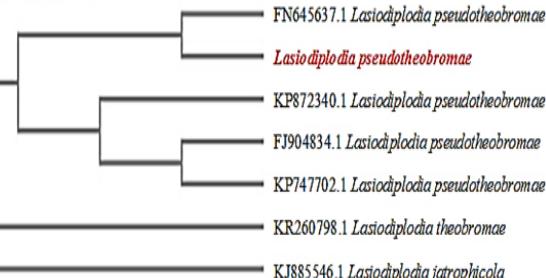
they were identified using morphological characters up to species level using the keys as proposed by Joseph Gilman (1950) and Barnett (1998). The isolates that were not able to identify through morphological characteristics were subjected to molecular characterization.

**Figure 2:** Dual culture method (Representative plates): A: control, B: *Trichoderma* sp.-isolate I, C: *Penicillium frequentans* D: *Lasiodiplodia pseudotheobromae* E: *Epicoccum sorghinum* F: *Cephalosporium acremonium*. Negative: *Penicillium frequentalis*.



For the molecular characterization of Isolate-1, the pure cultures of Isolate-1 were obtained by placing the hyphal tip of the mycelia aseptically on sterilized Potato Dextrose Broth and was incubated at 24°C for 6-7 days. The extraction of genomic DNA from the pure cultures of both the isolates was performed by - Cetyl trimethyl ammonium bromide (CTAB) method (Ausubel et al., 1994). The Polymerase chain reaction of the obtained DNA was performed with universal primers for fungal genome, namely ITS 1 - (5'-TCC GTA GGT GAA CCT GCC G-3') forward ITS 4 - (5'- TCC GCT TATTGATAT GC-3') reverse primers. These primers were obtained from Chromous Biotech Pvt. Ltd. Bangalore, India. ITS region of rDNA was visualized by UV trans-illumination (352 nm) and the expected DNA band was excised from the gel using a sterile scalpel and placed into a 1.5 ml microtube.

**Figure 3:** Cladogram of *Lasiodiplodia pseudotheobromae*



This DNA was purified using gel extraction kit (Chromous Biotech Pvt. Ltd. Bangalore, India) according to the manufacturer's specifications. The purified PCR product was sequenced at Chromous Biotech Pvt. Ltd.

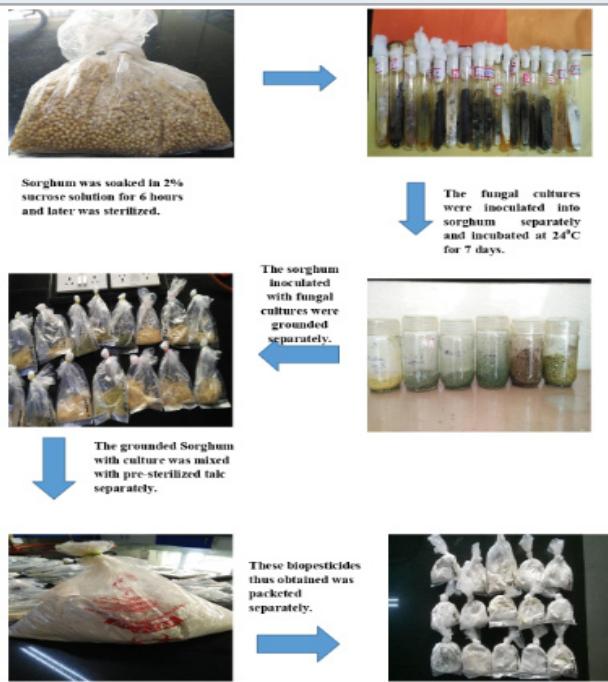
Bangalore, India. Sequences were determined by the chain termination method using an ABI 3130 Genetic Analyzer. Sequencing was done in the forward and reverse direction. The sequence was generated using data analysis software (Seq Analysis\_v 5.2). The obtained rDNA gene sequence was used to carry out BLAST with the database of the NCBI gene bank. Based on maximum identity scores first 10 sequences were selected and aligned using Multiple sequence alignment software program Clustal W2 to prepare Cladograms for the analyzed species. The rDNA sequence of an isolate was deposited in NCBI Genbank.

