

Evaluation of Inhibitory Activity of Bacteriocins from *Enterococcus italicus* BLN48 Against *Mycobacterium fortuitum* and its Toxicity Profiling

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ABSTRACT

Lactic Acid Bacteria (LAB) is an important group of microorganism due to their wide application in the food and dairy industries. They have extensively studied for the potential against various bacterial pathogens. The inhibitory activity of LAB is through their production of bacteriocins, organic acids, enzymes, hydrogen peroxide, etc., Bacteriocins are ribosomally synthesized antimicrobial peptides produced by various microorganisms. The incidence of non tuberculous mycobacterial infections increasing worldwide. *Mycobacterium fortuitum* is one of the rapidly growing non tuberculous mycobacteria which causes skin, bone, joint and pulmonary infections. Their antibiotic resistance and prolonged course of treatment necessitates the development of new candidate to fight against them. In this study, we have partially purified bacteriocins from four LAB strains using solvent extraction method and screened their activity against *M. fortuitum* MTCC1902 by colony forming unit (CFU) estimation method. The potential strain was identified by 16S rRNA sequencing and their sequences were submitted to Genbank database. The toxicity of potential LAB strain was assessed by both in vitro and in vivo method against Vero cell lines and zebra fish model respectively. The strain BLN48 showed 97.9% reduction in growth of *M. fortuitum* and 1.89 ± 0.32 log reduction in CFU/ml. The potential strain BLN48 was identified as *Enterococcus italicus*. *E. italicus* BLN48 exhibited cytotoxicity against vero cell lines in dose dependent manner whereas under in vivo conditions, 50% of zebrafish larvae survived upto 144 hours post fertilization (hpf) with normal morphological changes. Further purification and characterization of *E. italicus* BLN48 bacteriocin in future helps in the development of an efficient candidate against *M. fortuitum* as well as other mycobacterial pathogens.

KEY WORDS: PARTIALLY PURIFIED BACTERIOCIN, NON TUBERCULOUS MYCOBACTERIA, MYCOBACTERIUM FORTUITUM, CYTOTOXICITY, ZEBRAFISH.

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INTRODUCTION

Lactic Acid Bacteria (LAB) remains an industrially important group of microorganism due to their wide application in the food and dairy industries (Gomez et al., 2015). LAB are widely distributed in diverse habitats like marine, food products, dairy origin, etc.,. Usually, fermented foods were screened for bacterial isolates with antimicrobial properties as its microbiota is dominated by LAB. LAB also exists in gastrointestinal tracts, oral cavities of humans as well as animals (Bungenstock et al., 2020; Li et al., 2020). In raw milk, LAB are the predominant microorganisms and they contribute to the fermentation and food preservation process due to their various metabolite production (Rahmeh et al., 2019). LAB has studied extensively for their antagonistic activity against various bacterial pathogens such as food spoiling microorganisms like *Listeria monocytogenes*, gastrointestinal pathogens and other various gram positive and gram negative bacterial pathogens. Researchers have gained significant attention towards LAB due to their Generally Recognized As Safe (GRAS) status. The inhibitory activity of LAB is mainly through their production of various substances like organic acids, enzymes, bacteriocins, hydrogen peroxide, etc., (Gupta and Garg 2009; Rodrigues et al., 2006 Hussein et al., 2018).

Bacteriocins are ribosomally synthesized antimicrobial peptides produced by various microorganisms. The uses of bacteriocins in functional foods and as an alternative to antibiotics are their emerging application (Quwehand et al., 2004; Messi et al., 2001). Physical stability and non-toxic nature are the major advantages of bacteriocins (Morgan et al., 2005). Different classes of bacteriocins exerts diverse mechanisms against their target like disruption of cell wall, pore formation, inhibition of protein and nucleic acid synthesis, etc., (Cascales et al., 2007; Stevens et al., 1991). LAB produces a diverse nature of bacteriocins in different size, physicochemical properties, spectrum of activity, structures, etc. They secreted in an extracellular space during LAB growth (Venegas et al., 2019; Anbarasu et al., 2020). The crude and purified bacteriocins from LAB have found their potential use as biopreservative agents to enhance the quality and safety of various food products. The potential bacteriocins can be used in both combined and balanced mode as probiotics for human diseases also (Iseppi et al., 2019, Arrijoja et al., 2020).

