

Susceptibility status of dengue fever vector *Aedes aegypti*, (L.) in Republic of Yemen

Sadeq K. N. Alhag

Faculty of Sciences, Biology Department, Ibb University, Yemen

ABSTRACT

The aim of the present research work was to determine the current susceptibility level of larvae and adults of a field strain dengue fever vector *Aedes aegypti* (L.) to some conventional insecticides. Taking the values of LC50 (concentration required to kill 50 % of mosquito larvae), the results showed that the mosquito larvae of *A. aegypti* were more susceptible to the OP insecticides sumithion (0.007 ppm) than acifon (0.019 ppm), actillic (0.049 ppm) and of onac (1.87 ppm) respectively, while the pyrethroids fendona (0.09 ppm) was more effective against larvae than aralin (1.24 ppm) by about 13.8 times. On the other hand, the findings revealed that the exposure of mosquito adult females to the diagnostic dosages of deltamethrin(0.05%), lambdacyhalothrin(0.5%), permethrin(0.75%), cyfluthrin(0.15%), malathion(5%) and fenitrothion (1%) caused 81, 75, 90, 84, 64 and 29% mortality, respectively. According to WHO criteria, the data indicated that adult mosquitoes of the field strain *A. aegypti* were resistant to the insecticides lambdacyhalothrin, malathion and fenitrothion but were tolerant to deltamethrin, permethrin and cyfluthrin.

KEY WORDS: DENGUE FEVER, CONVENTIONAL MOSQUITOCIDES, AEDES AEGYPTI, INSECTICIDES RESISTANCE

INTRODUCTION

Dengue fever is an acute virus disease of the tropic and sub-tropic regions around the world, especially in urban and semi-urban areas (Halsted *et al.*, 2001). It is a mosquito-borne viral illness that is caused by one of the four serotypes of dengue virus, belonging to the family Flaviviridae and predominantly transmitted by *Aedes aegypti* and few other *Aedes* species. It has emerged as a major international health problem with an expanded geographic distribution and potential to cause major epidemics (Gubler, 2002; Fakeeh and Zaki, 2003; Rodriguez *et al.*, 2002, Fansiri *et al.*, 2006; WHO, 2010). The present study was conducted to determine the current susceptibility status of

mosquito larvae and adults of *A. aegypti* the primary vector of dengue fever to some conventional insecticides commonly used in mosquito control.

MATERIALS AND METHODS

MOSQUITO STRAIN

Tests were performed on a field strain of *A. aegypti* (L.) raised from wild larvae, collected from different localities of Ibb governorate, Yemen. The larvae were reared until pupation and adult emergence took place for maintaining the stock colony.

ARTICLE INFORMATION:

*Corresponding Author

Received 10th October, 2013

Accepted after revision 15th December, 2013

BBRC Print ISSN: 0974-6455

Online ISSN: 2321-4007

© A Society of Science and Nature Publication, 2013. All rights reserved.

Online Contents Available at: <http://www.bbrc.in/>

This strain was maintained at a room temperature of $27 \pm 1^\circ\text{C}$ and $70 \pm 5\%$ R. H. With a 14 : 10 L : D photoperiod throughout this study.

MOSQUITO LARVICIDES TESTED

The following insecticides were used:

- The synthetic pyrethroid Fendona (Alphacypermethrin 6%).
- The synthetic pyrethroid Aralin (Tetramethrin 2% + cypermethrin 11%).
- The organophosphates Ofonac (Pyridafenthion 40%).
- The organophosphates Acifon (Trichlorfon 50%).
- The organophosphates Actillic (Pirimiphos methyl 50%).
- The organophosphates Sumithion (Tetramethrin 2.5% + Phthothion 25%).

LARVAL BIOASSAY

Susceptibility tests of *A. aegypti* larvae were conducted according to the procedure recommended by WHO (1981). Early fourth instar larvae were exposed to serial concentrations of the tested insecticides for 24 hours, in groups of glass beakers containing 100 ml of tap water. Five replicates of 20 larvae each per concentration, and so for control trials were set up. Larval mortalities were scored at 24 hr post-treatment (WHO, 2005).

Data were subjected to probit analysis, for calculating LC50 and LC95 values using the Finney (1972) method and GW BASIC probit1 Statistical software.