For the *in vitro* screening of antagonistic fungi against the pathogen, the dual culture method (Skidmore and Dickinson, 1976) was followed to analyze the degree of antagonism exhibited by 15 fungal isolates against the pathogen. The inoculated sets were incubated at 28°C for 7 days. Triplicates of each test plate were maintained. The degree of antagonisms of each bio-agent against the test pathogen in dual culture was graded on a scale of 1-5 as proposed by Bell et al., (1982) and the percent inhibition of radial mycelial growth of the pathogen was calculated using the following formula given by Singh et al., (2002).

$$I = [(C-T)/C] \times 100$$

Where, I- the percent inhibition  
C - colony diameter in the control plate and  
T- the colony diameter of the pathogen

Figure 4: In vitro production of bio pesticides



For the preparation and biocontrol control studies of talc based biopesticides, antagonistic isolates, exhibiting a high degree of antagonism against the pathogen in

*vitro* conditions, were chosen for preparing talc-based biopesticides. Solid substrate fermentation was followed for the production of talc-based biopesticide (Mukesh et al., 2015) (Fig 04). The efficacy of the prepared biopesticides was studied to control the wilt disease of pigeon pea. It was tested by sowing pigeon pea seeds into artificially made pathogen infested soil as described by (Bell et al., 1982). 2g each of the 12 prepared talc powder-based biopesticide was added to each pot containing pigeon pea plants with the inoculation of *F. udum* in triplicates. The number of leaves, root and, shoot length was measured in each pot along with negative and positive control and it was subjected to statistical analysis.

Figure 5: In vivo studies: A: Triplicates of Pigeon pea plant; B: Representative figure-Trichoderma viridae.



Table 1. Percent (%) Inhibition of growth of *Fusarium udum* and *Fusarium oxysporum* f. sp lycopersici by different fungal isolates and Grade Assessment for the interaction between the pathogen and antagonistic fungi

Antagonistic organisms	Grades	% inhibition of radial growth of fungi <i>Fusarium udum</i>
<i>Aspergillus candidus</i>	3	78.82±0.02 <sup>i</sup>
<i>Aspergillus flavus</i>	1	78.52±0.02 <sup>j</sup>
<i>Aspergillus niger</i> -isolate I	1	78.51±0.02 <sup>f</sup>
<i>Aspergillus niger</i> -isolate II	1	78.22±0.01 <sup>g</sup>
<i>Cephalosporium Acremonium</i>	1	88.22±0.02 <sup>c</sup>
<i>Cladosporium</i> sp.-isolate I	5	11±0.10 <sup>l</sup>
<i>Cladosporium</i> sp.-isolate II	4	33.33±0.01 <sup>k</sup>
<i>Epicoccum sorghinum</i>	1	72.65±0.00 <sup>h</sup>
<i>Lasiodiplodia pseudotheobromae</i>	1	83.76±0.01 <sup>d</sup>
<i>Penicillium frequentalis</i>	1	0±0.0 <sup>m</sup>
<i>Penicillium frequentans</i>	5	82.63±0.02 <sup>e</sup>
<i>Trichoderma harzianum</i>	1	89.66±0.01 <sup>a</sup>
<i>Trichoderma</i> sp.-isolate I	1	78.25±0.02 <sup>g</sup>
<i>Trichoderma</i> sp.-isolate II	1	88.52±0.02 <sup>b</sup>
<i>Trichoderma viridae</i>	1	83.74±0.03 <sup>d</sup>

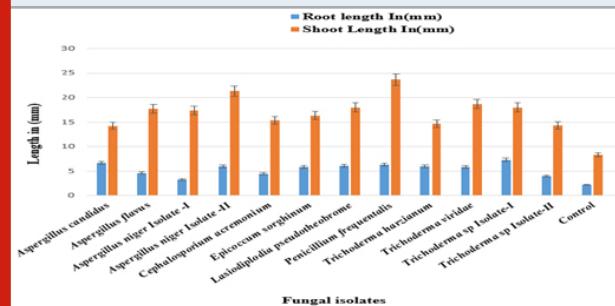
Means (n=3) in each column followed by the same letter are not significantly different ( $p<0.05$ ) from each other according to Duncan's multiple range test (DMRT)

For the data collection and statistical analysis, the data of the results obtained from in vitro and in vivo studies were tabulated and these were subjected to statistical analysis by using MSTAT-C (Version 1.41) and Microsoft excel software. Analysis of Variance (ANOVA) of differences in the treatments and the least significance tests were

carried out. Significance was evaluated at  $P<0.05$  for the tests. For each factor analyzed, the means of the different treatments were compared and ranked using Duncan multiple range test at  $P < 0.05$  using MSTAT-C (Version 1.41). The graphs were prepared using Microsoft Excel software.