Non Tuberculous Mycobacteria (NTM) are several mycobacterial species other than *M. tuberculosis* complex and *M. leprae*. NTM causes opportunistic infections in humans as well as animals and also it transmitted among environment, livestock, wildlife, etc. (Odoi et al., 2020). *Mycobacterium fortuitum* is one of the rapid growing NTM and is predominantly found in water systems like natural water, tap water, and water used in showers in hospitals and soil. It mainly causes skin, bone, joint infections and pulmonary diseases in immunocompromised and immunosuppressed patients. They also causes surgical site infections

(Okamori et al., 2018; Griffith et al., 2007; Goslee and Wolinsky 1976; Wolinsky and Rynearson 1968; Choudhary et al., 2020).

M. fortuitum are often isolated from skin and soft tissues and also from other clinical samples as it causes many types of infection (Garcia et al., 2020). The development of antibiotic resistance and prolonged course of treatment with multiple antibiotics in NTM infection surges the need for the development of new candidates with potential inhibitory substance and less toxicity to fight against these infections. In this study, we have evaluated bacteriocins from four LAB isolates for their anti *M. fortuitum* activity. The potential LAB isolate was identified through 16S rRNA sequencing and phylogenetic analysis. Toxicity profile of the bacteriocins from the potential LAB strain was also evaluated through in vitro and in vivo method.

MATERIAL AND METHODS

Mycobacterium fortuitum MTCC1902 strain was purchased from Microbial Type Culture Collection (MTCC), Chandigarh, India. de Man Rogosa Sharpe broth (Himedia), Middlebrook 7H9 broth (Himedia), Middlebrook 7H11 Agar (Himedia), Chloroform (SRL), PBS tablets (Sigma) were used in the study. Four lactic acid bacterial strains viz., BLN 34, BLN 36, BLN 39 and BLN 48 previously isolated from different cow milk samples were used in this study. Viability of all the cultures were maintained in MRS agar slants at 4°C (Revathy et al., 2019). Bacteriocin from four selected cultures was produced by submerged fermentation process and was partially purified by solvent extraction method using chloroform as described by Burianek et al., 2000 with few modifications. Briefly, 5ml of overnight grown culture in MRS broth was added to 500ml of sterile MRS broth and incubated for 18 hours in shaking incubator at 30°C. After incubation, the culture was centrifuged at 5000rpm for 10minutes to collect the cell free supernatant. For the extraction of crude bacteriocin, 50% v/v of chloroform has been added to the supernatant and kept in magnetic stirrer at 1000rpm for 20minutes. Then the mixture was subjected to centrifugation at 10000rpm for 30 minutes. After centrifugation, the precipitate in the interphase layer between solvent and aqueous phase was collected carefully and freed at -20°C following by lyophilization. The lyophilized form of partially purified bacteriocin (PPB) were stored at -20°C and used for further assays.

Evaluation of PPB against *M. fortuitum*: Ten mg concentration of partially purified bacteriocin in the form of lyophilized powder was dissolved in 1ml of PBS buffer in order to get 10mg/ml (w/v) concentration. Desired working concentration of PPB was prepared from the main stock using PBS buffer. Inhibitory activity of PPB prepared from all the four cultures was evaluated against *M. fortuitum* by colony forming unit (CFU) estimation (Gillespie et al. 2005). Briefly, *M. fortuitum* MTCC1902 suspension was prepared by inoculating a loopful of *M. fortuitum* culture into 0.3ml of Middlebrook 7H9

broth followed by vortexing. Then the volume of the suspension was made upto 5ml. using 7H9 broth. In a sterile cryovial, 400µl of 7H9 broth was used as growth control and 350µl of 7H9 broth with 50µl of bacteriocin was used in test vial. All the vials were added with 100µl of *M. fortuitum* suspension and incubated at 37°C for 48 hours. After incubation, 100µl of aliquot from each vial was serially diluted in 900µl of PBS buffer upto 10-12 dilution. 50µl of the dilution was spreaded onto Middlebrook 7H11 agar plate. Plating was done in triplicate and all the plates were incubated at 37°C for 48 hours. The plates with individual colonies were taken for the calculation of CFU/ml.

