

In vitro Phytochemical Profiling of Anti Diabetic and Anti Inflammatory Activities of *Premna rajendranii* – An Endemic Species

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ABSTRACT

The present study aims to find out the presence of phytochemicals and evaluate the anti diabetic and anti-inflammatory activities of *Premna rajendranii*. Anti diabetic assay was evaluated using α - amylase inhibitory assay by DNSA method, anti-inflammatory activity was tested using the method of Mizushima et al with simple modifications and the methods of Peach and Tracey (1956), Gibbs (1974), Harborne (1984), Trease and Evans (1985), Edeoga et al.,(2005), Khandewal (2008), Kokate et al.,(2001), Sofowara (2009) and Tiwari et al.,(2011) were used to identify the nature of phytochemical constituents present in *Premna rajendranii*. Anti diabetic analysis of methanolic leaf extract of *Premna rajendranii* showed optimal activity when compared to standard drug. Maximum inhibition showed at 100 μ g (79.3%) and minimum inhibition showed at 25 μ g (44.82%). Anti-inflammatory activity of methanolic leaf extract of *P.rajendranii* has shown high inhibition than standard at 50 μ l i.e. 33.88 \pm 0.01. Phytochemical screening shows the presence of Alkaloids, Carbohydrates, Flavonoids, Steroids, Glycosides, Phenols, Proteins, Tannins, Saponins, and Terpenoids in the leaf of *Premna rajendranii*. Anti diabetic and anti-inflammatory analysis showed the presence of bioactive compounds and the medicinal values of *Premna rajendranii*.

KEY WORDS: *PREMNA RAJENDRANII*, *ANTI DIABETIC*, *ANTI-INFLAMMATORY*, *PHYTOCHEMICAL ANALYSIS*.

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INTRODUCTION

Plants have an important role in the day to day life of mankind. Each and every plant contains phytochemicals either with medicinal properties or poisonous properties. Scientists and researchers are more focused on medicinal plants for the benefit of mankind. According to them each of the medicinal plant is a source of drug candidate. From ancient times India is considered a rich repository of medicinal plants and Indian traditional systems of medicine such as Ayurveda, Siddha and folk medicine are well known. Genus *Premna* has an indispensable value in Indian health care practice. Species of *Premna* are known for the preparation of famous Ayurvedic formulation Dashamoolaarishtam (Joshi *et al.*, 2017). *Premna*'s medicinal properties have been used in Indian traditional system of medicine especially for diarrhoea, stomach and hepatic disorders. The various biological activities including antioxidant, antibacterial, anti-inflammatory, cytotoxic and hepatoprotective have been displayed both at extract and pure compound level (Rekha, 2015). During the taxonomic revision of Indian Verbenaceae, Rajendran and Daniel (2002) recognized 31 species and 6 varieties of *Premna*. Recently, PrabhuKumar *et al.*

(2013) reported the discovery of a new species *Premna rajendranii* from Western Ghats (Chinnar of Kerala and Madukkarai of Tamilnadu). Apart from this, a research team comprising Robi, Augustin, Sasidharan and Udayan (2013) rediscovered an endemic and rare species *Premna paucinervis* (C. B. Clarke) Gamble from the Vagamon hills along South Western Ghats of Kerala after a lapse of 140 years of its original type collection by R.H. Beddome (1872) from the Anamalayas in the Western Ghats (Tamilnadu) (Bose, 2014). *Premna* included earlier in the family Verbenaceae, was recently transferred to the Lamiaceae family based on molecular data (A.P.G. IV. 2009; Francis *et al.*, 2019).

MATERIALS AND METHODS

Collection and authentication: The plant material for our investigation was collected from the scrub jungles of the Madukkarai Hills, Coimbatore

District in Tamilnadu, and authenticated by Dr. S. John Britto S.J, at the Rapinat Herbarium and Centre for Molecular Systematics, St. Joseph's College (Autonomous), Tiruchirappalli. The voucher Specimen (RHT 68887) was deposited in the Rapinat Herbarium.