## RESULTS AND DISCUSSION

**Figure 6:** Efficacy of fungal formulations on *Fusarium* wilt of *Cajanus cajan* (Pigeon pea) under greenhouse conditions (Root and shoot length)



Pigeon pea plants exhibited symptoms of wilt disease showing mild chlorosis followed by leaf abscission before succumbing to the disease. The pathogen isolated was identified as *F. udum* (Fig 01) and its pathogenicity was proved by reisolating the pathogen from the artificially infected plants. Similarly, Dehariya et al., 2015 performed the pathogenicity assay for *F. udum* and successfully reisolated the pathogen from the diseased pigeon pea plant. Following this antagonistic fungal species were isolated from the rhizosphere soil samples were identified till species level based on morphological characters.

**Table 2.** Efficacy of fungal formulations on *Fusarium* wilt of *C. cajan* under greenhouse conditions

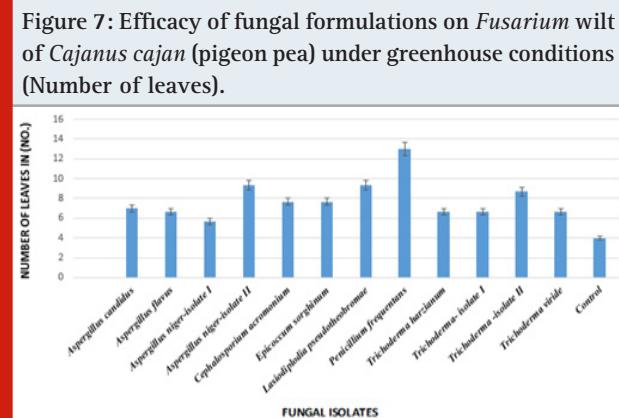
Biocontrol agents	Root length in (mm)	Shoot length in (mm)
<i>Aspergillus niger</i> Isolate-I	$3.33 \pm 0.849837^{\text{de}}$	$17.33 \pm 1.69^{\text{bc}}$
<i>Cephalosporium acremonium</i>	$4.50 \pm 0.408248^{\text{bcd}}$	$15.33 \pm 0.47^{\text{c}}$
<i>Epicoccum sorghinum</i>	$5.83 \pm 1.545603^{\text{abc}}$	$16.33 \pm 1.88^{\text{bc}}$
<i>Aspergillus niger</i> Isolate-II	$5.93 \pm 2.192918^{\text{abc}}$	$21.33 \pm 6.34^{\text{ab}}$
<i>Lasiodiplodia pseudotheobrome</i>	$6.07 \pm 0.492161^{\text{ab}}$	$18.00 \pm 2.16^{\text{bc}}$
<i>Trichoderma harzianum</i>	$6 \pm 2.160247^{\text{abc}}$	$14.67 \pm 3.68^{\text{c}}$
<i>Penicillium frequentans</i>	$6.33 \pm 1.247219^{\text{ab}}$	$23.67 \pm 3.39^{\text{a}}$
<i>Trichoderma</i> sp. Isolate-I	$7.33 \pm 1.649916^{\text{a}}$	$18.00 \pm 2.16^{\text{bc}}$
<i>Trichoderma</i> sp. Isolate-II	$4.00 \pm 0.816497^{\text{cde}}$	$14.33 \pm 4.92^{\text{c}}$
<i>Aspergillus flavus</i>	$4.6 \pm 0.432049^{\text{bcd}}$	$17.67 \pm 4.18^{\text{bc}}$
<i>Aspergillus candidus</i>	$6.67 \pm 1.545603^{\text{ab}}$	$14.23 \pm 3.02^{\text{c}}$
<i>Trichoderma viridae</i>	$5.83 \pm 0.849837^{\text{abc}}$	$18.67 \pm 0.47^{\text{abc}}$
Control	$2.23 \pm 0.20548^{\text{e}}$	$8.37 \pm 0.26^{\text{d}}$

Means ( $n=3$ ) in each column followed by the same letter are not significantly different ( $p<0.05$ ) from each other according to Duncan's multiple range test (DMRT)