Taxonomy of potential LAB strain: The genomic DNA of LAB strain BLN48 was isolated using solute ready genomic DNA kit. DNA was analyzed by gel electrophoresis and quantified using a spectrophotometer (NanoDrop ND-1000, Thermo Scientific, Gloucester, UK). The 16S rRNA gene sequence of the strain was amplified using the primers: 27F 5'AGAGTTTGATCMTGGCTCAG3' (forward) and 1492R 5'TACGGYTACCTGTTACGACTT3' (reverse) (Kumar Gothwal et al., 2007). The PCR amplified product of the strain was sequenced and analyzed at National Collection of Industrial Microorganisms (NCIM), National Chemical Laboratory (CSIR-NCL), Pune, India. The 16S rRNA gene sequence obtained from the strain BLN48 was aligned with similar sequences available in GenBank using MEGA 7 program. The aligned sequences of the strain BLN48 was used to construct the phylogenetic tree by following neighbor joining algorithm in MEGA 7 program (Saitou and Nei, 1987). The bootstrap estimation (Felsenstein, 1985) was used to determine the confidence of the branches of the phylogenetic tree. The partial 16S rRNA nucleotide sequence of all the four strains has been deposited in GenBank database.

In vitro toxicity analysis of PPB: The cytotoxicity of potential PPB which showed activity against *M. fortuitum* were assessed in vitro by adopting MTT (3-(4, 5-dimethyl thiazol-2yl)-2, 5-diphenyl tetrazolium bromide) assay using Vero cell lines. Briefly, in the 96 well plate, 100µl of RMPI 1640 medium was added with 100µl of desired concentration of PPB. Then 200µl of total volume is gently mixed well. 100µl of diluted PPB from the first well was serially diluted in next well till reaching the lowest concentration. The cultured Vero cell lines were harvested by trypsinization and pooled in 50ml vial. Then the cells were plated at a density of 1x100cells/ml. 200µl of vero cells without PPB was used as a control. The cells were incubated at 37°C in 5% CO2 incubator for 24 hours. After incubation, 20µl of MTT solution was added to all the wells and incubated for 4 hours at 37°C. The media and MTT was well mixed and the absorbance was measured at 450nm and the percentage of viability was calculated manually (Vijayarathna and Sasidharan, 2012).

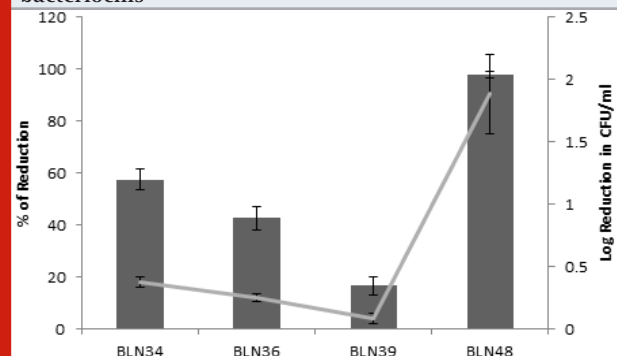
In vivo toxicity analysis of PPB: In vivo toxicity of PPB was evaluated using zebrafish as a model (Sisman et al. 2008). Zero day old zebrafish eggs were purchased from zebrafish aquarium in Kanchipuram

district, Tamil Nadu, India. Twenty healthy post hatched zebra fish eggs were transferred to the wells of a 24-well plate along with 1 ml of embryo water (60 mg of sea salt/litre of ultrapure water). Different concentrations of PPB of potential LAB strain BLN48 (10, 50 and 100 µg/ml) was added to the wells and incubated for 144 h at 28.5°C. Mortality of the zebra fish was noted after 24, 48, 96 and 144 h. The embryos appeared opaque and white in colour. The dead embryos were degraded soon, whereas the structures of intact embryos were more visible by 48 hours post fertilization (hpf) which allowed a clear distinction between the dead and alive. The mortality rate is calculated. At the end of the incubation period, the embryos were photographed using a light microscope at 10X magnification.