MOSQUITO ADULTICIDES TESTED

The insecticides used in the present study were diagnostic doses of WHO impregnated papers kindly supplied by Center of WHO, School of Biological Sciences, Universiti Sains Malaysia, 11800 Penang. The tested adulticide were the pyrethroids i.e., Deltamethrin (0.05%), Lambdacyhalothrin (0.5%), Permethrin (0.75%), Cyfluthrin (0.15%) and organophosphorus i.e., Malathion (5.0%) and Fenitrothion (1.0%).

ADULT BIOASSAY

The adult bioassay was conducted using WHO test kits (WHO, 1981). Non blood fed mosquito females aged 3-5 days old were exposed for 1hr to the tested insecticides. For each insecticide, four replicates were used, each containing 25 mosquito females.

Mosquitoes used as controls were exposed to papers without insecticides. After the exposure period, the mosquitoes were transferred to clean holding tubes and provided with cotton pads soaked with 10% sugar solution. Mortality counts were recorded 24 hr after the exposure period and compared with control trials. The resistance status was determined according to WHO criteria: population is considered to be susceptible to the test adulticide if the percent mortality is equal or greater than 98%, a resistant population is the one which shows less than 80% mortality (WHO, 1981).

RESULTS AND DISCUSSION

Susceptibility levels of *A. aegypti* mosquito larvae following treatment with different concentrations of the pyrethroid insecticides fendona and aralin as well as the organophosphates insecticide ofonac, acifon, actilic and sumithion are shown in Table 1. The effective concentrations of the above compounds against 4th larval instars ranged from 0.06 - 0.20 ppm; 0.50-4.00 ppm; 1.5-4.0 ppm; 0.01-0.1 ppm; 0.02-0.2 ppm and 0.005 - 0.05 ppm, respectively. The corresponding larval mortalities for these compounds were 17-93%; 23-87%; 21-93%; 27-97%; 18-96% and 22-93% respectively. LC50 values at 24 hours showed that the sumithion (0.007 ppm) proved to be the most effective compound, followed by the acifon (0.019 ppm), actilic (0.049 ppm), fendona (0.09), aralin (1.24 ppm) while the ofonac (1.87 ppm) was the least effective.

In other words, the results indicate that mosquito larvae of *A. aegypti* were more susceptible to the organophosphates sumithion than ofonac, actilic and acifon by about 267.14; 7 and 2.71 folds, respectively, while the pyrethroid fendona was more efficient than aralin against *A. aegypti* by about 13.8 times. However, it can be concluded that the change in the susceptibility levels of the present mosquito larvae is possibly due to the differential mode of action of the test compounds and its effective concentrations. The fluctuations in the percentage mortalities obtained for the different concentrations of different compounds tested against the present mosquito larvae support this conclusion (Saleh and Aly, 1987; Canyon and Hii, 1999; Nazniet *et al.*, 2005; Coleman *et al.*, 2006; Vittum *et al.*, 2008).

Data of table 2 show results of susceptibility status of mosquito adults of a field strain of *A. aegypti* against four pyrethroids (deltamethrin, Lambdacyhalothrin, permethrin and cyfluthrin) and two organophosphate adulticides (malathion and fenitrothion), at diagnostic dosages using WHO filter impregnated paper assays. The records showed that the exposure of mosquito females of *A. aegypti* to the diagnostic dosages of deltamethrin, lambdacyhalothrin, permethrin, cyfluthrin malathion and fenitrothion caused 81, 75, 90, 84, 64 and 29% mortality, respectively. Taking WHO criteria into consideration, the results indicate that *A. aegypti* mosquito females were resistant to lambdacyhalothrin, malathion and fenitrothion but were tolerant to deltamethrin, permethrin and cyfluthrin.

In Jeddah, the susceptibility status of mosquito adults of field strain *Culex pipiens* against some insecticides have been determined. The records showed that exposure of adult females to the diagnostic dosages of lambdacyhalothrin, permethrin, malathion and fenitrothion caused 54, 57, 56 and 23% mortality respectively, indicating resistance of the present mosquito species to the above insecticides (Mahyoub, 2011).