Isolate-1 which could not be identified based on morphological characters was subjected to molecular characterization for precise identification of organism till species level (Pushpa et al., 2014). Isolate-1 was identified as *Lasiodiplodia pseudotheobromae* as it showed 99% similarity with the sequence of *L. pseudotheobroma* deposited in NCBI- Genbank with the accession numbers - KP747702.1, KP872340.1, FN645637.1 and FJ904834.1. The phylogenetic position confirms that our isolate corresponds to *L. pseudotheobromae* and that it would have been evolved from *L. theobromae* as per the cladogram obtained (Fig 03). The isolate was deposited in Genbank with

the accession number: KX289694.1. Dual culture assay (Fig 02, Table 01) and Duncan's Multiple Range Test showed that 12 fungal isolates i.e., *T. viridae*, *T. harzianum*, *Trichoderma* sp.-isolate I, *Trichoderma* sp.-isolate II, *L. pseudotheobromae*, *E. sorghinum*, *A. niger*-isolate 1, *A. niger*-isolate II, *A. flavus*, *A. candidus*, *P. frequentans*, *C. acremonium* displaying class-1 and positive antagonism with percent of growth inhibition in the range of 72.66%-89.66%. While, *P. frequentans*, *Cladosporium* sp.-isolate I, *Cladosporium* sp.-isolate II showed poor inhibition rates of 33.33%, 11% and 0% respectively displaying class- 5 negative antagonism.

Greenhouse experiments revealed that *A. candidus*, *A. flavus*, *Trichoderma* sp. isolate-I on infected plants showed a moderate growth of root, shoot and number of leaves. Whereas, *A. niger*-isolate I, II, *C. acremonium*, *E. sorghinum*, *L. pseudotrichobrome*, *P. frequentans*, *T. harzianum* *Trichoderma* sp. -isolate-II, *T. viridaes* showed highest growth of root and shoot while it showed moderate growth of number of leaves (Fig 05, 06, 07; Table 02).



It can be concluded that the antagonistic organisms have an ability to produce zones of inhibition, which can be attributed for the production of various secondary metabolites. Similar studies have been reported by various investigators for the production of zones of inhibition by antagonistic fungi around the pathogen and the increasing concentrations of antifungal and fungistatic metabolites lead the dominating antagonistic organisms to overgrow the pathogens, thereby suppressing their growth (Royse and Ries, 1977; Odigie and Ikotun, 1982). Several scientists have worked on the in vitro screening of different strains of *Trichoderma viride* and *T. harzianum* against *F. udum* and proved significant inhibition of growth of various isolates of *F. udum* (Patel et al, 2011).

## CONCLUSION

The use of fungi as biological control agents has found its deserving slot in the sustainable management of a spectrum of fungal and bacterial diseases in recent years. The use of biocontrol agents to combat fungal diseases has been magnified recently, due to the increasing negative environmental impact of hazardous chemical pesticides. The present investigation provides a detailed insight on the use of biological means for managing the wilt pathogen of *C. cajan* caused by *F. udum* both in vitro and in vivo. Though moderate growth inhibition activity was exhibited by *L. pseudotrichobrome* and *E. sorghinum*, it was found that they could vigorously grow and overtake or suppress the growth of both the pathogens similar to that exhibited by all *Trichoderma* sp. which is referred to as hyper parasitism. These evidences prove that the wilt disease could be considerably reduced by the use of talc formulations of fungal antagonists. And also, that biocontrol agents other than *Trichoderma* sp. can be scaled-up for talc-based formulations and

mass production of fungal biopesticides for controlling *Fusarium* wilt diseases of *C. cajan*.