RESULTS AND DISCUSSION

Anti *M. fortuitum* activity: Among the four isolates tested, BLN48 showed significant inhibitory activity against *M. fortuitum* MTCC1902 viz., 1.89 log reduction of CFU/ml which corresponds to 97.9% reduction in growth when compared to growth control. Followed by, BLN34 showed slight inhibitory activity by 0.38 log reduction of CFU/ml with 57.56% reduction from growth control (Figure 1).

Figure 1: Anti *M. fortuitum* activity of Partially purified bacteriocins



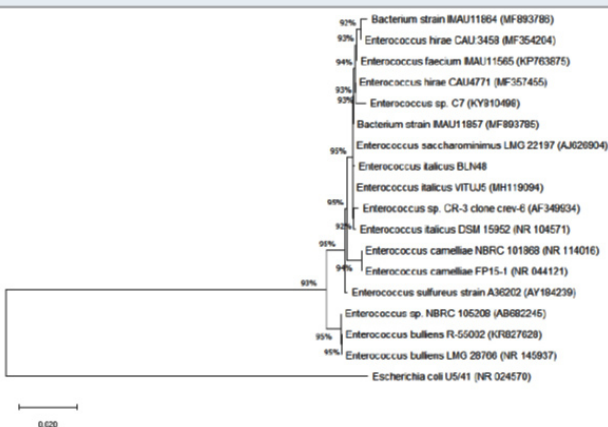
Taxonomy of potential LAB strain BLN48: Amplification of 16S rRNA gene from the strain BLN48 resulted in 1435 bp sequences. BLAST analysis showed 99.43% sequence similarity with 16S rRNA gene sequence of *Enterococcus italicus* DSM 15952. The phylogenetic tree also showed that the strain BLN48 is closely related to *Enterococcus italicus* (Figure 2). The nucleotide sequence of *E. italicus* BLN48 was submitted to Genbank with accession number MN880432.

Table 1: In vitro cytotoxicity analysis of BLN48 by MTT assay against vero cell lines

PPB	Test concentration	Percentage of Viability
BLN48	100mM	16.6
	10mM	60.4

In vitro toxicity analysis of PPB: The cytotoxicity analysis by MTT assay showed that the bacteriocin from BLN48 exhibited the cytotoxicity on vero cell lines in dose dependent manner. At the maximum of 60% of vero cells survived when treated at 10mM concentration of bacteriocin (Table 1).

Figure 2: Phylogeny of BLN48



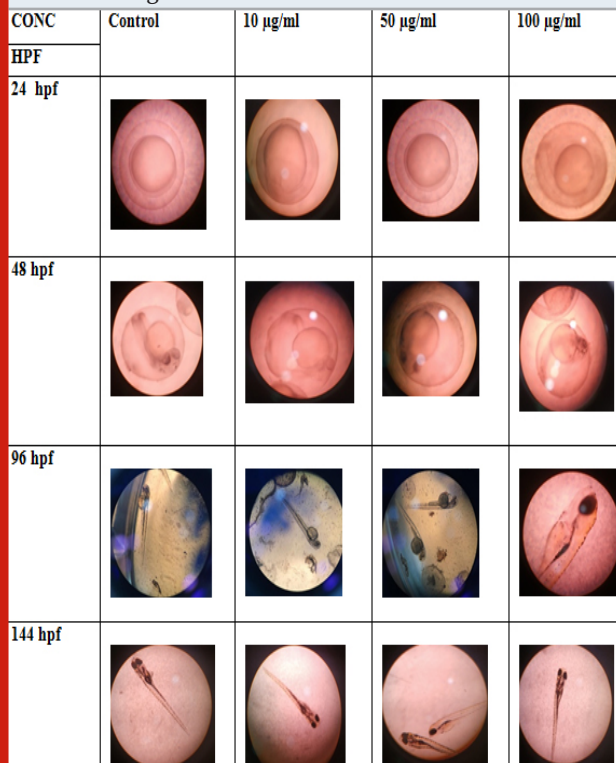
In vivo toxicity analysis of PPB: In the in vivo toxicity analysis of BLN48 done with zebrafish larvae, BLN48 have shown less toxicity viz., around 50% of larvae (compared to control) were survived upto 144 hpf at high concentration (100µg/ml) with healthy morphology under microscopic observation (Table 2). The features like fin movement, swimming nature, tail development are normal to the viable larvae (Figure 3).