In different parts of the world, laboratory and field trials in this respect have been carried out by several workers (Huong and Nguyen, 2000; Hemingway and Ranson, 2000; Rodriguez *et al.*, 2002; Huber *et al.*, 2003; Nazni *et al.*, 2005). In Port Sudan City - Red Sea State, *Aedes aegypti* were found to be susceptible to Deltamethrin 0.05%, Bendiocarb 0.1%, tolerant to Lambdacyhalothrin 0.05% and resistant to DDT 4% and Malathion 5% respectively (Husham *et al.*, 2010).

In India, Katyal *et al.*, (2001) used the diagnostic dosage of different insecticides against mosquito adults of *A. aegypti*. They found that the test mosquitoes were resistant to DDT (74% mortality) and dieldrin (46% mortality) but were tolerant to fenitrothion (91% mortality) and were susceptible to lambda-cyhalothrin (100% mortality). Liu *et al.*, (2004) detected high resistance values to some pyrethroid insecticides against *Culex* of unique *fasciatus* in Alabama and Florida. In Thailand, the field strain of *A. aegypti* was found to be susceptible to fenitrothion, resistant to permethrin and highly resistant to DDT (Paeporn *et al.*, 2005).

In general, it can be concluded that resistance status to insecticides within mosquito populations is a dynamic process

depending on the type of insecticide used, the frequency of use and the possible effect of environmental pollution following pest control measures (Orshan *et al.*, 2005).

On the other hand, variations in susceptibility are so great that susceptibility investigations should assess mosquito samples collected from many sites with an area, rather than relying on the results obtained from one area (Conyon and Hii, 1999). However, consideration must be taken regularly to monitor the susceptibility status of local mosquitoes to insecticides used in control programmes. Such records on insecticide susceptibility are essential in defining future control strategies against mosquito vectors.

TABLE 1: Susceptibility levels of a field strain of *A. aegypti* larvae to the insecticides fendona, aralin, ofonac, acifon, actillic and sumithion.

Larvicides	Effective Concentration (Ppm)	Larval Mortality (%)	Lc ₅₀ (LC ₉₅)	Slope
Fendona	0.06-0.20	17-93	0.09 (0.17)	3.1
Aralin	0.50-4.00	23-87	1.24 (11.6)	1.76
Ofonac	1.50-4.00	21-93	1.87 (1.87)	3.9
Acifon	0.01-0.10	27-97	0.019 (0.095)	2.5
Actillic	0.02-2.20	18-96	0.049 (0.19)	2.8
Sumithion	0.005-0.05	22-93	0.007 (0.073)	1.8