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## REFERENCES

- Aneja KR (2003) Experiments in Microbiology, Plant pathology and biotechnology. New age international (P) limited Publishers.4th Edition: 1-605.
- Anil Kumar R and Raj Kumar H Garampalli (2014) Antagonistic effect of rhizospheric *Aspergillus* species against *Fusarium oxysporum* f. sp. lycopersici, Research Article. International Journal of Chemical and Analytical Science.5 (1):39-42.
- Arunodhayam K, Eswara Reddy N P and Madhuri V (2014) Pathogenicity and management of *Fusarium* wilt of chickpea, *Cicer arietinum* L.-A review. Current Biotica 7(4): 343-358.
- Ausubel FM, Brent R, Kingston RE, Moore, D.D., Seidman, J.G., Smith, J.A. and Struhl, K (1994) Current Protocols in Molecular Biology. New York City, NY: John Wiley & Sons Inc.
- Barnett, H.L. and Hunter, B.B. (1998) Illustrated Genera of Imperfect Fungi. 4th Edition, APS Press, St. Paul, 218 pp
- Bell DK, Wells HD, Markham CR (1982) In vitro antagonism of *Trichoderma* species against six fungal plant pathogens. Phytopath. 72: 379-382.
- Bohl Stricker K, Harmon PF, Clay K, Goss EM and Flory SL (2014) Widespread independent emergence and accumulation of fungal pathogens reduce fecundity of an invasive grass Direct Submission Submitted University of Florida, Gainesville, FL32611, USA. GenBank database. Published only in GenBank database.
- Chen ZM, Fan QY, Chen HP, Li ZH, Feng T and Liu JK (2014) A novel C25 sterol peroxide from the endophytic fungus *Phoma* sp. Submitted School of Chemistry and Chemical Engineering, Lingnan Normal University, 29 Cunjin Road, Zhanjiang,
- Chet I, Hadar Y, Elad Y, Katan J and Henis Y (1979) Biological control of soil-borne pathogens by *Trichoderma harzianum*. In: Soil Borne Plant Pathogens (B. Schippers ed.), Academic press, London, UK 585.
- Couteaudier Y and Alabouvette C (1990) Survival and inoculum potential of conidia and chlamydospores of *Fusarium oxysporum* f. sp. lini in soil. Canadian Journal of Microbiology. 36: 551-556.
- Dehariya K, Shukla A, Sheikh I A, and Vyas D. (2015) *Trichoderma* and Arbuscular Mycorrhizal Fungi Based