Extraction: The leaves were shade dried and powdered using mechanical grinder. The powder sample was stored in an air tight container and the portion of the powder was taken in test tube and solvent (Methanol) was added to it such that plant powder soaked in it and shaken well. The solution then filtered with the help of muslin cloth and filtered extract was taken and used for antidiabetic and anti-inflammatory analysis.

Preliminary Phytochemical Analysis: Test's for Alkaloids: Mayer's Test: To a few ml of filtrate, one or two drops of Mayer's reagent were added by the side of the test tube. A white creamy precipitate indicated the test as positive. Wagner's Test: To a few ml of filtrate, few drops of Wagner's reagent were added by the side of the test tube. A reddish brown precipitate confirmed the test as positive. Hager's Test: To a few ml of filtrate 1 or 2 ml of Hager's reagent (Saturated aqueous Solution of picric acid) was added. A prominent yellow precipitate indicated the test as positive. (Kokate *et. al.*, 2001).

Test for flavonoids: Pews Test: To 2-3ml extract, was added zinc powder in a test tube, followed by drop wise addition of conc. HCL. Formation of purple red or cherry colour indicated the presence of flavonoids (Peach *et.al.*, 1956). Lead acetate test: 1ml extract was treated with 1ml 10% lead acetate (Pb(OAc)₄) solution. Formation of yellow colour precipitate indicated the presence of flavonoids. Alkaline reagent test: To 2ml test solution, sodium hydroxide solution was added to give a yellow or red colour (Khandewal *et.al.* 2008). Conc.H₂SO₄ test: 5ml of dilute ammonia solution was added to the extract followed by conc.H₂SO₄. Yellow colour indicated the presence of flavonoids. Tests for Phenolic Compounds and Tannins: Ferric Chloride Test: The extract (50 mg) was dissolved in 5 ml of distilled water. To this few drops of neutral 5% Ferric Chloride solution was added. A dark green colour indicated the presence of phenolic compounds.

Potassium dichromate test: To the extract add 5% potassium dichromate solution. Positive result was confirmed by a formation of brown precipitate (for phenol).

Lead Acetate Test: The extract (50 mg) was dissolved in distilled water and to this 3 ml of 10% Lead Acetate solution was added. A bulky white precipitate indicates the presence of phenolic compounds (Treare *et.al.*, 1985). Braymer's Test: To 2 ml extract, added 2 ml H₂O and followed with 2-3 drops of FeCl₃ (5%). Green precipitate proved presence of tannins. Tests for Saponins: Foam Test: To 1ml of extract, add 2ml of distilled water and shaken vigorously and allowed to stand for 10 min. There is the development of foam on the surface of the mixture. Then shake for 10 minutes, it indicates the presence of saponins (Khandewal *et.al.*, 2008). NaHCO₃ Test: To extract a drop of sodium bicarbonate was added. The mixture was shaken vigorously and kept for 3 min. A honey comb like froth was formed and

it showed the presence of saponins. Tests for Glycosides: Keller Kiliani Test (Test for cardiac glycoside): To 2 ml extract, was added 1 ml glacial acetic acid, one drop 5% FeCl₃ and 1 ml conc. H₂SO₄. A brown ring of the interface indicated the presence of cardiac glycosides (Kokate *et.al.*, 2001; Khandewal *et.al.*, 2008). Glycoside Test: To small amount of extract, was added 1 ml water and shake well. Then aqueous solution of NaOH was added. Yellow color appeared that indicated the presence of glycosides (Treare *et.al.*, 1985).

Molisch's Test: To 1ml of extract, 2drops of Molisch's reagent was added in a test tube and 2ml of con. H₂SO₄ was added carefully keeping the test tube slightly curved. Formation of violet ring at the junction indicated the presence of glycosides (Khandewal *et.al.*, 2008).