Table 2. Mortality rate in in vivo toxicity analysis of BLN48

Concentration HPF	Control	10 µg/ml	50 µg/ml	100 µg/ml
0 HPF	20	20	20	20
24 HPF	20	18	19	16
48 HPF	18	17	15	13
96 HPF	16	14	12	9
144 HPF	13	11	9	7

M. fortuitum group is responsible for 60-80% of post surgical infections caused by mycobacteria. The successful treatment outcome of *M. fortuitum* infections often limited by the multi drug resistance, need of combination therapy, prolonged course, etc. (Cynamon et al., 2012). Santos et al (2016) analysed the resistant profile of *M. fortuitum* isolates and showed their resistance to different classes of antibiotics. Antimicrobial peptides i.e., bacteriocins offers a solution to combat antibiotic resistance of various pathogenic microorganisms. Numerous studies have proved the efficacy of bacteriocins against various drug resistant pathogens (Regmi et al., 2017).

Figure 3: Morphological changes in zebrafish embryos after treating with BLN48



In the present study, the partially purified bacteriocin from *Enterococcus italicus* BLN48 has showed significant inhibition against *M. fortuitum*. In the various studies, numerous antibiotics and chemical compounds have been screened against *M. fortuitum* but there are very less reports on the screening of natural compounds of microbial source against *M. fortuitum* (Gay et al., 1984; Welch et al., 1979; Bagchi et al., 2007). While there are many other reports focuses on the screening of bacteriocins against other mycobacterium species like *M. tuberculosis*. A study by Sosunov et al (2007) has assessed the antimycobacterial of five bacteriocins against *M. tuberculosis* strains. In 2010, Carroll et al., compared the activities of two bacteriocins Lacticin 3147 and nisin against NTM species like *M. kansasii*, *M. avium* paratuberculosis.

Their study found that lacticin 3147 showed superior activity than nisin. A study by Aguilar-Pérez et al (2018) have found the inhibitory activity of bacteriocin AS-48 against *M. fortuitum* and found their MIC as 64µg/ml along with other mycobacterium species. They also found that there is no cytotoxicity obtained against various macrophage cell lines. Our result showed that the *E. italicus* BLN48 exhibits cytotoxicity on vero cell lines in dose dependent manner. However, the in vivo cytotoxicity assay using zebrafish showed that there is no side effect on the morphology of viable larvae in the presence of *E. italicus* BLN48 at three different concentrations. A survey by Fortina et al (2008) has described the safety and biotechnological properties of *E. italicus* of dairy origin. They suggested that *E.*

italicus presence in the cheese lowers the health risk and supports their applications in dairy industry. This study describes the safety profile of *E. italicus* for their wide application in future.

CONCLUSION

Mycobacterium fortuitum is one of the clinically significant rapidly growing mycobacteria which cause pulmonary, skin and soft tissue infections in immunocompromised and immunosuppressed patients. Partially purified bacteriocin of *Enterococcus italicus* BLN48 showed significant inhibitory activity against *M. fortuitum*. Their cytotoxicity assay under both in vitro and in vivo conditions shows that *E. italicus* BLN48 can be developed as potential candidate against *M. fortuitum*. Further analysis on their Minimum Inhibitory Concentration (MIC), purification and their characterization and screening them against other mycobacterial species will leads to the efficient use of *E. italicus* BLN48 for therapeutic purpose.

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