REFERENCES

- Abbott, W.S. (1925). A method of computing the effectiveness of an insecticide. *J. Econ. Entomol.* 18:256-269.
- Canyon, D.V. and J.K.L.Hii (1999). Insecticide susceptibility status of *Aedes aegypti* (Diptera: Culicidae) from Townsville. *Aust. J. Entomol.* 38:40-43.
- Coleman M., B. Sharp I. Seocharan and Hemingway J. (2006). Developing an evidence-based decision support system for rational insecticide choice in the control of African malaria vectors. *J. Med Entomol* 43(4):663-668.
- Fakeeh, M. and A. M. Zaki (2003). Dengue in Jeddah, Saudi Arabia, 1994-2002. *Dengue Bulletin* (27) 13:18.
- Fansiri T., U. Thavara, A. Tawatsin, S. Krasaesub and R. Sithiprasasna (2006). Laboratory and semifield evaluation of mosquito Dunks against *Aedes aegypti* and *Aedes albopictus* larvae (Diptera: culicidae). *Southeast Asian J. Trop. Med. Pulbl. Health.* 37(1):62-66.
- Finney, D. J. (1972). *Probit Analysis*, 3rd edition. Griffin, Cambridge University Press, Cambridge.
- Gubler, D. (2002). Epidemic dengue/dengue hemorrhagic fever as a public health, social and economic problem in the 21st century. *Trends in Micro* 10:100-103.
- Halstead, S. B., T. G. Streit, J. G. Lafontant, R. Putvatana, K. Russell, W. Sun, N. Kanesa-Thanan, C. G Hayes and D. M. Watts (2001). Haiti: absence of dengue hemorrhagic fever despite hyperendemic dengue virus transmission. *Am. J. Trop. Med. Hyg.* 65(3):180-183.
- Hemingway, J. and H. Ranson (2000). Insecticide resistance in insect vectors of human disease. *Annu. Rev. Entomol.* 45:371-391.
- Huber K., L. L. Loan, T. H. Hoang, T. K. T., F. Rodhain and Anna-Bella F. (2003). *Aedes aegypti* in South Vietnam: Ecology genetic structure, vectorial competence and resistance to insecticides. Vol 34 No. 1.
- Huong V. D. and Nguyen T. B. N. (2000). Susceptibility of *Aedes aegypti* to insecticides in South Vietnam. *Dengue Bulletin* Volume 23. Hanoi, Vietnam. 11. Sh Salari Lak.
- Husham A. O., M. A. Abdalmagid and Brair M. (2010). Status Susceptibility of dengue vector; *Aedes aegypti* to different groups of Insecticides in Port Sudan City - Red Sea State. *Sudanese J. of Public health*, vol(5):4.
- Katyal R., P. Tewari, S. J. Rahman, H. R. Pajni, K. Kumar and K. S. Gill (2001). Susceptibility status of immature and adult stages of *Aedes aegypti* against conventional insecticides in Delhi, India. *Dengue Bull.* 25:84-87.
- Liu H.; C. Eddie; A. Guo and N. Liu (2004). Insecticide resistance in Alabama and Florida mosquito strains of *Aedes albopictus*. *J. Med. Ent.* 41(5):946-952
- Mahyoub, J.A. (2011). Study of the Seasonal Activity and Dynamic Fluctuation of Medically Important Species of Mosquitoes with Reference to Testing the Susceptibility of the Dominant Species *Culex pipiens* L. 1758 to some Insecticides in Jeddah. Ph.D. Thesis. Faculty of Sciences, King Abdulaziz University, Jeddah-Saudi Arabia.
- Nazni W. A., H. L. Lee and A. H. Azahari (2005). Adult and larval insecticide susceptibility status of *Culex quinquefasciatus* (say) mosquitoes in Kuala Lumpur Malaysia. *Tropical Biomedicine* 22(1):63-68.
- Orshan L., Kelbert, M. and Perner, H. (2005). Patterns of insecticide resistance in Larval *Culex pipiens* Populations in Israel: Dynamics and trends. *J. Vector Ecol.* 30:289-294.

Paeporn P., P. S. Kasin, S. Sathantriphop and S. Sangkitporn (2005). Insecticides Susceptibility of *Aedes aegypti* in Tsunami-affected areas in Thailand. *Dengue Bull.* 29:210-213.

Rodriguez, M.M.; J. Bisset; M. Ruiz, and A. Soca (2002). Cross-resistance to pyrethroids and organophosphorus insecticides induced by selection with temephos in *Aedes aegypti* (Diptera: Culicidae) from Cuba. *J. Entomol.* 39:882-888.

Saleh, M.S. and M.I. Aly (1987). The biological effects of three insect growth regulators on *Culex pipiens* L. *Anz. Schadlingskunde, pflanzenschutz, Umweltschutz*, 60:34-37.

Vittum P. and Silcox C. A. (2008). Development of resistance by turf insects to insecticides. *GCM* 100-106. World Health Organization (1981). Instruction for determining the susceptibility of adult mosquito

to organochlorine, organophosphate and carbamate insecticides establishment of base line. Geneva: WHO/VBC/81.806.PP1-7.

World Health Organization (2005). Prevention and control of dengue and dengue hemorrhagic fever. WHO, Regional Publication, seral No. 29. 134 pgs.

World Health Organization (2010). First WHO report on neglected tropical diseases: working to overcome the global impact of neglected tropical diseases. WHO/HTM/NTD/1.

Zahirnia A. H., H. Vatandoost M., Nateghpour and E. Djavadian (2002). Insecticide Resistance/Susceptibility Monitoring in *Anopheles pulcherrimus* (Diptera: Culicidae) in Ghasreghand District, Sistan and Baluchistan Province, Iran. *Iranian J. Publ. Health*, 31:11-14.

BBRC