- Biocontrol of *Fusarium udum* Butler and Their Growth Promotion Effects on Pigeon Pea. J. Agr. Sci. Tech. 17: 505-517.
- Fernandez D Calvino, Pedro Soler-Rovira, and Cesar Plaza. (2010) Enzyme activities in vineyard soils long-term treated with copper-based fungicides. Soil Biology and Biochemistry. 42(12):2119-2127.
- Janisiewicz WJ, Tworkoski TJ and Sharer C (2000) Characterizing the mechanism of biological control of postharvest diseases on fruits with a simple method to study competition for nutrients. Phytopathology. 90(11):1196-2000.
- John F Leslie and Brett A Summerell (2006) The Fusarium Laboratory Manual, Photographs by Suzanne Bullock. Blackwell Publishing: 1-369.
- Joseph C Gilman. (1950) A manual of Soil Fungi, Botany department, IOWA state college, The Iowa State College Press, Ames, Iowa, USA: 1-390.
- Konovalova OP (2015) Direct Submission Submitted Faculty of Biology, Moscow State University, Soyuznii street, Moscow, Moscow 111396, Russian Federation. GenBank database. Published only in GenBank database.
- Leite TS, Fassoni AC, Pereira OL, Mizubuti EG, Araujo EF and Queiroz MV. Diversity of endophytic fungi in two soybean cultivars using two different methods for isolation, GenBank database. Published only in GenBank database.
- Leslie JF, Summerell BA (1991) The Fusarium Laboratory Manual. Blackwell publications.388p
- Li Q, Huang S, Guo T and Mo J. (2015) Molecular phylogenetics of mango gummosis-causing agents in Guangxi, Submitted by Institute of plant protection, Guangxi Academy of Agricultural Sciences, 174 Daxue Road, Nanning, Guangxi 530007, China. GenBank database. Published only in GenBank database.
- Lin Z, Que Y and Zhang M (2013) Phoma species associated with sugarcane twisting and curling of crown leaves in china, Plant diseases. 98:850
- Loro M, Valero-Jimenez CA, Nozawa S and Marquez LM. (2012) Diversity and composition of fungal endophytes in semiarid Northwest Venezuela J. Arid Environ. 85:46-55.
- Marx DH (1972) Ectomycorrhizae as biological deterrents to pathogenic root infections. Annu Rev Phytopathol. 10:429-454.
- Molli M Newmana, Nigel Hoilettb, Nicola Lorenzc, Richard P Dickc, Mark R Lilesd, Cliff Ramsiere, Joseph W. Kloeppe, Editor: D. Barcelo (2016).Glyphosate effects on soil rhizosphere-associated bacterial communities, Sci Total Environ. Feb 1;543(Pt A):155-160
- Mukesh Srivastava, Vipul Kumar, Mohammad Shahid, Sonika Pandey and Anuradha Singh. (2016). *Trichoderma*- a potential and effective bio fungicide and alternative source against notable phytopathogens: A review. AJAR. 11(5): 310-316.
- Nikam PS, Jagtap GP and Sontakke PL. (2011) Survey, surveillance and cultural characteristics of chickpea wilt caused by *Fusarium oxysporum* f. sp. *ciceris*. African Journal of Agricultural Research.6 (7): 1913-1917.
- Njuguna JW, Barklund P, Ihrmark K and Stenlid J (2009) Direct Submission Submitted by Forest Pathology and Mycology, Swedish University of Agricultural Sciences, UllsVag 26A P.O. Box 7026, Uppsala SE-750 07, Sweden. GenBank database. Published only in GenBank database.
- Odigie EE, Ikotun T (1982) In vitro and in vivo inhibition of growth of *Phytophthora palmivora* by antagonistic microorganisms Fitopatologia Brasileira. 7: 157 -167.
- Patel SI, Patel RL, Desai AG and Patel DS (2011). Biocontrol of *Fusarium udum* through *Trichoderma*. Journal of Pharma and Bio Sciences. 2(4):215-222.
- Pushpa H, Purushothama K B, Ramesh Thirumalesh D H (2012). Taxonomic studies and molecular characterisation of *Tricholomagiganteum* and *Calocybe indica* isolates from Bangalore.J Biochem Tech. 3(5): S218-S220.
- Royse DJ, Ries SM (1977). The influence of fungi isolated from peach twigs on the pathogenicity of *Cytosporacinata*.*Phytopathol*. 63: 603-607.Sambrook J. and Russell DW. (2001) Molecular Cloning. A laboratory manual. 3rd ed. CSHL press, NY, USA.
- Saremi H and Burgess LW (2000). Effect of Soil Temperature on Distribution and Population Dynamics of *Fusarium* Species. J. Agr. Sci. Tech. 2: 119-125.
- Shuttleworth LA, Roux J, Slippers B and Wingfield MJ (2015) Lasiodiplodia sp. associated with native Myrtaceae in southern Africa, description of two species, and evidence of hybridisation. Submitted by Forestry & Agricultural Biotechnology Institute, University of Pretoria, FABI Ring Road, Hatfield, Pretoria, Gauteng 0002, South Africa GenBank database. Published only in GenBank database.
- Silva-Rojas HV, Cortes-Cueto AL and Molina-Gayosso E (2012). Identification of culturable fungi associated with declining sugarcane plants based on genealogical analyses Submitted Produccion de Semillas. Proyecto Sagarpa-Conacyt, Colegio de Postgraduados, Km. 36.5 carretera Mexico-Texcoco, Texcoco, Mexico 56230, Mexico. GenBank database. Published only in GenBank database.
- Singh R, Singh BK, Upadhyay RS, Rai B, Lee YS (2002) Biological control of *Fusarium* wilt disease of pigeonpea. Plant Pathol. J., 18: 279-283
- Skidmore AM, Dickinson CH. (1976). Colony interactions

and hyphal interference between *Septoria nodorum* and phylloplane fungi. Trans.Brit. Mycol. Soc. 66: 57-64.

Sumitra Patil Kulkarni (2006) Studies on *Fusarium oxysporum* Schlecht Fr f. sp. *gladioli* (Massey) Snyd. And Hans. causing wilt of gladiolus. Ph.D Thesis submitted to the University of Agricultural Sciences, Dharwad.

Thilagam R, Kalaivani G and Hemalatha N (2018) Isolation and identification of phytopathogenic fungi from infected plant parts. International Journal of Current Pharmaceutical Research. 10. 26.

Zhao JP, Lv Q and Zhang XY. (2009) *Lasiodiplodia*

*pseudotrichobromae*, a new record of pathogenic endophytic fungi in southern China, Submitted by Zhao J.P., Forestry College, Hebei Agricultural University, Southroad of Lucky Street, Baoding, Hebei 071000, China. GenBank database. Published only in GenBank database.

Zhu H, Qin WQ, Liu L and Yan W (2015) First Report of Leaf Spot of Clustering Fishtail Palm (*Caryota mitis*) Caused by *Lasiodiplodia jatrophicola* in China, Plant Dis. 99 (7), 1038. GenBank database. Published only in GenBank database.