Tests for Carbohydrates: Molish's Test: To 2 ml of filtrate two drops of alcoholic solution of α - naphthol was added, the mixture was shaken

Table: 1.1 Preliminary phytochemical analyses in *Premna rajendranii* (leaf)

	Test	Chloroform	Acetone	Ethanol	Methanol	Aqueous
Alkaloids	Wager's	+	+	++	+++	++
	Hager's	-	+	++	+++	++
	Mayer's	-	+	++	+++	+
Flavonoids	Pew's	+	+	+	++	-
	Lead Acetate	+	+++	+++	+++	+++
	NaOH	-	+++	+++	+++	+++
	Con.H ₂ SO ₄	+	+++	+++	+++	++
Phenol & tannin	FeCl ₃	+	+	+	++	+
	K ₂ Cr ₂ O ₇	+	+	+	+	+
	Lead Acetate	-	-	+	++	+
	Braymers	+	-	+	+	+
Saponins	Foam	-	-	-	-	-
	NaHCO ₃	-	-	-	-	-
Glycosides	Keller kiiani	-	+++	+++	+++	+++
	Glycosides	-	+++	+++	+++	+++
	Molish	-	++	+++	+++	++
Carbohydrates	Molish	-	+	++	++	++
	Benedicts	-	-	+	+	-
Terpenoids	Salkowskis	-	+	+	++	+
Quinones	Quinones	-	-	-	++	+++
Sterols	Salkowskis	-	-	-	-	-
	Keller kiiani	-	+	+	++	++
Protein	Biuret	-	+	+	+	+

well and 1 ml of con. H₂SO₄ was added slowly along the sides of the test tube and allowed to stand. A violet ring indicated the presence of carbohydrates. Benedict's test: To 0.5 ml of filtrate, 1 ml of Benedict's reagent was added. The mixture was heated on a boiling water bath for 2 mins. A characteristic coloured precipitate indicated the presence of sugar. Test for Terpenoids: Salkowski's Test: 2 ml of chloroform and 1 ml of conc. H₂SO₄ was added to 1 ml of extract and observed for reddish brown color that indicated the presence of terpenoids. Tests for quinones: 1ml of extract was treated with alcoholic potassium hydroxide solution. Quinines give coloration ranging from red to blue. Test for sterols: Salkowski's Test: To 2 ml of extract, was added 2 ml chloroform and 2 ml conc. H₂SO₄ from the side of the test tube. Chloroform layer appeared red and acid layer showed greenish yellow fluorescence indicated the presence of sterols (Khandewal et.al., 2008).

Tests for Proteins and Amino Acids: Biuret Test: An aliquot of 2 ml of filtrate was heated with 1 drop of 2 % H₂SO₄ solution. To this 1 ml of ethanol (95%) was added, followed by excess of KOH Pellets. Pink colour in the ethanolic layers indicated the presence of proteins. Conc. H₂SO₄ Test: 2 ml extract was treated with few drops of conc. H₂SO₄. Formation of white precipitate indicated the presence of proteins.

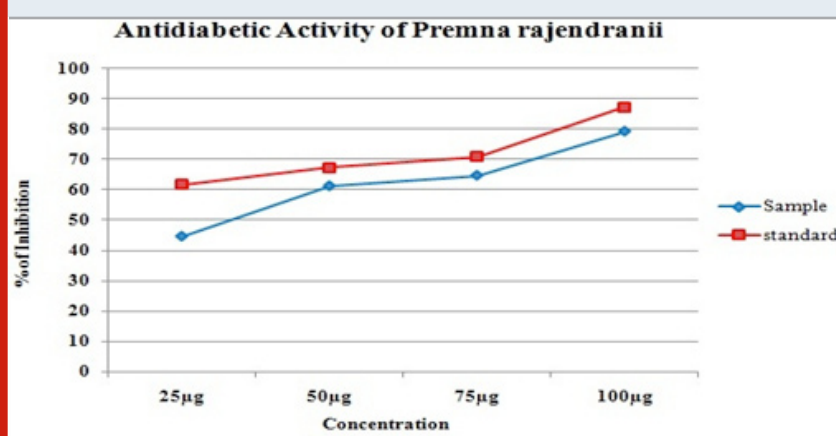
Xantho proteins Test: 2 ml extract was treated with few drops of conc. HNO₃ and NH₃ solution. Formation of reddish orange precipitate indicated the presence of xantho proteins.

Anti-Diabetic Assay (A-Amylase Inhibitory Assay) By Dnsa Method: α -amylase was dissolved in phosphate buffer saline (PBS, 0.02 mol/L, pH6.8) at a concentration of 0.1 mg/ml. Various concentrations of sample solutions (0.25 ml) were mixed with α -amylase solution (0.25 mL) and incubated at 37 °C for 5 min. Then the reaction was initiated by adding 0.5 mL 1.0% (w/v) starch substrate solution to the incubation medium. After incubation at 37 °C for 3 min, the reaction was stopped by adding 0.5 ml DNS reagent (1% Dinitrosalicylic acid, 0.05% Na₂SO₃ and 1% NaOH solution) to the reaction mixture and boiling at 100 °C for 5 min. After cooling to room temperature, the absorbance (Abs) at 540 nm was recorded by a spectrophotometer. The inhibition percentage was calculated by the following equation:

Table 2. Antidiabetic Activity of Methanolic leaf extract of *Premna rajendranii*

SL.NO:	Concentration in (μ g)	% of inhibition	
		Sample	Standard
1	25	44.82 \pm 0.5	61.7 \pm 0.01
2	50	61.37 \pm 0.6	67.3 \pm 0.01
3	75	64.82 \pm 0.4	70.9 \pm 0.01
4	100	79.3 \pm 0.7	87.4 \pm 0.21
IC ₅₀ value	288.8	246.7	

Figure 1. Antidiabetic Activity of *Premna rajendranii*



$$\text{Percentage Inhibition} = \frac{[(\text{Abs1} - \text{Abs2})/\text{Abs1}] \times 100}{\times 100}$$

Where, Abs1= control and Abs2= sample.

Anti-Inflammatory Activity: Method of Mizushima et al was followed with simple modifications. The reaction mixture (0.5 ml) consisted of 0.45 ml bovine serum albumin (3% aqueous solution) and varying concentration of compound (25, 50, 100,200µg/ml of final volume), pH was adjusted to 6.3 using small amount of 1N hydrochloric acid. The samples were incubated at 37°C for 20 min and then heated at 80°C for 2min. After cooling the samples, 2.5 ml phosphate buffer saline (pH 6.3) was added to each tube. The absorbance was measured using spectrophotometer at 660nm. The percentage inhibition of protein denaturation was calculated as follows:

$$\text{Percentage of inhibition} = \frac{[(\text{Abs Control} - \text{Abs Sample}) / \text{Abs control}] \times 100}{\times 100}$$

RESULTS AND DISCUSSION

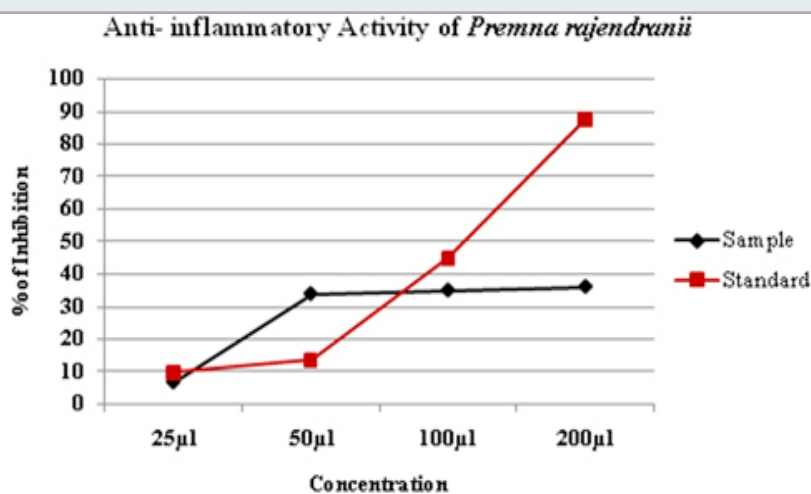
The result of phytochemical profiling of leaves of *P.rajendranii* indicates the presence of Alkaloids, Flavonoids, Phenols, Tannins, Glycosides, Carbohydrates, Terpenoids, Quinones, Sterols and Proteins. High concentrations of bioactive compounds were found in methanolic, ethanolic, aqueous, acetone extracts while very low concentrations in chloroform extract. Alkaloids, Flavonoids and Glycosides were mainly seen in most of the samples except chloroform extraction (Table 1).

Diabetes mellitus is one of the major metabolic disorders (Keerthana et al., 2013), and is characterized by high blood glucose levels. According to World Health Organization, 180 million people are currently suffering from DM and the rate is rapidly growing. There are many effective medicines for DM but with serious side effects. Recently researchers are in the experiments to find effective medicine for DM without any side effect. More than 1200 plants have a potential antidiabetic capacity (Modak et. al., 2007; Bailes, 2002; Chitturi and George, 2002; Kesari et. al., 2007,Prabhu et al 2018). Methanolic leaf extract of *P.rajendranii* was investigated for in vitro antidiabetic activity with respect to

Table 3. Anti-inflammatory Activity of *Premna rajendranii*

SL.NO:	Concentration in (µg)	% of inhibition	
		Sample	Standard
1	25	6.88±0.02	9.79±0.01
2	50	33.88±0.01	13.54±0.02
3	100	34.98±0.03	44.86±0.05
4	200	36.08±0.1	87.78±0.01

Figure 2. Anti-inflammatory Activity of *Premna rajendranii*



inhibition of α -amylase. The concentrations of samples used for testing the inhibitory activity are 25, 50, 75 and 100 (μ g). The inhibition activity of test samples showed optimum activity when compared to commercial drug at different concentrations. The maximum inhibition showed at 100 μ g concentration was 79.3% and minimum inhibition showed at 25 μ g concentration was 44.82%. IC50 value of sample (288.8) is higher than standard drug (246.7). Acarbose was used as standard drug (Table 2, Fig: 1). Protein denaturation is the major cause of Inflammation (Mizushima and Kobayashi, 1968) and it is a response to negative stimuli including injury, infection (Lumeng and Saltiel, 2011). Basically it was meant to destroy invading microorganisms, inactivate toxins, repairing and healing of injuries which might lead to life threatening hypersensitivity reactions. Inflammation is the major cause for several diseases including neurological, cardiovascular, intestinal, dental and renal disorders and also linked to diabetics, ageing, obesity, multiple sclerosis, pancreatitis, cancer (Sakat *et al.*, 2010; Burns *et al.*, 2001; Kuek *et al.*, 2006; Grivennikov *et al.*, 2010; Jenny, 2012; Hoque *et al.*, 2012; Marchant *et al.*, Wyss- Coray *et al.*, 2012).

Prescribed medicines for inflammation are mainly non steroidal anti- inflammatory drugs. These drugs have certain side effects causing gastric bleeding, ulceration, renal failure. Medicinal plants have a prolonged history of use as a remedy for inflammation (Insel, 1996; Rang *et al.*, 2007). *Premna serratifolia* and *Premna tomentosa* have been reported to possess anti-inflammatory activity (Alam *et al.*, 1993; Habtemariam *et al.*, 2015). Methanolic leaf extract of *P.rajendranii* has shown high inhibition than standard at 50 μ l i.e. 33.88 \pm 0.01. The inhibition activity of other test samples (25 μ l, 100 μ l, 200 μ l) showed moderate activity when compared to standard drug. Diclofenac was used as the standard drug.

CONCLUSION

Our study demonstrates that the *P.rajendranii* acts as an antidiabetic and anti-inflammatory agent. The antidiabetic and anti-inflammatory activities may be due to the presence of various

bioactive compounds especially flavonoids. More purification needs to be done and further research on *P.rajendranii* is necessary for isolating bioactive compounds and their mode of action.

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